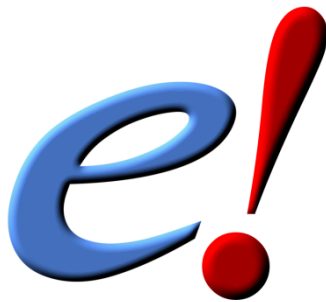


Browsing Genomes with Ensembl



www.ensembl.org
www.ensemblgenomes.org

Coursebook v72

<http://tinyurl.com/Camb0613>

Cambridge - 21st June 2013

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Introduction to Ensembl

Getting started with Ensembl

www.ensembl.org

Ensembl is a joint project between the EBI ([European Bioinformatics Institute](http://www.ebi.ac.uk)) and the [Wellcome Trust Sanger Institute](http://www.wellcome-trust.org) that annotates **chordate** genomes (i.e. vertebrates and closely related invertebrates with a notochord such as sea squirt). Gene sets from model organisms such as yeast and worm are also imported for comparative analysis by the Ensembl 'compara' team. Most annotation is updated every two months, leading to increasing Ensembl versions (such as version 70), however the gene sets are determined less frequently. A sister browser at www.ensemblgenomes.org is set up to access non-chordates, namely bacteria, plants, fungi, metazoa, and protists.

Ensembl provides genes and other **annotation** such as regulatory regions, conserved base pairs across species, and sequence variations. The Ensembl gene set is based on protein and mRNA evidence in **UniProtKB** and **NCBI RefSeq** databases, along with manual annotation from the **VEGA/Havana** group. All the data are freely available and can be accessed via the web browser at www.ensembl.org. Perl programmers can directly access Ensembl databases through an Application Programming Interfaces (**Perl APIs**). Gene sequences can be downloaded from the Ensembl browser itself, or through the use of the **BioMart** web interface, which can extract information from the Ensembl databases without the need for programming knowledge by the user.

Synopsis – What can I do with Ensembl?

- View genes with other annotation along the chromosome.
- View alternative transcripts (i.e. splice variants) for a given gene.
- Explore homologues and phylogenetic trees across more than 60 species for any gene.
- Compare whole genome alignments and conserved regions across species.
- View microarray sequences that match to Ensembl genes.
- View ESTs, clones, mRNA and proteins for any chromosomal region.
- Examine single nucleotide polymorphisms (SNPs) for a gene or chromosomal region.
- View SNPs across strains (rat, mouse), populations (human), or breeds (dog).
- View positions and sequence of mRNAs and proteins that align with Ensembl genes.
- Upload your own data.
- Use BLAST, or BLAT against any Ensembl genome.
- Export sequence or create a table of gene information with BioMart.
- Determine how your variants affect genes and transcripts using the Variant Effect Predictor.
- Share Ensembl views with your colleagues and collaborators.

Need more help?

- ❓ Check Ensembl [documentation](#)
- ❓ Watch [video tutorials](#) on YouTube
- ❓ View the [FAQs](#)
- ❓ Try some [exercises](#)
- ❓ Read some [publications](#)
- ❓ Go to our [online course](#)

Stay in touch!

- ❖ [Email](#) the team with comments or questions at helpdesk@ensembl.org
- ❖ Follow the Ensembl [blog](#)
- ❖ Sign up to a [mailing list](#)

Further reading

Flicek, P. *et al*
Ensembl 2013
Nucleic Acids Res. Advanced Access (Database Issue)
<http://www.ncbi.nlm.nih.gov/pubmed/23203987>

Ensembl Methods Series
<http://www.biomedcentral.com/series/ENSEMBL2010>

Xosé M. Fernández-Suárez and Michael K. Schuster
Using the Ensembl Genome Server to Browse Genomic Sequence Data.
UNIT 1.15 in Current Protocols in Bioinformatics, Jun 2010.

