Package 'tilingArray'

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Title Transcript mapping with high-density oligonucleotide tiling arrays

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Imports strucchange, affy, vsn, genefilter, RColorBrewer, grid, stats4

Description The package provides functionality that can be useful for the analysis of high-density tiling microarray data (such as from Affymetrix genechips) for measuring transcript abundance and architecture. The main functionalities of the package are: 1. the class 'segmentation' for representing partitionings of a linear series of data; 2. the function 'segment' for fitting piecewise constant models using a dynamic programming algorithm that is both fast and exact; 3. the function 'confint' for calculating confidence intervals using the strucchange package; 4. the function 'plotAlongChrom' for generating pretty plots; 5. the function 'normalizeByReference' for probe-sequence dependent response adjustment from a (set of) reference hybridizations.

Reference Huber W, Toedling J, Steinmetz, L. Transcript mapping with high-density oligonucleotide tiling arrays. Bioinformatics 22, 1963-1970 (2006).

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Description

tilingArray package overview

Details

The package provides some functionalities that can be useful for the analysis of high-density tiling microarray data (such as Affymetrix genechips) for measuring transcript abundance and architecture. The main functionalities of the package are:

- The segmentation class for representing partitionings of a linear series of data (such as microarray intensity readings along a chromosome strand).
- The function segment for fitting piecewise constant models using a dynamic programming algorithm that is both fast and exact, and confint for calculating confidence intervals using the strucchange package. Please see the vignette *Segmentation demo* in the file inst/doc/segmentation.pdf (source file inst/scripts/segmentation.Rnw).

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• The function plotAlongChrom for generating pretty plots of segmentations along with genomic features. Please also see the vignette Segmentation demo.

• The function normalizeByReference for probe-sequence dependent response adjustment from a (set of) reference hybridizations. Please see the vignette *Assessing signal/noise ratio before and after normalization* in the file inst/doc/assessNorm.pdf (source file inst/scripts/assessNorm.Rnw).

Author(s)

W. Huber <huber@ebi.ac.uk>

breakpointsPretend

Accessor methods for breakpointsPretend objects - not to be called by the user.

Description

Accessor methods for breakpointsPretend objects - not to be called by the user.

These functions are used in the interface between the segmentation class and the confint.breakpointsfull method of the strucchange package. This method calls breakpoints and residuals methods for its first argument, and since we pass an argument of S3 class breakpointsPretend, we can avoid the overhead of the corresponding methods for breakpointsfull objects. These functions are of no interest to the user.

Usage

```
## S3 method for class 'breakpointsPretend'
residuals(object, breaks, ...)
breakpoints.breakpointsPretend(obj, breaks, ...)
```

Arguments

```
object a breakpointsPretend object.
obj a breakpointsPretend object.
breaks dummy argument, is ignored.
... futher arguments.
```

Value

residuals and breakpoints.

Author(s)

```
W. Huber <huber@ebi.ac.uk>
```

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comparisonPlot	Plot a vertical layout of panels for the comparison of different along-chromosome profiles.

Description

This function is used for Figure 5 in the David et al. (PNAS 2006) paper and in the Huber et al. methods paper.

Usage

```
comparisonPlot(x, y, xscale=range(x), yscale, anno, ticks, pch=20, cex=1, bgcol="#f2f2f2")
```

Arguments

X	numeric vector.
у	list of numeric vector, each of same length as x.
xscale	numeric vector of length 2.
yscale	matrix with 2 rows and columns corresponding to the elements of x.
anno	dataframe with columns start, end, and name, each row corresponds to one gene CDS to be plotted at the bottom.
ticks	numeric vector, where to plot the ticks.
pch	A numeric or character vector indicating what sort of plotting symbol to use, see grid.points.
cex	Multiplier applied to fontsize, see gpar.
bgcol	Color to use as background for some of the plot panels.

Value

Function is called for its side-effect.

Author(s)

W. Huber <huber@ebi.ac.uk>

References

•••

Examples

##

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costMatrix

Segmentation cost matrix

Description

This function calculates the cost matrix for the segmentation model

Usage

```
costMatrix(x, maxk)
```

Arguments

Х

Numeric vector of length n or matrix with n rows and d columns, where n is the number of sample points and d the number of replicate measurements (e.g. from multiple arrays).

maxk

Positive integer.

Details

See the package vignette Calculation of the cost matrix.

Value

Matrix with maxk rows and length(x) columns.

Author(s)

W. Huber

Examples

```
d = 4
x = apply(matrix(rnorm(200), ncol=d), 2, cumsum)
maxk = 50

G = costMatrix(x, maxk=maxk)

G.pedestrian = matrix(NA, nrow=nrow(G), ncol=ncol(G))
for(i in 1:(ncol(G)))
  for(k in 1:min(nrow(G), nrow(x)-i+1))
    G.pedestrian[k, i] = (k*d-1)*var(as.vector(x[i:(i+k-1), ]))

stopifnot(identical(is.na(G), is.na(G.pedestrian)))
stopifnot(max(abs(G-G.pedestrian), na.rm=TRUE) <= 1e-6)</pre>
```

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findSegments	Fit a piecewise constant curve to a sequence of numbers – OBSO- LETE, please use function segment instead.
--------------	---

Description

This function is only here for backward compatibility - please use segment.

