

# MassArray

October 25, 2011

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MassArray.example.data  
*MassArray Data object*

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## Description

This data contains MassArray spectral information for two samples.

## Usage

MassArray.example.data

## Format

The format is: Formal class 'MassArrayData' [package "MassArray"] with 17 slots ..@ sequence : chr "CCAGGTCCAAAGGTTTCAGACCAGTCTGAACCTGTCCAGGGGCACTCCATATTTTCCTACCTGTCCCTCTTTGCTTGTA AAAACAAATTA AACAGGGATCCCAGCAACTTCGGGGGCATGTGTGTAAC T\_\_truncated\_\_ ..@ chr : chr(0) ..@ start : int(0) ..@ end : int(0) ..@ strand : chr "+" ..@ fwd.tag : chr "AGGAAGAGAG" ..@ rev.tag : chr "AGCCTTCTCCC" ..@ fwd.primers : num 29 ..@ rev.primers : num 27 ..@ lower.threshold : num 1500 ..@ upper.threshold : num 9000 ..@ fragments.T :List of 89 .. .\$ :Formal class 'MassArrayFragment' [package "MassArray"] with 21 slots .. ..@ ID : int 1 .. ..@ assay.name : chr "" .. ..@ name : chr "" .. ..@ sequence : chr "GGGAGAAGGCT" .. ..@ position : int 385 .. ..@ length : int 11 .. ..@ CpGs : int 0 .. ..@ MW : num 3913 .. ..@ collisions : int 0 .. ..@ collision.IDs :List of 1 .. ..@ .. .\$ : int(0) .. ..@ CG.collisions : int 0 .. ..@ CG.collision.IDs : list() .. ..@ type : chr "T" .. ..@ direction : chr "+" .. ..@ extra : chr "5PPP-3P" .. ..@ bisulfite.converted: logi TRUE .. ..@ assayable : logi TRUE .. ..@ conversion.control : logi FALSE .. ..@ required : logi FALSE .. ..@ ignored : logi FALSE .. ..@ primer : logi TRUE ..@ samples :List of 2 .. .\$ :Formal class 'MassArraySpectrum' [package "MassArray"] with 9 slots .. ..@ sample : chr "A" .. ..@ rxn : chr "T" .. ..@ strand : chr "+" .. ..@ peaks :List of 184 .. ..@ .. .\$ :Formal class 'MassArrayPeak' [package "MassArray"] with 16 slots .. ..@ ID : int 1 .. ..@ MW.theoretical : num 1111 .. ..@ MW.actual : num NA .. ..@ probability : num 0 .. ..@ SNR : num 0 .. ..@ height : num NA .. ..@ sample.intensity: num NA .. ..@ ref.intensity : num 0.1 .. ..@ ..@ sequence : chr "ACACAAT" .. ..@ adduct : chr "" .. ..@ type : chr "Modified" .. ..@ charge : int 1 .. ..@ collisions : int 0 .. ..@ components : int 0 .. ..@ missing : logi TRUE .. ..@ new : logi FALSE .. ..@ quality.conversion : num [1:4] 0.0529 0 0 0 .. ..@ quality.spectra : num NA .. ..@

```
quality.primerdimer: num [1:7] 8.51 15.83 3.28 1.04 0 ... ..@ quality.contaminant: num NA
.. ..@ quality.adducts : num [1:114] 1 1 0.97 0.231 0.412 ... ..@ groups : chr(0) ..@ CpG.data :
num [1:2, 1:18] 0.0322 0.0449 0.1468 0.3641 0.1468 ... ..@ CpG.data.combined: num [1:2, 1:18]
0.0322 0.0449 0.1468 0.3641 0.1468 ...
```

### Source

Thompson et al. 2009

### Examples

```
data(MassArray.example.data)
```

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MassArrayData-class

*Class "MassArrayData"*

---

### Description

A data structure containing MassArray data and associated information for a single amplicon

### Objects from the Class

Objects can be created by calls of the form `new("MassArrayData", sequence, file, verbose, fwd.tag, rev.tag, fwd.primer, rev.primer, strand, lower.threshold, upper.threshold, header, skip, sep, comment.char, fill, method, position, ...)`.

### Slots

`sequence`: Nucleotide sequence for unconverted amplicon

`chr`: Chromosomal position of amplicon

`start`: Chromosomal position of amplicon

`end`: Chromosomal position of amplicon

`strand`: DNA strand used for amplicon (can be '+' or '-')

`fwd.tag`: Nucleotide tag sequence 5' of the forward primer

`rev.tag`: T7-containing nucleotide tag sequence 5' of the reverse primer

`fwd.primer`: Length (in bp) of forward primer

`rev.primer`: Length (in bp) of reverse primer

`lower.threshold`: Lower limit (in Da) of usable mass window (default: '1500')

`upper.threshold`: Upper limit (in Da) of usable mass window (default: '7000')

`fragments.T`: List containing objects of class `MassArrayFragment`, corresponding to the T-cleavage reaction for the amplicon on the specified `strand`

`fragments.C`: List containing objects of class `MassArrayFragment`, corresponding to the C-cleavage reaction for the amplicon on the specified `strand`

`samples`: List containing object of class `MassArraySpectrum`, each corresponding to spectral data from a single sample

**groups:** List of the group name to which each sample belongs

**CpG.data:** Matrix containing analyzed methylation data, where each row is a sample and each column is a CG dinucleotide site

**CpG.data.combined:** Matrix containing methylation data combined from multiple objects (or collapsed from within a single object), where each row is a sample and each column is a CG dinucleotide site

### Methods

`\$ signature(x = "MassArrayData"):...`

`\$<- signature(x = "MassArrayData"):...`

`[ signature(x = "MassArrayData"):...`

`initialize signature(.Object = "MassArrayData"):...`

### Author(s)

Reid F. Thompson (<rthompso@aecom.yu.edu>), John M. Greally (<jgreally@aecom.yu.edu>)

### Examples

```
showClass("MassArrayData")
```

---

```
MassArrayFragment-class
      Class "MassArrayFragment"
```

---

### Description

A data structure containing information for a single fragment of an amplicon

### Objects from the Class

Objects can be created by calls of the form `new("MassArrayFragment", ID, sequence, assay.name, name, position, type, direction, extra, bisulfite.converted, assayable, primer, ...)`.