Giulietta M Spudich and Xosé M Fernández-Suárez
Touring Ensembl: A practical guide to genome browsing
BMC Genomics 2010, 11:295 (11 May 2010)

Exploring the Ensembl genome browser

Demo: Ensembl species

The front page of Ensembl is found at ensembl.org. It contains lots of information and links to help you navigate Ensembl:

The screenshot shows the Ensembl homepage with several callout boxes:

- Link back to homepage**: Points to the Ensembl logo in the top left navigation bar.
- Ensembl tools**: Points to the 'Tools' link in the top navigation bar.
- Blue bar remains visible on every Ensembl page**: Points to the dark blue navigation bar at the top of the page.
- Search**: Points to the search bar in the top right corner.
- Search**: Points to the search bar in the main content area.
- Drop-down list of species**: Points to the 'All species' dropdown menu in the main search area.
- News**: Points to the 'What's New in Release 71' section on the right side of the page.

Click on [View full list of all Ensembl species](#).

Click on the common name of your species of interest to go to the species homepage. We'll click on [Human](#).

Search

Information and statistics

News

Links to example features in Ensembl

To find out more about the genome assembly and genebuild, click on **More information and statistics**.

Information

Tables of statistics

Statistics Summary	
Assembly:	GRCh37.p10, Feb 2009
Database version:	71.37
Base Pairs:	3,320,602,131
Golden Path Length:	3,101,804,739
Genebuild by:	Ensembl
Genebuild method:	Full genebuild
Genebuild started:	Jul 2010
Genebuild released:	Apr 2011
Genebuild last updated/patched:	Feb 2013

Let's take a look at the Ensembl Genomes homepage at ensemblgenomes.org.

Links to the tax-specific sites

Link back to Ensembl

News

Ensembl Genomes: Extending Ensembl across the taxonomic space.

NEW! in EnsemblBacteria

Over 6000 bacterial genome sequences have been annotated and deposited in the public archives of the members of the [International Nucleotide Sequence Database Collaboration](#). This site provides access to complete, annotated bacterial genomes (present in the [European Nucleotide Archive](#)) through the Ensembl graphical user interface (genome browser). More details about the integration are provided [here](#)

Programmatic access is available through the Ensembl Perl and REST-ful APIs and through publicly accessible mysql databases, along with full data dumps (including DNA sequence and protein sequence in FASTA format, annotations in GTF format, and mysql dump files). Due to the large number of these databases, there is some modification to the APIs, and database and FTP site structure, compared to that used for other branches of the taxonomy (e.g. the storage of many genomes in one database; the provision of lookup services to identify genomes by INSDC identifiers, taxonomy identifiers, or partial names). [Full details are available here.](#)

BioMart access is not available, but we are working on providing new, more powerful data mining tools to allow users to exploit these genomes. A selection of over 100 key bacterial genomes has been included in the pan-taxonomic Compara, and genes from all genomes are classified into families using HAMAP and PANTHER ([more details](#))

Ensembl Bacteria has been completely revised, and now contains more than 6000 genomes that have been deposited in the European Nucleotide Archive [Find out more...](#)

Hordeum vulgare (Barley) variation **NEW!**

Wheat sequence search **NEW!**

GO annotation extensions **NEW!**

Highlighting annotations in gene trees

Click on the different taxa to see their homepages. Each one is colour-coded.

EnsemblProtists

Search: All species (e.g. Pfl02120 or cypr)

Popular genomes

- Plasmodium falciparum
- Dicotylestium discoidium
- Physiphthora infestans
- Leishmania major

What's new in Release 17 (January 2013)

- Updated schema
- Added new species *Glandia lamella*
- Protein features updated for all the protist species
- Minor internal fixes for the comparative genomics database
- Updated BioMart

Protists

EnsemblFungi

Search: All species (e.g. MAT2 or abobob)

Popular genomes

- Saccharomyces cerevisiae
- Schizosaccharomyces pombe
- Aspergillus nidulans
- Ustilago maydis
- Magnaporthe oryzae
- Zygomorpha tritici