The function fits a piecewise constant curve to a sequence of numbers using a simple least squares cost function and the dynamic programming algorithm described by Picard et al. (see reference).

Usage

```
findSegments(x, maxcp, maxk, verbose=TRUE)
```

Arguments

x Numeric (real) vector.

maxcp Integer (length 1): maximum number of segments (= 1 + maximum number of

change points).

maxk Integer (length 1): maximum length of a segment.

verbose Logical: if this parameter has a positive value, various diagnostic output is

printed.

Details

The complexity of the algorithm is length(x)*maxk in memory and length(x)*maxk*maxcp in time.

Value

An object of class "segmentation" A list with elements

J likelihood criterion

th matrix of segment start points
dat the data used for the segmentation

call the function call

.

See the vignette, and the paper cited below for details.

Note

This function is depracated and replaced by function segment, but still included for backward compability.

Author(s)

W. Huber <huber@ebi.ac.uk>, Joern Toedling <toedling@ebi.ac.uk>

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References

A statistical approach for CGH microarray data analysis. Franck Picard, Stephane Robin, Marc Lavielle, Christian Vaisse, Gilles Celeux, Jean-Jacques Daudin, Rapport de recherche No. 5139, Mars 2004, Institut National de Recherche en Informatique et en Automatique (INRIA), ISSN 0249-6399. The code of this function is based on the Matlab implementation presented at http://www.inapg.fr/ens_rech/mathinfo/recherche/mathematique/outil.html, but it has evolved.

Examples

```
x = rep(sin((0:4)/2*pi), each=3) + rnorm(3*5, sd=0.1)
res = findSegments(x, maxcp=6, maxk=15)
```

gffSub

Example of a genomic feature object

Description

Example of a genomic feature object

Usage

```
data(gffSub)
```

Details

gffSub is a data frame that contains the features from 35000bp - 50000bp in yeast chromosome one.

Author(s)

Zhenyu Xu <zhenyu@ebi.ac.uk>

Examples

```
data(segnf)
data(gffSub)
nmLabel = colnames(segnf$"1.+"@y)
plotAlongChrom(segnf,chr=1, coord=c(35000,50000),what="heatmap",
    gff=gffSub,rowNamesHeatmap=nmLabel)
```

normalizeByReference Probe-specific normalization of hybridization intensities from an oligonucleotide microarray

Description

Adjust the hybridization intensities from an oligonucleotide microarray for probe-specific response effect by using one or several reference hybridizations. If x contains more than one array, vsnMatrix from the vsn package is called for between array normalization.

Usage

normalizeByReference(x, reference, pm, background, refSig, nrStrata=10, cutoffQuantile=0.05, plotFileNames, verbose=FALSE)

Arguments

X	ExpressionSet containing the data to be normalized.
reference	ExpressionSet with the same number of features as x, containing the reference signal, on the raw scale (non-logarithmic). This argument can be used to directly input the data from a set of replicate DNA hybridizations. Alternatively, the argument refSig can be specified.
pm	Indices specifying the perfect match features in reference (see Details). This can be either an integer vector with values between 1 and nrow(exprs(reference)) or a logical vector.
background	Indices specifying a set of background features in x (see Details). This can be either an integer vector with values between 1 and nrow(exprs(x)) or a logical vector.
refSig	A numeric vector of the same length as pm with estimates of probe response effects, on a logarithm-like scale. This argument can be specified alternatively to reference.
nrStrata	Integer (length 1), number of strata for the estimation of the background function.
cutoffQuantile	Numeric (length 1), the probes whose reference signal is below this quantile are thrown out.
plotFileNames	Character vector whose length is the same as the number of arrays in x. Optional, if missing, no plots are produced.
verbose	Logical of length 1, if TRUE, some messages about progress are printed.

Details

The intensities in x are adjusted according to the reference values. Typically, the reference values are obtained by hybridizing a DNA sample to the array, so that the abundance of target is the same for all reference probes, and their signal can be used to estimate the probe sequence effect. A reference probe is a probe that perfectly matches the target genome exactly once. Usually, not all probes on a chip are reference probes, hence the subset of those that are is specified by the argument pm.

The background signal is estimated from the probes indicated by the argument background. They need to be a strict subset of the reference probes. I.e., they need to uniquely match the target

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organism's DNA, but are not expected to match any of its transcripts. A robust estimation method is used, so a small fraction of background probes that do hit transcripts is not harmful.

A limitation of this normalization method is that it only makes sense for the data from reference probes, NA values are returned for all other probes.

The functions PMindex and BGindex can be used to produce the pm and background arguments from a probeAnno environment such as provided in the davidTiling package.

To summarize, a reference probe (indicated by argument pm) is a probe that perfectly matches the target genome exactly once, a background probe (indicated by argument background) is a reference probe which we expect not to be transcribed. These should not be confused with what is called 'perfect match' and 'mismatch' probes in Affymetrix annotation.

Value

A copy of x with the normalized intensities.

Author(s)

W. Huber <huber@ebi.ac.uk>

References

The method implemented in this function is described in detail in Section 2.3 of the article Huber W, Toedling J, Steinmetz, L. Transcript mapping with high-density oligonucleotide tiling arrays. Bioinformatics 22, 1963-1970 (2006).