### Slots

**ID:** Unique integer indexing the fragment's position within a potential list of multiple fragments

**assay.name:** (currently not supported)

**name:** (currently not supported)

**sequence:** Bisulphite converted nucleotide sequence of fragment

**position:** Relative position of fragment within the amplicon

**length:** Length (in bp) of fragment sequence

**CpGs:** Number of CG dinucleotides contained within the fragment

**MW:** Predicted molecular weight(s) of fragment, including methylated and unmethylated mass, adducts, etc.

**collisions:** Number of fragments that share the same molecular weight as the current fragment

`collision.IDs`: IDs of other fragments that share the same molecular weight as the current fragment

`CG.collisions`: Number of CG-containing fragments that share the same molecular weight as the current fragment

`CG.collision.IDs`: IDs of other CG-containing fragments that share the same molecular weight as the current fragment

`type`: Specifies either 'T' or 'C' cleavage reaction

`direction`: DNA strand used for fragment sequence (can be '+' or '-')

`extra`: One of "5PPP-3P", "5OH", or "5PPP-3OH" (default)

`bisulfite.converted`: Logical indicating whether the fragment sequence represents bisulfite converted sequence

`assayable`: Logical indicating whether or not the fragment molecular weight is within the usable mass window

`conversion.control`: Logical indicating whether or not the fragment is designated as a potential conversion control

`required`: Logical indicating whether or not the fragment is designated as 'required' by the user

`ignored`: Logical indicating whether or not the fragment is to be ignored

`primer`: Logical indicating whether or not the fragment overlaps with primer or tagged sequence

## Methods

```
\$ signature(x = "MassArrayFragment"): ...
\${<- signature(x = "MassArrayFragment"): ...
initialize signature(.Object = "MassArrayFragment"): ...
```

## Author(s)

Reid F. Thompson (<rthompso@aecom.yu.edu>), John M. Greally (<jgreally@aecom.yu.edu>)

## See Also

See Also [MassArrayData](#)

## Examples

```
showClass("MassArrayFragment")
```

---

```
MassArrayPeak-class
      Class "MassArrayPeak"
```

---

## Description

A data structure containing information and data for a single peak from a single spectrum

## Objects from the Class

Objects can be created by calls of the form `new("MassArrayPeak", ID, MW.theoretical, MW.actual, probability, SNR, height, sample.intensity, ref.intensity, sequence, adduct, type, charge, collisions, components, missing, new, ...)`.

## Slots

**ID:** Peak ID indicating indexed position within a potentially large list of peaks  
**MW.theoretical:** Expected molecular weight of peak based on nucleotide sequence  
**MW.actual:** Observed molecular weight from experimental data  
**probability:** Object of class "numeric" ~~  
**SNR:** Signal-to-noise ratio  
**height:** Raw peak height  
**sample.intensity:** Raw sample intensity  
**ref.intensity:** Object of class "numeric" ~~  
**sequence:** Nucleotide composition or sequence(s) corresponding to peak  
**adduct:** One of 'Na', 'K', or '' indicating whether or not peak represents a salt adduct of another expected peak  
**type:** Object of class "character" ~~  
**charge:** Degree of ionization of fragment (default is '1' indicating a single positive charge per fragment)  
**collisions:** Number of peaks that share the same molecular weight as the current peak  
**components:** Number of fragments expected to give rise to a peak of this molecular weight  
**missing:** Logical indicating whether or not the expected peak is missing from the spectral data  
**new:** Logical indicating whether or not the observed peak is unexpected given the amplicon sequence

## Methods

```
\$ signature(x = "MassArrayPeak"): ...  
\$<- signature(x = "MassArrayPeak"): ...  
initialize signature(.Object = "MassArrayPeak"): ...
```

## Author(s)

Reid F. Thompson (<rthompso@aecom.yu.edu>), John M. Greally (<jgreally@aecom.yu.edu>)

## See Also

See Also [MassArrayData](#)

## Examples

```
showClass("MassArrayPeak")
```

---

MassArraySpectrum-class

*Class "MassArraySpectrum"*

---

### Description

A data structure containing MassArray spectral data for a single sample

### Objects from the Class

Objects can be created by calls of the form `new("MassArraySpectrum", sample, rxn, strand, peaks, quality.conversion, quality.spectra, quality.primerdimer, quality.contaminant, quality.adducts, ...)`.

### Slots

`sample`: Sample name

`rxn`: Cleavage reaction (either 'T' or 'C')

`strand`: DNA strand for amplicon (either '+' or '-')

`peaks`: List containing objects of class `MassArrayPeak`

`quality.conversion`: Overall level(s) of remnant unconverted cytosines, as measured by one or more conversion controls

`quality.spectra`: (currently not supported)

`quality.primerdimer`: (currently not supported)

`quality.contaminant`: (currently not supported)

`quality.adducts`: Overall ratio(s) of Na and/or K adduct peak heights to expected peak heights

### Methods

`\$` signature(x = "MassArraySpectrum"): ...

`\$<-` signature(x = "MassArraySpectrum"): ...

**initialize** signature(.Object = "MassArraySpectrum"): ...

### Author(s)

Reid F. Thompson (<rthomps@aecon.yu.edu>), John M. Greally (<jgreally@aecon.yu.edu>)

### See Also

See Also [MassArrayData](#)

### Examples

```
showClass("MassArraySpectrum")
```

---

ampliconPrediction *Amplicon prediction*

---

### Description

Function to predict amplicon fragmentation pattern and details for T&C reactions on the plus and minus strands

### Usage

```
ampliconPrediction(sequence, lower.threshold = 1500, upper.threshold = 7000, fwd
```

### Arguments

sequence	Nucleotide sequence input as a character string
lower.threshold	Lower limit (in Da) of usable mass window (default: '1500')
upper.threshold	Upper limit (in Da) of usable mass window (default: '7000')
fwd.tag	Nucleotide tag sequence 5' of the forward primer
rev.tag	T7-containing nucleotide tag sequence 5' of the reverse primer
plot	Logical specifying whether or not to display graphical representation of fragmentation profiles (default is TRUE)
table	Logical specifying whether or not to return tabular representation of fragmentation profiles (default is TRUE)
lwd	The line width used for fragmentation display, a positive number, defaulting to 1
cex	A numerical value (defaulting to 1) giving the amount by which plotting text and symbols should be magnified relative to the default
multiple.conversion	Logical value specifying whether or not to include multiple CGs on the same conversion control fragment where possible (default is FALSE).