What's new in Release 17 (February 2013)

- New genomes
 - Glomerella perniciosa*
 - Ascochyta blight*
 - Zygomorpha tritici*
 - Glomerella graminicola*
- Updated genomes
 - Schizosaccharomyces pombe* (latest data from EuroBall)
- New data
 - Glomerella graminicola* EST alignments
 - Magnaporthe oryzae* EST alignments
 - Leishmania major* EST alignments
 - DNA comparative alignments of *Pyrenophora sp.*
- Updated data
 - Updated peptide comparative genomics
 - Updated BioMarts

Fungi

Metazoa

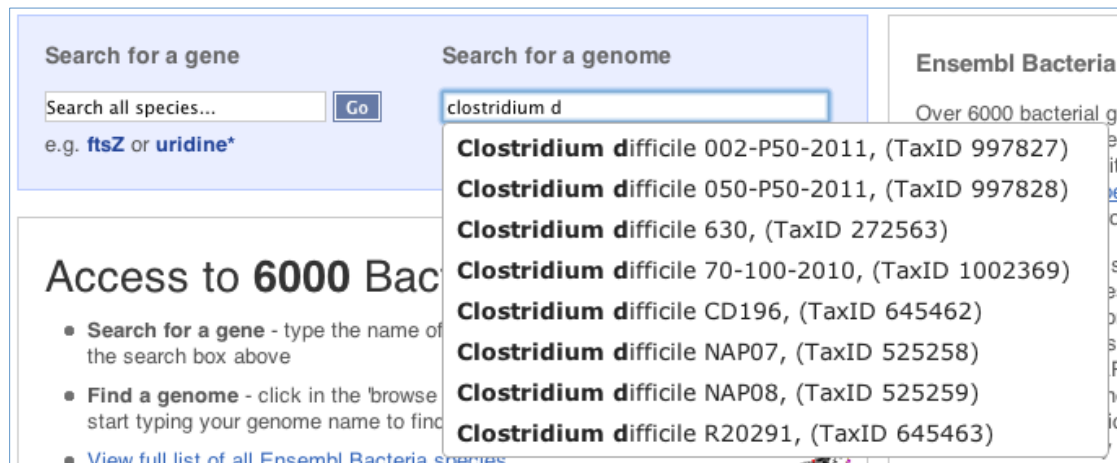
Plants

Bacteria

You can navigate most of the taxa in the same way as you would with Ensembl, but Ensembl Bacteria has a large number of genomes, so needs slightly different methods. Let's look at it in more detail.

There's no full species list for bacteria as it would be hard to navigate with the number of species. To find a species, start to type the species name into the species search box. A drop down list will appear with possible species.

For example, to find a substring of *Clostridium difficile* type in **Clostridium d.**



The drop down contains various strains of *Clostridium difficile*. Let's choose **Clostridium difficile 630**. This will take us to another species homepage, where we can explore various features.



Exercises: Ensembl species

Exercise 1 – Panda

(a) Go to the species homepage for Panda. What is the name of the genome assembly for Panda?

(b) Click on [More information and statistics](#). How long is the Panda genome (in bp)? How many genes have been annotated?

Exercise 2 – Zebrafish

(a) What's new in release 71 for zebrafish?

(b) What previous release is available for zebrafish?

Exercise 3 – Mosquitos

(a) Go to Ensembl Metazoa. How many species of the genus *Anopheles* are there?

(b) Who published the genome sequence for *Anopheles gambiae*?

Exercise 4 – Bacteria

Go to Ensembl Bacteria and find the species *Belliella baltica*. How many coding and non-coding genes does it have?

Demo: The Region in detail view


Start at the Ensembl front page, ensembl.org. You can search for a region by typing it into a search box, but you have to specify the species.

Type (or copy and paste) [human 4:123792818-123867893](#) into either search box.