See Also

PMindex, BGindex

Examples

see vignette assessNorm.Rnw in inst/scripts directory

otherStrand

Return the name of the opposite strand

Description

Return the name of the opposite strand

Usage

otherStrand(x)

Arguments

Х

Character vector whose elements are "+" or "-".

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Details

This is a rather trivial convenience function.

An alternative would be to code strands with integers -1 and +1, in which case the inversion would be a trivial builtin operation. However, many genomic databases and input data files use the character string / factor notation.

Value

Character vector of same length as x, with strands reversed.

Author(s)

W. Huber <huber@ebi.ac.uk>

Examples

```
otherStrand(c("+", "-"))
```

plotAlongChrom

Plot signals and segmentation for a region of a chromosome

Description

Plot signals and segmentation for a region of a chromosome

Usage

```
plotAlongChrom(segObj, y, probeAnno, gff,
    isDirectHybe=FALSE,
    what = c("dots"), ## "heatmap"
    chr, coord, highlight,
    colors, doLegend=FALSE,
    featureExclude=c("chromosome", "nucleotide_match", "insertion"),
    featureColorScheme=1, extras,
    rowNamesHeatmap, rowNamesExtras, ylab, ylabExtras, main,
    colHeatmap=colorRamp(brewer.pal(9, "YlGnBu")),
    colExtras=colorRamp(brewer.pal(9, "Reds")),
    sepPlots=FALSE, reOrder=TRUE, ...)
```

Arguments

segObj Either an environment or an object of S4 class segmentation. See *Details*.

y a numeric vector or matrix containing the signal to be plotted. See *Details*.

probeAnno environment with probe annotations. See *Details*, and package davidTiling for an example.

gff data frame with genome annotation from the GFF file.

isDirectHybe logical scalar: if TRUE, the mapping of probes to genomic strands is reversed with respect to the default. This is appropriate for data from a direct RNA hy-

bridization that used no reverse transcription.

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what character scalar indicating which signal visualization to plot. Can be either dots

to plot each probe intensity with a point, or heatmap to produce a colorscale

representation of the intesities.

chr integer of length 1 indicating the chromosome number to plot.

coord integer vector of length 2 containing the start and end coordinates (in bases) for

the plot.

highlight (optional) list with two elements: a single numeric value coord and a character

strand. If present, this position is marked by a vertical red bar on the coordinate

axis. The color can be changed using the colors argument below.

colors (optional) named character vector. If missing, a default color scheme is used:

c("+"="#00441b", "-"="#081d58", "duplicated"="grey", "cp"="#101010", "highlight"="red", "threshold"="grey"), where the first three elements refer to the colors of data points and the last three to the colors of lines in the

olot.

doLegend logical: should the plot contain a legend?

featureExclude character vector of names of feature types (in gff) that should not be plot-

ted. Default is "chromosome", "nucleotide_match" and "insertion". Additional possibilities include: "ARS", "repeat_region", "repeat_family"

and "nc_primary_transcript".

featureColorScheme

numeric scalar, used to select a color scheme for the boxes representing genomic features such as coding sequences, ncRNAs etc. Currently the only value supported is 1 (see plotAlongChromLegend or plotFeatures for further infor-

mation).

extras a matrix containing additional values to be plotted along the chromosome in

a separate panel (such as p-values). This option is only available when y is

specified. These values should be on the scale [0,1].

rowNamesHeatmap

character vector of row names for the main heatmap.

rowNamesExtras character vector of row names for the extra heatmap.

ylab character label for y-axis of main plot.

ylabExtras character label for y-axis on extras panel (if specified).

main character: plot title.

colHeatmap function describing color scheme for the main heatmap plot (defaults to Y1GnBu

from RColorBrewer package).

colExtras function describing color scheme for the extra heatmap plot (if specified) (de-

faults to Reds from RColorBrewer package).

sepPlots logical scalar. If TRUE, each column of intensities in segObj or y is plotted

separately (maximum of 3) in the same figure. When FALSE, the average is

plotted. This argument is only used when what is set to dots.

reOrder logical scalar (only used when sepPlots is TRUE). If TRUE, the first column

of intensities is printed at the bottom of each plot, and the subsequent columns are plotted above. If FALSE, the first appears at the top, and the subsequent

columns are plotted below.

... further arguments that can be passed to the functions that implement the what

option above (see plotSegmentationDots and plotSegmentationHeatmap) or

gff plotting (see plotFeatures and plotAlongChromLegend).

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Details

Intensities: There are two alternative, mutually exclusive ways of providing the intensities that are to be plotted to this function.

- 1. Via the parameters y and probeAnno. In this case, y is a matrix of intensities, whose rows correspond to probes on the array, and its columns to different conditions, time points, etc. It is also acceptable that y is provided as a vector, in which case it is converted to an nrow(y) x 1 matrix. probeAnno is an environment whose elements correspond to target sequences (e.g. chromosome strands) and that contain integer vectors of length nrow(y) with information about the probes: start and end positions of their alignment to the target sequence, their row indices in y, the type of alignment (is it perfect? is is unique?). For example, the start positions and indices of probes for the + strand of chromosome 1 would be described by environment elements "1.+.start" and "1.+.index".
- 2. Via the parameter seg0bj.

segObj: This can be either an object of S4 class segmentation or an environment that by convention contains a certain set of objects. Future work on this package will focus on the S4 class segmentation. The environment option is provided for backward compatibility.