### Details

Plotted fragmentation patterns contain a number of detailed features including: CG positions, molecular weight overlaps, conversion controls, fragment assayability, and more.

Note that the graphical output does not contain a built-in legend at this time, but the plot may be interpreted as follows: Putative fragmentation patterns are shown for T and C-cleavage reactions on both the plus and minus strands of an input amplicon, with the T-forward, T-reverse, C-forward, and C-reverse shown in descending order. CG dinucleotides (filled circles) are numbered and colored in blue. Other fragments are colored according to their ability to be assayed: fragment molecular weight outside the testable mass window (gray), fragment molecular weight overlapping with another fragment (red), fragment containing a potential conversion control (green), or fragment uniquely assayable but containing no CGs (black). Linked arrowheads denote molecular weight overlaps between multiple CG-containing fragments. Yellow highlights represent tagged or primer sequences, while lavender highlights denote user-specified "required" sites.

**Value**

If `table` is `TRUE`, returns a list containing the following items:

<code>summary</code>	A summary matrix of logical values specifying whether or not each CG is assayable by a given combination of cleavage reaction and DNA strand
<code>counts</code>	A numerical tally of the quantity of CGs that are assayable by each assay

**Author(s)**

Reid F. Thompson (<rthompso@aeom.yu.edu>), John M. Grealley (<jgreally@aeom.yu.edu>)

**Examples**

```
ampliconPrediction("TGGAACACCCAGCAAAGATCAAGCAGGAAAGGGCGCACGCAGCCTTCGTTGCTAACCTCCTCTGGACTC")
```

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analyzeCpGs	<i>Analyze CG methylation</i>
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**Description**

Function to determine percent methylation for all CGs from input fragmentation

**Usage**

```
analyzeCpGs(fragments, peaks, method = c("weighted", "proportion"))
```

**Arguments**

<code>fragments</code>	List of <code>MassArrayFragment</code> objects
<code>peaks</code>	List of <code>MassArrayPeak</code> objects comprising spectral data for a given assay
<code>method</code>	Specifies which algorithm to use when calculating percent methylation (either "weighted" or "proportion")

**Details**

Wrapper function for `calcMeth()`, takes fragmentation pattern and spectral data as input and applies percent methylation calculation for all CG-containing, non conversion control fragments

**Value**

Returns a list of numerical values corresponding to percent methylation for each CG dinucleotide, with 0

**Author(s)**

Reid F. Thompson (<rthompso@aeom.yu.edu>), John M. Grealley (<jgreally@aeom.yu.edu>)

**See Also**

See Also [calcMeth](#)



**Examples**

```
data(MassArray.example.data)
cpg.data <- analyzeCpGs(MassArray.example.data$fragments.T, MassArray.example.data$sample)
barplot(cpg.data, xlab="CpG (Number)", ylim=c(0,1), ylab="Methylation (Percent)")
```

---

bisConvert	<i>Bisulphite conversion</i>
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**Description**

Bisulphite convert nucleotide sequence input

**Usage**

```
bisConvert(sequence)
```

**Arguments**

sequence      Nucleotide sequence in the form of a character string

**Value**

Returns a character value corresponding to the bisulphite converted input sequence.

**Author(s)**

Reid F. Thompson (<rthomps@aecon.yu.edu>), John M. Greally (<jgreally@aecon.yu.edu>)

**Examples**

```
bisConvert("AAATTCGGAACCC")
```

---

calcMW	<i>Calculate molecular weight</i>
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---

**Description**

Function to calculate molecular weight of a fragment generated by the MassCLEAVE assay for either the T or C cleavage reactions

**Usage**

```
calcMW(sequence, extra = c("5OH", "5PPP-3P", "5PPP-3OH"), adduct = c("", "Na", "
```

**Arguments**

sequence	Nucleotide sequence input
extra	One of "5OH" (default), "5PPP-3P", or "5PPP-3OH" describing 5' and/or 3' modifications of the fragment
adduct	One of 'Na', 'K', or '' (default) specifying whether or not the molecular weight should be calculated for a salt adduct
rxn	One of 'T' or 'C' indicating which cleavage reaction is employed to generate the fragment

**Value**

Returns a numerical output corresponding to the molecular weight (in Da) of sequence input. Note that the output may actually represent multiple molecular weights if/whenever the input sequence contains one or more degenerate bases (e.g. R or Y).

**Author(s)**

Reid F. Thompson (<rthompso@aeocom.yu.edu>), John M. Greally (<jgreally@aeocom.yu.edu>)

**Examples**

```
calcMW("AAATCCC")
calcMW("AARTYCC")
```

---

calcMeth	<i>Calculate percent methylation</i>
----------	--------------------------------------

---

**Description**

Function to calculate percent methylation from a collection of peaks corresponding to a single fragment.

**Usage**

```
calcMeth(SNR.list, fragments = rep(1, length(SNR.list)), non.cg.fragments = nume
```

**Arguments**

SNR.list	List of signal-to-noise ratios, sorted from low to high MWs, corresponding to the unmethylated and methylated peaks for a given set of fragments
fragments	List of all fragments contributing to each of the input peaks, automatically defaulting to a single fragment
non.cg.fragments	List of fragments (without CGs) contributing to any of the input peaks, automatically defaulting to numeric(0)
method	Specifies which algorithm to use when calculating percent methylation (either "weighted" or "proportion")

<code>prune.non.cg.peaks</code>	Boolean value determining whether or not to remove non-CG-containing fragments prior to analysis or whether to include them in the calculating model (note that setting this option to <code>FALSE</code> could result in a considerable increase in analytical time); option automatically defaults to <code>TRUE</code>
<code>na.rm</code>	Boolean value determining whether or not to return an error on input of any unspecified data (NA), automatically defaulting to <code>FALSE</code>

### Details

Note that the current release of this function performs as expected for the large majority of cases. However, certain complex combinations of peak overlaps are not handled at this time. This may affect data for a minority of points, particularly those containing multiple overlaps with alternative fragments. Please ensure more in-depth review of such loci.