Search: for
 e.g. BRCA2 or rat X:100000..200000 or coronary heart disease

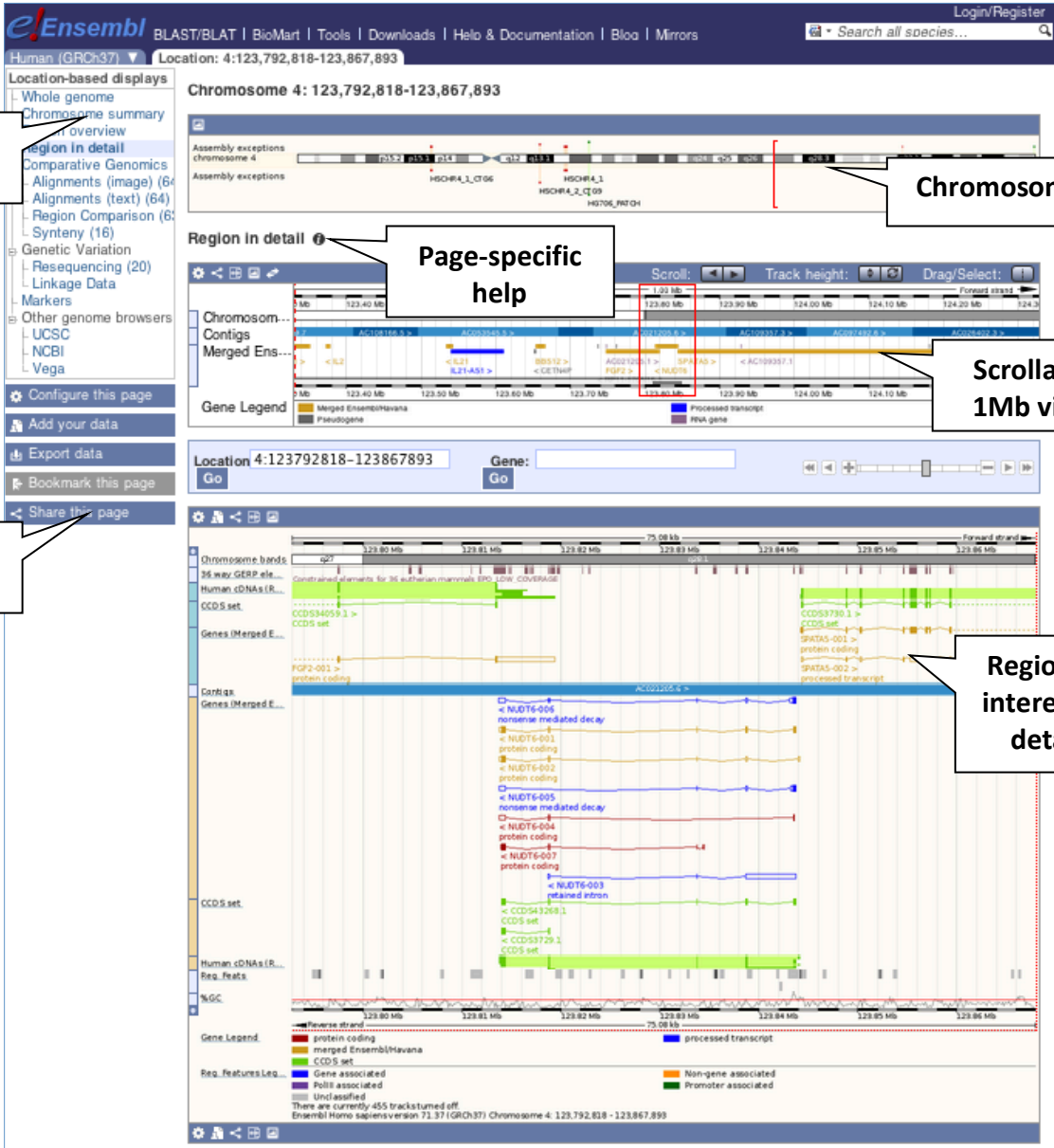
or

Press **Enter** or click **Go** to jump directly to the **Region in detail** Page.