Explanation of the environment: the intended workflow is as follows: Use the script segment.R (in the inst/scripts directory of this package) to generate segmentations. This can be run in parallel on several processors, separately for each chromosome and strand. The results of this are stored in files of the name 1.+.rda, 1.-.rda, 2.+.rda, and so forth, typically within a dedicated directory. Then use the script readSegments.R to collect the R objects in these .rda files into the environment. It contains three types of data:

- microarray intensities in along-chromosome order.
- the segmentation objects (output of findSegments).
- a dataframe named segScore with segment scores; it can be missing iff nrBasesPerSeg is present,
- a numeric scalar names the Threshold, which is used to draw a horizontal "threshold" line in the plot.

... and the different signal visualization methods (what option): If what=="dots", the argument showConfidenceIntervals can be a logical scalar to choose whether vertical dashed lines are drawn for the confidence interval. In any case, these are only drawn if they are present in the segmentation object in segObj.

Author(s)

Wolfgang Huber <huber@ebi.ac.uk>

Examples

```
## 1. see viewSegmentation.R script in the inst/scripts directory
## 2. (newer): segmentation.Rnw
## 3. (newer): see the plotALongChrom vignette
data(segnf)
data(gffSub)
nmLabel = colnames(segnf$"1.+"@y)
plotAlongChrom(segnf,chr=1,coord=c(35000,50000),
    gff=gffSub,rowNamesHeatmap=nmLabel) ##the dots
plotAlongChrom(segnf,chr=1,coord=c(35000,50000),what="heatmap",
    gff=gffSub,rowNamesHeatmap=nmLabel) ##the heatmap
```

```
plotAlongChrom(segnf,chr=1,coord=c(35000,50000),gff=gffSub,
    showConfidenceIntervals=FALSE) ##do not show the segment confidence interval
```

plotAlongChromLegend Plot a legend for genomic features

Description

Plot a legend for genomic features

Usage

```
plotAlongChromLegend(vpr, nr=2,
    featureColorScheme=1,
    featureExclude=c("chromosome", "nucleotide_match", "insertion"),
    mainLegend, cexLegend=0.35, cexMain=1)
```

Arguments

vpr vector specifying where to place the legend in figure (set up by using the viewport

function from the grid package. When this function is called directly by the user

this argument should be left missing.

nr numeric scalar, specifying the number of rows to plot legend over (default value

is 2).

featureColorScheme

numeric scalar, used to select a color scheme for the boxes representing genomic features such as coding sequences, ncRNAs etc. Currently the only value

supported is 1.

featureExclude character vector of names of feature types (in gff) that should not be plotted.

Default is "chromosome", "nucleotide_match" and "insertion". Additional possible candidates include: "ARS", "repeat_region", "repeat_family"

and "nc_primary_transcript".

mainLegend character vector specifying legend title.

cexLegend numeric scalar specifying the magnification to be used for the legend text rela-

tive to the current text size.

cexMain numeric scalar specifying the magnification to be used for the legend title rela-

tive to the current text size.

Details

This function is usually called by plotAlongChrom when doLegend is TRUE. It can also be called directly by the user to produce a separate legend.

```
The following features are included in the legend (unless excluded using the featuredExclude option): "chromosome", "nucleotide_match", "pseudogene", "uORF", "nc_primary_transcript", "region", "repeat_family", "repeat_region", "transposable_element", "transposable_element_gene", "ARS", "centromere", "telomere", "insertion", "CDS", "CDS_dubious", "ncRNA", "tRNA", "snRNA", "snRNA", "binding_site" and "TF_binding_site".
```

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Author(s)

Wolfgang Huber <huber@ebi.ac.uk>

Examples

```
## plotAlongChromLegend(mainLegend="Legend")
```

plotFeatures

Plot genomic features for a region along a chromosome

Description

Plot genomic features for a region along a chromosome

Usage

```
plotFeatures(gff, chr, xlim, strand, vpr, featureColorScheme=1,
             featureExclude=c("chromosome", "nucleotide_match", "insertion"),
             featureNoLabel=c("uORF", "CDS"), ...)
```

Arguments

gff data frame with genome annotation from the GFF file. chr integer of length 1 specifying the chromosome to plot the features for. x1im integer of length 2 with start and end coordinates (in bases) for plotting. character scalar which should be set to either + or - to indicate which strand of strand DNA to plot the features from. vpr which viewport to plot the features in. featureColorScheme numeric scalar, used to select a color scheme for the boxes representing genomic features such as coding sequences, ncRNAs etc. Currently the only value supported is 1. featureExclude character vector of names of feature types (in gff) that should not be plotted. Default is "chromosome", "nucleotide_match" and "insertion". Additional possible candidates include: "ARS", "repeat_region", "repeat_family" and "nc_primary_transcript".

featureNoLabel character vector, names of feature types (in gff) that should not be labelled with their names (if they are plotted).

additional arguments.

Details

This function is called by plotAlongChrom when the gff argument has been specified. It should not be called directly by the user.

Author(s)

Wolfgang Huber <huber@ebi.ac.uk>

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plotPenLL

Plot the log-likelihood and penalized log-likelihoods (AIC, BIC)

Description

Plot the log-likelihood and two versions of penalized log-likelihoods (AIC, BIC) for a segmentation object.