### Value

Returns a numerical values corresponding to percent methylation, with 0

### Author(s)

Reid F. Thompson (<rthomps@aeom.yu.edu>), John M. Greally (<jgreally@aeom.yu.edu>)

### References

Coolen, M.W., et al. (2007) Genomic profiling of CpG methylation and allelic specificity using quantitative high-throughput mass spectrometry: critical evaluation and improvements, *Nucleic Acids Research*, **35**(18), e119.

### See Also

See Also [MassArrayPeak](#)

### Examples

```
data(MassArray.example.data)
frags <- MassArray.example.data$fragments.T[[6]]$"MW"
peaks <- findPeaks(frag, unlist(lapply(MassArray.example.data$samples[[1]]$peaks, slot,
SNRs <- unlist(lapply(MassArray.example.data$samples[[1]]$peaks[peaks], slot, "SNR"))
frag.list <- list(1:3, 1:3, 1:3, 1:3)
calcMeth(SNRs, fragments=frag.list, method="weighted")
calcMeth(SNRs, fragments=frag.list, method="proportion")
```

---

calcPercentAdduct *Calculate percent salt adducts*

---

### Description

Function to calculate ratio of salt adduct peak heights to reference/unmodified peaks

### Usage

```
calcPercentAdduct(peaks)
```



**Author(s)**

Reid F. Thompson (<rthompso@aeacom.yu.edu>), John M. Greally (<jgreally@aeacom.yu.edu>)

**See Also**

See Also [calcMeth](#), [convControl](#)

**Examples**

```
data(MassArray.example.data)
conversion.data <- calcPercentConversion(MassArray.example.data$fragments.T, MassArray.ex
mean(conversion.data)

# NOTE: conversion control data may already be contained within a MassArrayData object; t
conversion.data <- unlist(lapply(lapply(MassArray.example.data$samples, slot, "quality.co
barplot(conversion.data)
```

---

combine-methods      *Combine MassArrayData objects (methods)*

---

**Description**

Methods for joining two MassArrayData objects by sequence positions and samples, or simply operating on a single MassArrayData object to combine samples, depending on input

**Methods**

**x = "MassArrayData", y = "MassArrayData"** Combine two MassArrayData objects by position and then by sample

**x = "MassArrayData", y = "missing"** Combine duplicate samples within the same MassArrayData object

**See Also**

See Also [combine](#)

---

combine      *Combine MassArrayData objects*

---

**Description**

Function to join two MassArrayData objects by sequence positions and samples

**Usage**

```
combine(x, y, ...)
```

**Arguments**

<code>x</code>	MassArrayData object
<code>y</code>	MassArrayData object
<code>...</code>	Other arguments passed to combine not supported at this time.

**Value**

Returns a single MassArrayData object that contains a union of samples and amplicons and spectral data from both MassArrayData objects in input

**Author(s)**

Reid F. Thompson (<rthompso@aecom.yu.edu>), John M. Grealley (<jgreally@aecom.yu.edu>)

**See Also**

See Also [MassArrayData](#)

**Examples**

```
data(MassArray.example.data)
samples(MassArray.example.data)
combined.data <- combine(MassArray.example.data[2,], MassArray.example.data[1,])
samples(combined.data)
```

---

convControl

*Conversion control*

---

**Description**

Function to identify all potential conversion controls in a given input sequence, for a given list of fragments

**Usage**

```
convControl(sequence, fragmentation, multiple = FALSE)
```

**Arguments**

<code>sequence</code>	Nucleotide sequence input as a character string
<code>fragmentation</code>	List of MassArrayFragment objects corresponding to the fragmentation pattern of the sequence input
<code>multiple</code>	Logical value specifying whether or not to include multiple CGs on the same conversion control fragment where possible (default is FALSE).

**Details**

Potential conversion controls are identified from the nucleotide sequence input through pattern recognition of fragments that contain non-CG cytosines. Any conversion controls that contain CG dinucleotides or have a molecular weight outside of the usable mass window are screened out. Additionally, conversion controls that are located in identified primer sequence or have molecular weight overlap with other fragments are removed from consideration. Lastly, if the consideration of the fragment as a conversion control will cause new molecular weight overlap(s) with one or more other fragments, the control is also removed from consideration.

**Value**

Returns a list of `MassArrayFragment` objects identical to the input, with the exception that conversion controls are labeled and updated accordingly.

**Author(s)**

Reid F. Thompson (<rthompso@aeacom.yu.edu>), John M. Greally (<jgreally@aeacom.yu.edu>)

**See Also**

See Also as [MassArrayFragment](#)

**Examples**

```
data(MassArray.example.data)
MassArray.example.data$fragments.T[[54]]
conversion.data <- convControl(MassArray.example.data$sequence, MassArray.example.data$fr
conversion.data[[54]]
```

---

countCGs

*Count number of CGs*

---

**Description**

Function to count the number of CG dinucleotides in a given sequence (can include special characters for degenerate bases - i.e. 'Y' or 'R')

**Usage**

```
countCGs(sequence)
```

**Arguments**

sequence      Nucleotide sequence input as a character string

**Value**

Returns a numerical count of the number of CG dinucleotides in a given sequence, 'NA' if sequence input is 'NA'

**Author(s)**

Reid F. Thompson (<rthompso@aeacom.yu.edu>), John M. Greally (<jgreally@aeacom.yu.edu>)

**Examples**

```
countCGs ("AAACGCGAAAAAAAYGAAA")
```

---

```
createWiggle          Create wiggle track
```

---

**Description**

Function to create and write a wiggle track (UCSC Genome Browser format) to flat file from methylation data contained in a MassArrayData object

**Usage**

```
createWiggle(x, file = "", append = FALSE, colors = NULL, na.rm = FALSE, sep = "
```

**Arguments**

x	MassArrayData object containing methylation data for at least one sample.
file	location of file to write wiggle track information; if "", wiggle track prints to the standard output connection: see <a href="#">cat</a> .
append	logical; if 'TRUE', the output is appended to an existent wiggle track file. If 'FALSE' (default), a new file with a new header is created and any existing file of the same name is destroyed.
colors	vector of colors, indicates which colors to use for which wiggle track
na.rm	logical; if 'TRUE' (default), missing values are removed from data. If 'FALSE' any missing values cause an error
sep	a string used to separate columns. Using 'sep = "\t"' (default) gives tab-delimited output.