Click on the button  to view page-specific help.

The help pages provide links to **Frequently Asked Questions**, a **Glossary**, **Video Tutorials**, and a form to **Contact HelpDesk**.

There is a help video on this page at <http://youtu.be/tTKEvgPUq94>.

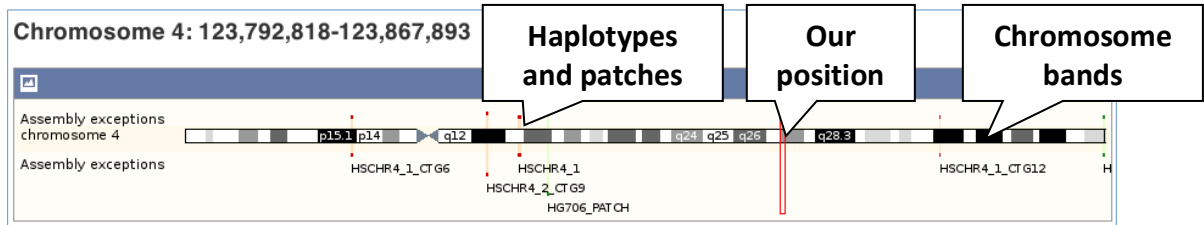


The screenshot shows the Ensembl genome browser interface for Human (GRCh37) at location 4:123,792,818-123,867,893. The interface is annotated with several callouts:

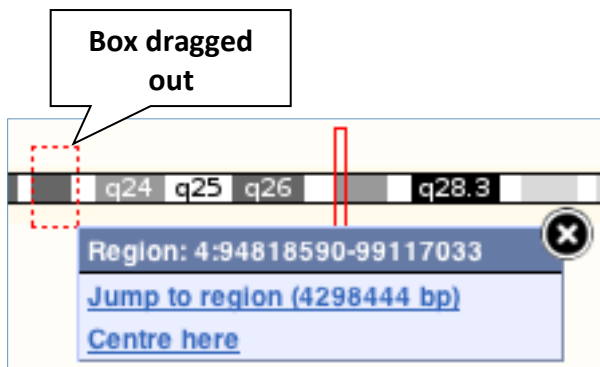
- Location views:** Points to the left-hand navigation menu containing options like 'Whole genome', 'Chromosome summary', 'region in detail', 'Comparative Genomics', 'Alignments (image) (64)', 'Alignments (text) (64)', 'Region Comparison (6)', 'Synteny (16)', 'Genetic Variation', 'Resequencing (20)', 'Linkage Data', 'Markers', and 'Other genome browsers' (UCSC, NCBI, Vega).
- Chromosome:** Points to the top track showing the assembly exceptions for chromosome 4, with genes HSOX4_1_CT06, HSOX4_1, HSOX4_2_CT09, and HSOX4_WT04 visible.
- Page-specific help:** Points to the 'Region in detail' button in the top navigation bar.
- Scrollable 1Mb view:** Points to the 'Region in detail' track, which shows a zoomed-in view of the region with a scroll bar and track height controls.
- Tool buttons:** Points to the bottom-left navigation icons for home, back, forward, and search.
- Region of interest in detail:** Points to a zoomed-in view of a specific gene region, showing tracks for 'Chromosome bands', '35-way GERP etc.', 'Human cDNAs (RefSeq)', 'CCDS set', 'Genes (Merged Ensembl)', 'Contigs', 'Genes (Merged Ensembl)', 'CCDS set', 'Human cDNAs (RefSeq)', 'Req. Feats.', and '%GC'. The gene legend at the bottom identifies features like protein coding, merged Ensembl/Havana, CCDS set, Gene associated, PolII associated, Unclassified, Non-gene associated, and Promoter associated.