Usage

```
plotPenLL(seg, extrabar=numeric(0), type="b", lty=1, pch=16, lwd=2, ...)
```

Arguments

```
seg A segmentation object. extrabar In addition to the location of maximmal BIC, vertical bars are drawn at these x-positions as well. type, pch, lty, lwd, \dots Get passed on to matplot.
```

Details

This function is used in the vignette: *How to use the segment function to fit a piecewise constant curve.*

Value

The function is called for its side effect, which is creating a plot in the current graphics device.

Author(s)

Wolfgang Huber < huber@ebi.ac.uk>

Examples

```
x = rep( sin((0:4)/2*pi), each=3) + rnorm(3*5, sd=0.1)
res = segment(x, maxseg=8, maxk=15)
plotPenLL(res)
```

Description

Plot points for a region along a chromosome

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Usage

Arguments

dat	list containing data to be plotted (see <i>Details</i> section below for particulars).
xlim	integer vector of length 2 with start and end coordinates (in bases) for plotting.
ylim	numeric vector containing the y limits of the plot.
ylab	character scalar (if sepPlots=FALSE) or vector containing y-axis label(s).
threshold	numeric scalar indicating the threshold of expression (default value is NA, for no threshold. If a value is supplied, it is subtracted from the intensity measures in dat\$y.
chr	integer of length 1 indicating the chromosome to be plot (defaults to 1).
strand	character scalar which should be set to either + or - to indicate which strand of DNA to plot the intensity values from (defaults to "+").
vpr	which viewport to plot the figure in. If this function is called directly by the user this argument should be left missing.
colors	named character vector, optional. If missing, a default color scheme is used: $c("+"="\#00441b", "-"="\#081d58", "duplicated"="grey", "cp"="#101010" "highlight"="red", "threshold"="grey"), where the first three elements refer to the colors of data points and the last three to the colors of lines in the plot.$
main	character vector specifying plot title.
pointSize	an object of class unit which specifies the size of each point. Default value is unit(0.6, "mm").
showConfidenceI	ntervals
	logical scalar indicating whether confidence intervals for each change-point are to be plotted (only available once segmentation has occurred).
sepPlots	logical scalar indicating whether the intensities are plotted separately for each array (if dat\$y has multiple columns). Defaults to FALSE, in which case the average intensity for each probe is plotted. When TRUE, up to 3 arrays can be plotted separately (more than 3 gets crowded).
cexAxisLabel	numeric scalar specifying the magnification to be used for the y-axis label relative to the current test size.
cexAxis	numeric scalar specifying the magnification to be used for the y-axis annotation relative to the current text size.

Details

. . .

This function is called by plotAlongChrom when the argument what is set to dots. Although this function can be called directly by the user, this is not recommended.

The dat list contains the following items: items x: x-coordinates (in bases) along chromosome,

1. y: intensity matrix of probes along chromosome,

additional arguments.

- 2. flag: indicates probe uniqueness in the genome. Possibilities are 3: multiple perfect matches, 2: has no PM but one or more near-matches, 1: has exactly one PM and some near-matches in the genome, 0: has exactly one PM and no near-matches.
- 3. extras: (optional) matrix of additional values (such as test-statistics/p-values) to be plotted.

Author(s)

Wolfgang Huber < huber@ebi.ac.uk>

 $\verb"plotSegmentationHeatmap"$

Plot a heatmap diagram for a region along a chromosome

Description

Plot a heatmap diagram for a region along a chromosome

Usage

Arguments

dat	list containing data to be plotted (see <i>Details</i> section below for particulars).
xlim	integer vector of length 2 with start and end coordinates (in bases) for plotting.
ylab	character scalar specifying y-axis label.
rowNames	character vector specifying a name for each row in the heatmap plot.
chr	integer of length 1 indicating the chromosome to plot (defaults to 1).
strand	character scalar which should be set to either + or - to indicate which strand of DNA to plot the intensity values from (defaults to "+").
vpr	which viewport to plot the figure in. If this function is called directly by the user this argument should be left missing.
colors	named character vector, optional. If missing, a default color scheme is used: $c("+"="\#00441b", "-"="\#081d58", "duplicated"="grey", "cp"="\#101010", "highlight"="red", "threshold"="grey"), where the first three elements refer to colors of data points and the last three to those of lines in the plot.$
colHeatmap	function describing color scheme for the heatmap plot (defaults to Y1GnBu from RColorBrewer package).

showConfidenceIntervals

logical scalar indicating whether confidence intervals for each change-point are to be plotted (only available once segmentation has occurred).

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just

character vector specifying the justification of the supplied values to the given coordinates; setting the first entry to "left" indicates that the supplied x-coordinates are the start positions of the probes, change this to "centre" if the x-coordinates are the probe middle positions. Usually the second entry should be "centre" (see grid.rect)

main

character vector specifying plot title.

makeRasterImage

logical scalar indicating whether to plot the heatmap image by the grid.raster (see grid.raster) or the grid.rect (see grid.rect) function in grid package. The default is to generate raster image, as it can be displayed much faster with a relatively smaller file size.

... additional arguments.