**Author(s)**

Reid F. Thompson (<rthomps@aecon.yu.edu>), John M. Greally (<jgreally@aecon.yu.edu>)

**References**

UCSC Genome Browser, <http://genome.ucsc.edu/goldenPath/help/customTrack.html>: Kent, W.J., Sugnet, C. W., Furey, T. S., Roskin, K.M., Pringle, T. H., Zahler, A. M., and Haussler, D. The Human Genome Browser at UCSC. *Genome Res.* **12**(6), 996-1006 (2002).

**See Also**

[write](#), [cat](#)

**Examples**

```
data(MassArray.example.data)
createWiggle(MassArray.example.data)
```



---

`estimatePrimerDimer`*Estimate Primer Dimers*

---

## Description

Function to estimate level of signal due to primer dimers in a given spectrum

## Usage

```
estimatePrimerDimer(fragments, peaks, method = c("ratio", "mann-whitney"))
```

## Arguments

<code>fragments</code>	List of <code>MassArrayFragment</code> objects corresponding to the sample
<code>peaks</code>	List of <code>MassArrayPeak</code> objects comprising spectral data for a complete assay
<code>method</code>	Specifies which algorithm to use when estimating primer dimer levels (either "ratio" or "mann-whitney")

## Details

Primer dimers are calculated by: 1) identifying fragments that occur within the expected primer sequence, 2) identifying which of these fragments is assayable, and 3) comparing the overall signal for primer peaks and peaks from the rest of the amplicon.

## Value

Returns a list containing primer dimer ratios or significance estimates (i.e. p-values) depending on the analytical method specified ("ratio" or "mann-whitney", respectively). Returns "NA" in cases where insufficient data is present to calculate primer dimer levels.

## Author(s)

Reid F. Thompson (<rthompso@aecom.yu.edu>), John M. Greally (<jgreally@aecom.yu.edu>)

## See Also

See Also [MassArrayData](#)

## Examples

```
data(MassArray.example.data)
primer.data <- estimatePrimerDimer(MassArray.example.data$fragments.T, MassArray.example.
mean(primer.data)
primer.data <- estimatePrimerDimer(MassArray.example.data$fragments.T, MassArray.example.
mean(primer.data)
```

---

 evaluateSNPs

*Evaluate SNPs*


---

### Description

Function to analyze a `MassArrayData` object for all potential single nucleotide polymorphisms (SNPs) indicated by new and/or missing peaks in the the spectral data for one or more samples

### Usage

```
evaluateSNPs(x, verbose = TRUE, plot = TRUE)
```

### Arguments

<code>x</code>	MassArrayData object containing spectral data for one or more samples
<code>verbose</code>	Logical specifying whether or not to display descriptive progress updates as SNPs are analyzed
<code>plot</code>	Logical specifying whether or not to display graphical representation of fragmentation profiles (default is TRUE)

### Details

This function performs an exhaustive search for all potential SNPs (single base pair substitutions or deletions) that may give rise to new and/or missing peaks. Graphical output is displayed by default, and extensive data describing putative SNPs is also returned.

Note that the graphical output does not contain a built-in legend at this time, but the plot may be interpreted as follows: In the uppermost panel the T-cleavage fragmentation profile is shown for a given amplicon (C-cleavage reactions occupy a split screen whenever relevant). CG dinucleotides (filled circles) are numbered and colored in blue. Other fragments are colored according to their ability to be assayed: fragment molecular weight outside the testable mass window (gray), fragment molecular weight overlapping with another fragment (red), fragment containing a potential conversion control (green), or fragment uniquely assayable but containing no CGs (black). Putative SNPs are shown directly below their location within the amplicon fragmentation profile. Each row represents analysis from a single sample. Small, gray symbols represent potential SNPs that do not have sufficient evidence (presence of a new peak with corresponding absence of an expected peak). Larger black symbols indicate a potential SNP with both new peaks and missing expected peaks. Triangles indicate base pair substitution, while circles indicate single base pair deletion.

### Value

Returns a list of potential SNPs for each identified new peak in the spectral data. Note that each new peak may be explained by any number of potential SNPs; the list returned only includes the most reliable hits, but the redundant nature of the data necessitates returning a nested list, such that each new peak is associated with the following list elements:

SNP	Contains a list of SNPs, each of which takes the form "position:base" where <code>position</code> is the base pair location within the amplicon sequence, and <code>base</code> is the mutated character
SNR	Contains a numerical list of signal-to-noise ratios corresponding to the expected original peak for the fragment mapping to the identified SNP position

fragment	Contains a numerical list of fragment IDs which map the SNP position to a specific fragment
SNP.quality	Contains a numerical list (values ranging from 0 to 2, with 0 being a highly unlikely SNP and 2 being a SNP with increased likelihood. This number is calculated as a function of new peak SNR and expected peak SNR.
samples	Contains a list of samples whose spectral data contained the given new peak
count	Specifies the number of unique SNP and sample pairs, exactly equivalent to the length of SNP, SNR, fragment, SNP.quality, or samples

**Author(s)**

Reid F. Thompson (<rthompso@aecon.yu.edu>), John M. Grealley (<jgrealley@aecon.yu.edu>)

**See Also**

See Also [identifySNPs](#)

**Examples**

```
data(MassArray.example.data)
SNP.data <- evaluateSNPs(MassArray.example.data[2,])
```

---

expandSequence      *Expand nucleotide sequence*

---

**Description**

Function to process shorthand form of a nucleotide sequence, where a given base pair followed by a number specifies a run of the indicated nucleotide for the specified length (ex: "A6TTCGA4")