The Region in detail page is made up of three images, let's look at each one on detail.

The first image shows the chromosome:

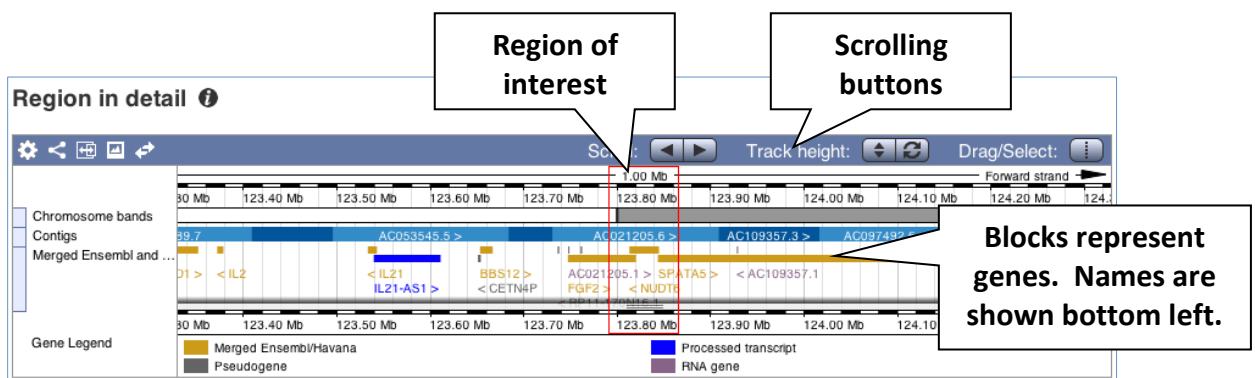



You can jump to a different region by dragging out a box in this image. Drag out a box on the chromosome, a pop-up menu will appear.



If you wanted to move to the region, you could click on [Jump to region \(###bp\)](#). For now, we'll close the pop-up by clicking on the X on the corner.


The second image shows a 1Mb region around our selected region. This view allows you to scroll back and forth along the chromosome.

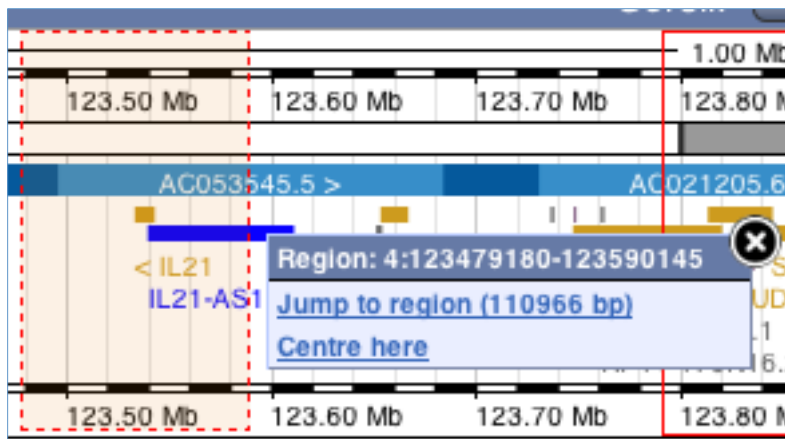


At the moment the gene track is set to a fixed height. Click on the [Automatic track height button](#)  to expand the image to include all possible data in the track.

Scroll along the chromosome by clicking and dragging within the image. As you do this you'll see the image below grey out and two blue buttons appear. Clicking on [Update this image](#) would jump the lower image to the region central to the scrollable image. We want to go back to where we started, so we'll click on [Reset scrollable image](#).