Details

This function is called by plotAlongChrom if the argument what is set to heatmap. Although this function can be called directly by the user, this is not recommended. The dat list contains the following items:

- x x-coordinates (in bases) along chromosome
- y intensity matrix of probes along chromosome

flag indicates probe uniqueness in the genome. Possibilities are 3: multiple perfect matches, 2: has no PM but one or more near-matches, 1: has exactly one PM and some near-matches in the genome, 0: has exactly one PM and no near-matches.

extras (optional) matrix of additional values (such as test-statistics/p-values) to be plotted

Author(s)

Wolfgang Huber <huber@ebi.ac.uk>

Examples

```
data(segnf)
data(gffSub)
nmLabel = colnames(segnf$"1.+"@y)
plotAlongChrom(segnf,chr=1,coord=c(35000,50000),what="heatmap",
    gff=gffSub,rowNamesHeatmap=nmLabel) ##using raster image
```

PMindex

Find the index of the exact match (PM) or background probes from a probeAnno environment

Description

Find the index of the exact match (PM) or background probes from a probeAnno environment

Usage

```
PMindex(probeAnno)
BGindex(probeAnno)
```

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Arguments

probeAnno

environment with probe annotations. See package davidTiling for an example (?probeAnno).

Details

These functions extract the exact match probes (PM) or background probes (from intergenic regions outside of known annotations) indices from probeAnno. These indices can be used to select the relevant rows of intensity data from the ExpressionSet object for plotting and normalization.

Value

Numeric vector of indices.

Author(s)

Matt Ritchie <ritchie@ebi.ac.uk>

Examples

```
## library(davidTiling)
## data(davidTiling)
## data(probeAnno)
## pmind <- PMindex(probeAnno)
## mmind <- MMindex(probeAnno)
## bgind <- BGindex(probeAnno)
## boxplot(as.data.frame(log2(exprs(davidTiling))[pmind,]), outline=FALSE)</pre>
```

posMin

Find the smallest positive number in a vector

Description

Find the smallest positive number in a vector

Usage

```
posMin(x, ...)
```

Arguments

x Numeric vector.

... Further arguments that get passed on to min.

Details

This is a rather trivial convenience function.

Value

Numeric of length 1.

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Author(s)

W. Huber <huber@ebi.ac.uk>

Examples

```
x = runif(5)
posMin(x-0.5)
posMin(x-2)
```

qcPlots

Generate simple diagnostic plots for Affymetrix tiling array data

Description

Generate simple diagnostic plots for Affymetrix tiling array data

Usage

extracted from probeAnno.

Arguments

x	ExpressionSet containing the data to be plotted.
html	logical scalar. If TRUE an html summary page 'qcsummary.htm' is generated. If FALSE, no summary page is generated.
plotdir	optional character string specifying the filepath where the plots will be saved. Defaults to current working directory.
probeAnno	environment with probe annotations. See package davidTiling for an example (?probeAnno).
gff	data frame with genome annotation from the GFF file.
chr	integer of length 1 indicating the chromosome number to plot.
coord	integer vector of length 2 containing the start and end coordinates (in bases) for the along chromosome intensity plot.
nr	integer, indicating the number of probes in each row on the array (2560 for yeast tiling arrays).
nc	integer, indicating the number of probes in each column on the array (2560 for yeast tiling arrays).
ylimchrom	numeric vector containing the y limits of the along chromosome intensity plot.
nucleicAcid	character vector or factor indicating what sample has been hybridised to each array. Used to color the boxplots and smoothed histograms of intensities.
pmindex	integer vector of indices of PM probes in x. If missing, this information is

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pgm	logical scalar. If TRUE, image plots will be saved as .pgm files. Otherwise (FALSE), they are converted to jpegs. On windows machines, this argument should be set to TRUE.
ext	character string indicating the file extension.
ranks	logical scalar. If TRUE, imageplots will show ranks of standardised probe intensities. Otherwise (FALSE, default), the standardised probe intensities are plotted.
	further arguments that can be passed to the plotting function plotSegmentationDots.

Details

This function creates boxplots, smoothed histogram (density) plots, imageplots and along chromosome plots of the raw (log base 2) probe intensity data.

An html page called 'qcsummary.htm' which displays the results, is created when html=TRUE.

Imageplots of standardised intensities (i.e. (probe intensity - minimum probe intensity) divided by the difference between the maximum and minimum probe intensities, all on log base 2 scale) or the ranks of these standardised intensities are plotted depending on the ranks argument.

The individual plots are named by replacing the file extension (specified by ext) of each 'celfile.ext', with 'density.png' for smoothed histogram plots, 'gencoord.jpg', for along chromosome plots and either 'log.pgm' ('log.jpg' if pgm=FALSE) or 'rank.pgm' ('rank.jpg' if pgm=FALSE) for the image-plots, depending on the ranks argument.

Author(s)

Matt Ritchie <ritchie@ebi.ac.uk> and Wolfgang Huber <huber@ebi.ac.uk>

Examples

```
## library(davidTiling)
## data(davidTiling)
## data(probeAnno)
## qcPlots(davidTiling, probeAnno)
```

readCel2eSet

Read celfiles into an ExpressionSet object.

Description

This is a wrapper for ReadAffy that returns an ExpressionSet object rather than an AffyBatch. This is particularly usefiles for arrays for which we have or need no CDF environment.