**Usage**

```
expandSequence(sequence)
```

**Arguments**

sequence      Nucleotide sequence input as a character string

**Value**

Returns an expanded nucleotide sequence as a character string

**Author(s)**

Reid F. Thompson (<rthompso@aecon.yu.edu>), John M. Grealley (<jgrealley@aecon.yu.edu>)

**Examples**

```
expandSequence("A6TTCGA4")
expandSequence("C10C10")
expandSequence("AT1CG")
```

---

findCollisions      *Find peak collisions*

---

### Description

Function to determine which subset(s) of peaks collide with each other (i.e. molecular weights are indistinguishable given the specified resolution)

### Usage

```
findCollisions(peaks, resolution = 0.5)
```

### Arguments

peaks	Numerical list of molecular weights (in Da) corresponding to a set of peaks
resolution	Resolution (in Da), used to specify the ability to distinguish two different molecular weights. For a resolution of '0.5' (default), two molecular weights are considered identical if they are less than '0.5' Da apart.

### Value

Returns a list of peak collisions for each peak in the original list, thus the data object returned is in the form of a nested list.

### Author(s)

Reid F. Thompson (<rthompso@aecom.yu.edu>), John M. Greally (<jgreally@aecom.yu.edu>)

### Examples

```
findCollisions(1:5, 1.5)
```

---

findFragments      *Find fragments*

---

### Description

Function to identify which fragment(s) in a list of fragments match a given molecular weight

### Usage

```
findFragments(MW, fragments, resolution = 1)
```

### Arguments

MW	Molecular weight target (in Da)
fragments	List of molecular weights corresponding to unique fragments
resolution	Resolution (in Da), used to specify the ability to distinguish two different molecular weights. For a resolution of '1' (default), two molecular weights are considered identical if they are less than '1' Da apart.

**Value**

Returns the index or indices of fragment(s) within the input list that have a molecular weight which matches that specified as input

**Author(s)**

Reid F. Thompson (<rthompso@aecon.yu.edu>), John M. Grealley (<jgrealley@aecon.yu.edu>)

**Examples**

```
data(MassArray.example.data)
findFragments(3913,MassArray.example.data$fragments.T, resolution=0.1)
findFragments(3913,MassArray.example.data$fragments.T, resolution=0.5)
```

---

findPeaks

*Find peaks*

---

**Description**

Function to determine which peak(s) in a list of peaks match a given molecular weight.

**Usage**

```
findPeaks(MW, peaks, resolution = 1)
```

**Arguments**

MW	Molecular weight target (in Da)
peaks	List of molecular weights corresponding to unique peaks
resolution	Resolution (in Da), used to specify the ability to distinguish two different molecular weights. For a resolution of '1' (default), two molecular weights are considered identical if they are less than '1' Da apart.

**Value**

Returns the index or indices of peak(s) within the input list that have a molecular weight which matches that specified as input

**Author(s)**

Reid F. Thompson (<rthompso@aecon.yu.edu>), John M. Grealley (<jgrealley@aecon.yu.edu>)

**Examples**

```
findPeaks(3.1, 6:1, res=0)
findPeaks(3.1, 6:1, res=0.2)
```

---

identifySNPs      *Identify SNPs*

---

### Description

Function to identify potential single nucleotide polymorphisms (SNPs) which allow mapping of a novel peak sequence to the expected amplicon sequence

### Usage

```
identifySNPs(peak.sequence, sequence, rxn = c("T", "C"))
```

### Arguments

peak.sequence	Nucleotide sequence (can also be base composition - ex: "A6G2C1T3") as a character string
sequence	Nucleotide sequence for wildtype/expected amplicon as a character string
rxn	One of "T" or "C" specifying which cleavage reaction to use for SNP analysis

### Details

The algorithm steps through the `sequence`, substituting one nucleotide at a time with the other three base pairs or a blank character (deletion), in order to determine a base compositional match to the input `peak.sequence` which represents a peak not found in the native `sequence`.

### Value

Returns a list of potential SNP matches for the input `peak.sequence`. Each element of the list contains multiple items as follows:

sequence	corresponds to <code>peak.sequence</code>
position	corresponds to the matched position within <code>sequence</code>
base	corresponds to the altered nucleotide (i.e. "A", "T", "C", "G", or "")
type	corresponds to the class of SNP (i.e. "substitution" or "deletion")

### Author(s)

Reid F. Thompson (<rthompso@aeom.yu.edu>), John M. Greally (<jgreally@aeom.yu.edu>)

### Examples

```
## SINGLE SUBSTITUTION
identifySNPs("AAGT", "AATTTT")
## MULTIPLE SUBSTITUTION POSSIBILITIES
identifySNPs("A1G1T1", "AATTTT")
## DELETION
identifySNPs("AAT", "AGATTTT")
```

---

importEpiTyperData *Import EpiTyper data (v.1.0)*

---

**Description**

Function to read and import an EpiTyper datafile (v.1.0) and store it as a MassArraySpectrum objects

**Usage**

```
importEpiTyperData(data, MassArrayObject, verbose = TRUE)
```

**Arguments**

data	location of EpiTyper datafile as a character string
MassArrayObject	Pre-existent MassArrayData object in which store relevant sample and spectral information from datafile
verbose	Logical specifying whether or not to display descriptive progress updates as datafile is processed

**Details**

EpiTyper v.1.0 datafiles must only contain a single amplicon, thus the user must export peak data for one amplicon at a time.

**Value**

Returns a list of MassArraySpectrum objects each populated by spectral data

**Author(s)**

Reid F. Thompson (<rthompso@aecon.yu.edu>), John M. Grealley (<jgrealley@aecon.yu.edu>)

**See Also**

See Also [MassArraySpectrum](#)

---

importEpiTyperData.new  
*Import EpiTyper data (v.1.0.5)*

---

**Description**

Function to read and import an EpiTyper datafile (v.1.0.5) and store it as a list of MassArraySpectrum objects

**Usage**

```
importEpiTyperData.new(data, MassArrayObject, verbose = TRUE)
```

**Arguments**

data	location of EpiTyper datafile as a character string
MassArrayObject	Pre-existent MassArrayData object in which store relevant sample and spectral information from datafile
verbose	Logical specifying whether or not to display descriptive progress updates as datafile is processed

**Details**

EpiTyper v.1.0.5 datafiles must only contain a single amplicon, thus the user must export peak data for one amplicon at a time.