Usage

```
readCel2eSet(filename, adf, path=".", rotated=FALSE, ...)
```

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Arguments

filename Character vector with CEL file names. Either filename or adf need to be spec-

ified, but not both.

adf Object of class AnnotatedDataFrame.

path Character scalar with path to CEL files.

rotated Logical scalar, see details.

... Further arguments that are passed on to new("ExpressionSet").

Details

The rotate options allows to deal with different versions of the scanner software. Older versions rotated the image by 90 degrees, newer ones do not. Use the default rotated=FALSE for CEL files produced by the newer version.

Value

ExpressionSet object.

Author(s)

W. Huber

Examples

```
## To test the rotation, look at the scatterplot between two DNA hybes
## that were measured with scanner software that rotated (041120) and did
## not rotate (060125)
## cp /ebi/research/huber/Projects/tilingArray/Celfiles/041120_S96genDNA_re-hybe.cel.gz ~/p/tmp
## cp /ebi/research/huber/Projects/allelicTranscription/celfiles_allelictrans/060125_S96_genomicDNA.zip ~/p.
## cd ~/p/tmp
## gunzip 041120_S96genDNA_re-hybe.cel.gz
## unzip 060125_S96_genomicDNA.zip
##
## Not run:
library("affy")
options(error=recover)
e1 = readCel2eSet("041120_S96genDNA_re-hybe.cel", rotated=TRUE)
e2 = readCel2eSet("060125_S96_genomicDNA.CEL")
smoothScatter(log(exprs(e1)), log(exprs(e2)), nrpoints=0)
## End(Not run)
```

sampleStep 23

camp	leSten
samo	resten

Sampling of ascending numbers to ensure minimal spacing.

Description

Given a vector of ascending numbers and a step width, sample the numbers such that the difference between consecutive numbers is greater than or equal to step.

Usage

```
sampleStep(x, step)
```

Arguments

x Numeric or integer vector.

step Numeric scalar.

Details

The simple algorithm works greedily from x[1] to x[length(x)]. First, x[1] is selected. Then, if x[i] is selected, all numbers x[j] with j>i and x[j]-x[i]<step are dropped. Then, i is set to the smallest j with x[j]-x[i]>=step.

Value

A logical vector of the same length as x, representing the selected subsample.

Author(s)

W. Huber <huber@ebi.ac.uk>

Examples

```
x = sort(as.integer(runif(20)*100))
sel = sampleStep(x, step=10)
x
x[sel]
```

segChrom

Fit a piecewise constant curve to along chromosome data (wrapper function)

Description

Wrapper around the segment function for each strand of one or more chromosomes specified by the user. It does some typical preprocessing and I/O.

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Usage

```
segChrom(y, probeAnno, chr=1:17, strands=c("+", "-"),
nrBasesPerSegment = 1500, maxk = 3000, step = 7, confint = FALSE,
confintLevel = 0.95, useLocks=TRUE, verbose=TRUE, savedir)
```

Arguments

y ExpressionSet or matrix containing the data to be segmented.

probeAnno an object of class probeAnno (defined in the Ringo package) or an environment

with probe annotations. For the latter, see the package davidTiling for an

example (?probeAnno).

chr integer scalar or vector specifying which chromosome(s) to segment.

strands character scalar or vector specifying which strands to segment; can also be NA.

nrBasesPerSegment

integer (length 1): the parameter maxseg of the segment function is calculated as the length of the chromosome divided by nrBasesPerSegment. Thus, it de-

termines the average segment length in the finest segmentation.

maxk passed on to the function segment.

step integer scalar, indicating the minimum distance between consecutive probes. In

cases when probes are offset by less than step bases, the probes are sampled to

achieve the desired spacing.

confint logical scalar. If TRUE, confidence intervals for each change-point are calculated.

confintLevel numeric scalar between 0 and 1 indicating the probability level for the confi-

dence intervals that are calculated for each change-point.

useLocks logical scalar. Should a file locking mechanism be used that allows for a simple-

minded parallelization of this function.

verbose logical scalar. Should we be chatty about our progress?

savedir character scalar. If specified, resulting segmentation objects are saved (with

save) to this directory.

Details

This function is a wrapper for the segment function. It is provided in this package for illustration. For applications to different datasets, you will likely need to adapt it to some extent, please refer to its source code.

Value

An environment containing S4 objects of class "segmentation" called "1.+", "1.-", etc. (depending on the values in chr and strands), where "+" and "-" indicate the strand and the preceding number refers to the chromosome. If savedir is specified, there is also the side-effect that a series of files "1.+.rda", "1.-.rda", etc. is saved in that directory.

Author(s)

Wolfgang Huber < whuber@embl.de>

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Examples

```
## Not run:
    library("davidTiling")
    data("davidTiling")
    data("probeAnno")
    isDNA = seq(1:3)
    yn = normalizeByReference(davidTiling[,-isDNA],davidTiling[,isDNA], probeAnno=probeAnno)
    seg = segChrom(yn, probeAnno) ## this will take a while to run!
## End(Not run)
```

segment

Fit a piecewise constant curve: segmentation by dynamic programming

Description

The function fits a piecewise constant curve to one or multiple sequences of measurements, using a least squares cost function and an O(n) dynamic programming algorithm (see references).