**Value**

Returns a list of MassArraySpectrum objects each populated by spectral data

**Author(s)**

Reid F. Thompson (<rthompso@aeom.yu.edu>), John M. Grealley (<jgreally@aeom.yu.edu>)

**See Also**

See Also [MassArraySpectrum](#)

---

inSilicoFragmentation

*In silico fragmentation*

---

**Description**

Function to perform a complete in silico fragmentation of input sequence. Provides wrapper to a number of different functions, each of which determines additional information about each fragment.

**Usage**

```
inSilicoFragmentation(sequence, fwd.tag = "", rev.tag = "", type = c("T", "C"),
```

**Arguments**

sequence	Nucleotide sequence input as a character string
fwd.tag	Nucleotide tag sequence 5' of the forward primer
rev.tag	T7-containing nucleotide tag sequence 5' of the reverse primer
type	One of 'T' or 'C' indicating which cleavage reaction to use
lower.threshold	Lower limit (in Da) of usable mass window (default: '1500')
upper.threshold	Upper limit (in Da) of usable mass window (default: '7000')



`fwd.primers`      Length (in bp) of forward primer  
`rev.primers`      Length (in bp) of reverse primer  
`multiple.conversion`  
                          Logical value specifying whether or not to include multiple CGs on the same  
                          conversion control fragment where possible (default is FALSE).

### Details

In silico fragmentation analysis includes RNase A digestion, peak mapping and overlap detection, CG detection, assayability and conversion controls.

### Value

Returns a list of `MassArrayFragment` objects, each with extensive contextual and other information

### Author(s)

Reid F. Thompson (<rthompso@aecon.yu.edu>), John M. Grealley (<jgrealley@aecon.yu.edu>)

### See Also

See Also as [MassArrayFragment](#)

### Examples

```
inSilicoFragmentation("GGGTTAGTCC")
```

---

<code>isAssayable</code>	<i>Is assayable?</i>
--------------------------	----------------------

---

### Description

Function to determine whether or not a given molecular weight is assayable (i.e. within the usable mass window specified)

### Usage

```
isAssayable(MW, lower.threshold = 1500, upper.threshold = 7000)
```

### Arguments

`MW`                      Numerical input corresponding to molecular weight  
`lower.threshold`  
                          Lower limit (in Da) of usable mass window (default: '1500')  
`upper.threshold`  
                          Upper limit (in Da) of usable mass window (default: '7000')

### Value

Returns a logical corresponding to whether or not the molecular weight input falls within the usable mass window specified

**Author(s)**

Reid F. Thompson (<rthompso@aeacom.yu.edu>), John M. Greally (<jgreally@aeacom.yu.edu>)

**Examples**

```
isAssayable(5000)
isAssayable(1200)
```

---

numCollisions	<i>Count peak collisions</i>
---------------	------------------------------

---

**Description**

Function to count the number of peak collisions (i.e. molecular weights are indistinguishable given the specified resolution)

**Usage**

```
numCollisions(peaks, resolution = 0.5)
```

**Arguments**

peaks	Numerical list of molecular weights (in Da) corresponding to a set of peaks
resolution	Resolution (in Da), used to specify the ability to distinguish two different molecular weights. For a resolution of '0.5' (default), two molecular weights are considered identical if they are less than '0.5' Da apart.

**Value**

Returns a list of peak collision counts for each peak in the original list.

**Author(s)**

Reid F. Thompson (<rthompso@aeacom.yu.edu>), John M. Greally (<jgreally@aeacom.yu.edu>)

**Examples**

```
numCollisions(1:5, 1.5)
```

---

plot.MassArrayData *Plot MassArrayData*

---

### Description

Function to generate graphical output for methylation data in a `MassArrayData` object

### Usage

```
## S3 method for class 'MassArrayData'  
plot(x, ..., collapse = TRUE, bars = TRUE, scale = TRUE, sequence = TRUE, labels = TRUE)
```

### Arguments

<code>x</code>	Object of class <code>MassArrayData</code>
<code>...</code>	Other arguments to <code>plot</code> , currently not supported at this time
<code>collapse</code>	Logical specifying whether or not to combine samples by unique group (see <a href="#">MassArrayData</a> ). If <code>TRUE</code> , each methylation values are averaged across all samples in each unique group. If <code>FALSE</code> , all samples are retained and plotted individually
<code>bars</code>	Logical specifying whether or not to display error bars. If <code>TRUE</code> (and <code>collapse</code> is <code>TRUE</code> ), the median absolute deviation is calculated for each group and plotted as an error bar for each methylation value. If <code>FALSE</code> , no error bars are displayed
<code>scale</code>	Logical specifying whether or not to keep the x axis to scale. If <code>TRUE</code> , methylation values are plotted as a function of relative position within the amplicon sequence. If <code>FALSE</code> , positional information is ignored and methylation values are evenly spaced across the plot window.
<code>sequence</code>	Logical specifying whether or not to display the nucleotide sequence for the amplicon
<code>labels</code>	Logical specifying whether or not to display data labels
<code>colors</code>	Logical specifying whether or not to plot in color. If <code>TRUE</code> , colors are used. If <code>FALSE</code> , plotting occurs in black and white and grayscale.
<code>main</code>	Label/title for overall plot (default is " ")
<code>width</code>	Numerical value specifying the display width to use for each methylation value; number corresponds to the number of base pairs to include in both directions from the methylation position (default is 1.5)

### Author(s)

Reid F. Thompson (<rthompso@aecon.yu.edu>), John M. Greally (<jgreally@aecon.yu.edu>)

### See Also

See Also [MassArrayData](#)

### Examples

```
data(MassArray.example.data)  
plot(MassArray.example.data, collapse=FALSE, bars=FALSE, scale=FALSE)
```