Usage

```
segment(y, maxseg, maxk)
```

Arguments

y Numeric matrix. Rows correspond to the x-variable, columns to replicate mea-

surements at the same value of x. Breakpoints are fitted along the x-axis. For example, the x-variable can be genomic coordinates or time. The segmentation

will be along the rows of y.

maxseg integer of length 1, maximum number of segments (= 1 + maximum number of

change points).

maxk integer of length 1, maximum length of a single segment.

Details

The complexity of the algorithm is length(x)*maxk in memory and length(x)*maxk*maxseg in time.

Value

An object of class segmentation.

Author(s)

```
W. Huber <huber@ebi.ac.uk>
```

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References

[1] Transcript mapping with high-density oligonucleotide tiling arrays. Huber W, Toedling J, Steinmetz, L. Bioinformatics 22, 1963-1970 (2006).

[2] A statistical approach for CGH microarray data analysis. Franck Picard, Stephane Robin, Marc Lavielle, Christian Vaisse, Gilles Celeux, Jean-Jacques Daudin. BMC Bioinformatics. 2005 Feb 11; 6:27.

Examples

```
x = rep(sin((0:4)/2*pi), each=3) + rnorm(3*5, sd=0.1)
res = segment(x, maxseg=6, maxk=15)
```

segmentation

The class segmentation represents a segmentation result.

Description

This class represents the result of a segmentation, usually a call to the function segment.

Objects from the Class

Objects can be created by calls of the function segment or by calls of the form new("segmentation", \dots).

Slots

- y: A matrix with the data (the dependent variable(s)), see segment.
- x: A numeric vector with the regressor variable. The length of this vector must be either the same as nrow(y), or 0. The latter case is equivalent to x=1:nrow(y).
- flag: An integer vector, whose length must be either the same as nrow(y), or 0. This can be used to *flag* certain probes for special treatment, for example by plotAlongChrom.
- breakpoints: List of segmentations. The element breakpoints[[j]] corresponds to a segmentation fit of j segments, i.e. with j-1 breakpoints. It is a matrix with (j-1) rows and 1 or 3 columns. It always contains a column named estimate with the point estimates. Optionally, it may contain columns lower and upper with the confidence intervals. The point estimates are the row indices in y where new segments start, for example: let z=breakpoints[[j]], then the first segment is from row 1 to z[1, "estimate"]-1, the second from row z[1, "estimate"] to z[2, "estimate"]-1, and so on.
- logLik: Numeric vector of the same length as breakpoints, containing the log-likelihood of the piecewise constant models under the data y.
- hasConfint: Logical vector of the same length as breakpoints. TRUE if the confidence interval estimates are present, i.e. if the matrix breakpoints[[j]] has columns lower and upper.
- nrSegments: A scalar integer, value must be either NA or between 1 and length(breakpoints). Can be used to select one of the fits in breakpoints for special treatment, for example by plotAlongChrom.

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Methods

confint The method confint(object, parm, level=0.95,het.reg=FALSE, het.err=FALSE, ...) computes confidence intervals for the change point estimates of the segmentation. Typically, these were obtained from a previous call to the function segment that created the object. This is just a wrapper for the function confint.breakpointsfull from the strucchange package, which does all the hard computations. Parameters: object an object of class segmentation, parm an integer vector, it determines for which of the segmentation fits confidence intervals are computed. See also segment. The other parameters are directly passed on to confint.breakpointsfull.

logLik The method logLik(object, penalty="none", ...) returns the log-likelihoods of fitted models. Valid values for the argument penalty are none, AIC and BIC.

plot The method plot(x, y, xlim, xlab="x", ylab="y", bpcol="black", bplty=1, pch=16, ...) provides a simple visualization of the result of a segmentation. Parameters: x an object of class segmentation, y an integer between 1 and length(x@breakpoints), selecting which of the fits contained in x to plot, bpcol and bplty color and line type of breakpoints. The plot shows the numeric data along with breakpoints and if available their confidence intervals.

show summary.

Author(s)

Wolfgang Huber < huber@ebi.ac.uk>

See Also

segment

Examples

```
## generate random data with 5 segments:
y = unlist(lapply(c(0,3,0.5,1.5,5), function(m) rnorm(10, mean=m)))
seg = segment(y, maxseg=10, maxk=15)
seg = confint(seg, parm=c(3,4,5))
if(interactive())
   plot(seg, 5)
show(seg)
```

segnf

Example of a segmentation output object

Description

Example of a segmentation output object

Usage

```
data(segnf)
```

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Details

segnf is an environment which contains two segmentation object for the "+" and "-" strand of yeast chromosome one. Each segmentation object contains the tiling array expression profiling between condition YPE and YPD, 3 replicates each. The raw array data are normalized, mapped to the yeast genome and segmented using the segChrom function. The object is the output of segChom with some small modification. Only a small region (35000bp-50000bp) is included in this data

Author(s)

Zhenyu Xu <zhenyu@ebi.ac.uk>

Examples

```
data(segnf)
data(gffSub)
nmLabel = colnames(segnf$"1.+"@y)
plotAlongChrom(segnf,chr=1, coord=c(35000,50000),what="heatmap",
    gff=gffSub,rowNamesHeatmap=nmLabel)
```

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