---

position-methods     *Operate on positional information (methods)*

---

### Description

Methods to access (and/or assign) positional information for a `MassArrayData` object

### Methods

**object = "MassArrayData"** Access positional information for `MassArrayData` object

**object = "MassArrayData", value = "missing"** Handle empty function call, simply return the `MassArrayData` object

**object = "MassArrayData", value = "character"** Assign position of `MassArrayData` object to `value`

### See Also

[position](#)

---

position     *Operate on positional information*

---

### Description

Function to access (and/or assign) positional information for a `MassArrayData` object

### Usage

```
position(object)
position(object) <- value
```

### Arguments

<code>object</code>	Object of class <code>MassArrayData</code>
<code>value</code>	Character string containing positional information of the form "chrXX:XXXX-XXXX"

### Value

Returns a character string containing positional information of the form "chrXX:XXXX-XXXX" if accessing a `MassArrayData` object. If updating a `MassArrayData` object, function returns the object with updated positional information

### Author(s)

Reid F. Thompson (<rthompso@aecom.yu.edu>), John M. Grealley (<jgrealley@aecom.yu.edu>)

### See Also

See Also [MassArrayData](#)

## Examples

```
data(MassArray.example.data)
position(MassArray.example.data)
position(MassArray.example.data) <- "chrB:2001-2374"
position(MassArray.example.data)
```

---

revComplement-methods  
*Reverse complement (methods)*

---

## Description

Methods for reverse complement

## Methods

**x = "MassArrayData"** Finds reverse complement of a MassArrayData object, a function that consists of altering sequence, strand, fragmentation, and methylation data

**x = "character"** Calculates reverse complement from nucleotide sequence as character input

## Author(s)

Reid F. Thompson (<rthompso@aeom.yu.edu>), John M. Grealley (<jgrealley@aeom.yu.edu>)

## See Also

[revComplement](#)

---

revComplement      *Reverse complement*

---

## Description

Function to find the reverse complement

## Usage

```
revComplement(x)
```

## Arguments

**x**                      sequence input to use for reverse complement. **x** can be a character string or a MassArrayData object.

## Value

Returns the reverse complement of a character string or MassArrayData object, depending upon input data type.

**Author(s)**

Reid F. Thompson (<rthompso@aeom.yu.edu>), John M. Grealley (<jgrealley@aeom.yu.edu>)

**See Also**

[revComplement-methods](#)

**Examples**

```
revComplement("AATCCGGGGGAA")
```

---

rnaDigest

*RNA digest*

---

**Description**

Function to perform an in silico RNase A digest for either the T or C cleavage reactions

**Usage**

```
rnaDigest(sequence, type = c("T", "C"))
```

**Arguments**

sequence	Nucleotide sequence input
type	One of either 'T' or 'C', specifying which cleavage reaction mixture was used

**Value**

Returns a list of `MassArrayFragment` objects, each containing information about a given fragment generated by the RNA digest

**Author(s)**

Reid F. Thompson (<rthompso@aeom.yu.edu>), John M. Grealley (<jgrealley@aeom.yu.edu>)

**See Also**

See Also as [MassArrayFragment](#)

**Examples**

```
rnaDigest("AAAACCCCTGCGGAGAGAGGCCGACAAAA", type="T")
```

---

samples-methods      *Operate on sample name(s) (methods)*

---

**Description**

Methods to access (and/or assign) sample name information for a `MassArrayData` object

**Methods**

**object = "MassArrayData"** Access sample name information for `MassArrayData` object

**object = "MassArrayData", value = "missing"** Handle empty function call, simply return the `MassArrayData` object

**object = "MassArrayData", value = "character"** Assign sample name of `MassArrayData` object to `value`

**See Also**

[samples](#)

---

samples      *Operate on sample name(s)*

---

**Description**

Function to access (and/or assign) sample name information for a `MassArrayData` object

**Usage**

```
samples(object)
samples(object) <- value
```

**Arguments**

<code>object</code>	Object of class <code>MassArrayData</code>
<code>value</code>	List of character strings containing sample name information, one for each sample contained in the <code>MassArrayData</code> object

**Value**

Returns a list of character strings containing sample name information for each sample in `MassArrayData` object. If updating a `MassArrayData` object, function returns the object with updated sample name information

**Author(s)**

Reid F. Thompson (<rthompso@aecom.yu.edu>), John M. Grealley (<jgrealley@aecom.yu.edu>)

**See Also**

See Also as [MassArrayData](#)

**Examples**

```
data(MassArray.example.data)
samples(MassArray.example.data)
samples(MassArray.example.data)[2] <- "SECOND"
samples(MassArray.example.data)
```

---

```
sum.MassArraySpectrum
```

*Sum MassArraySpectrum objects*

---

**Description**

Function to collapse multiple MassArraySpectrum objects into a single MassArraySpectrum representing the sum of each

**Usage**

```
## S3 method for class 'MassArraySpectrum'
sum(x, ..., trim = 0, na.rm = TRUE)
```

**Arguments**

x	One or multiple MassArraySpectrum objects to include in sum
...	Any additional MassArraySpectrum objects to include in sum
trim	Numerical value between 0 and 0.5 specifying the proportion of spectra to remove from consideration on a per peak basis, such that the SNR of each peak is calculated as the trimmed mean of the same peak across all included spectra.
na.rm	Logical value passed to mean, indicating whether NA values should be stripped before the computation proceeds.

**Value**

Returns a single MassArraySpectrum object that represents the union of all unique peaks from the component MassArraySpectrum objects, with SNR for each peak representing the average value of that peak across all spectra

**Author(s)**

Reid F. Thompson (<rthomps@aecon.yu.edu>), John M. Greally (<jgreally@aecon.yu.edu>)

**See Also**

See Also as [MassArraySpectrum](#)

**Examples**

```
data(MassArray.example.data)
MassArray.example.data$samples[[1]]$peaks[[11]]$height
MassArray.example.data$samples[[1]] <- sum.MassArraySpectrum(MassArray.example.data$samp
MassArray.example.data$samples[[1]]$peaks[[11]]$height
```



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