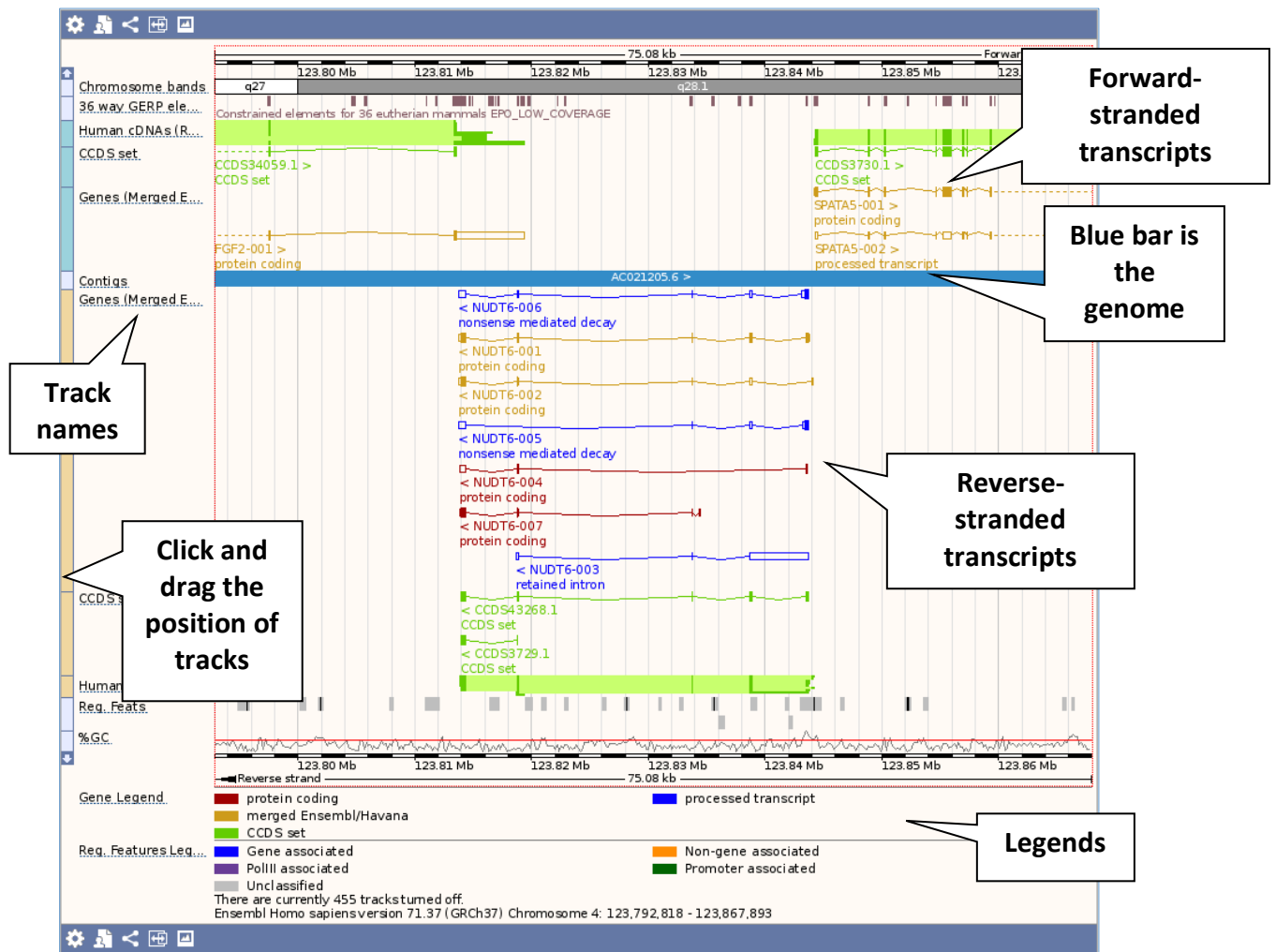


You can also drag out and jump to a region. Either hold down **shift** and drag in the image, or click on the [Drag/Select button](#)  to change the action of your mouse click, and drag out a box.



Click on the **X** to close the pop-up menu.

The third image is a detailed, configurable view of the region.

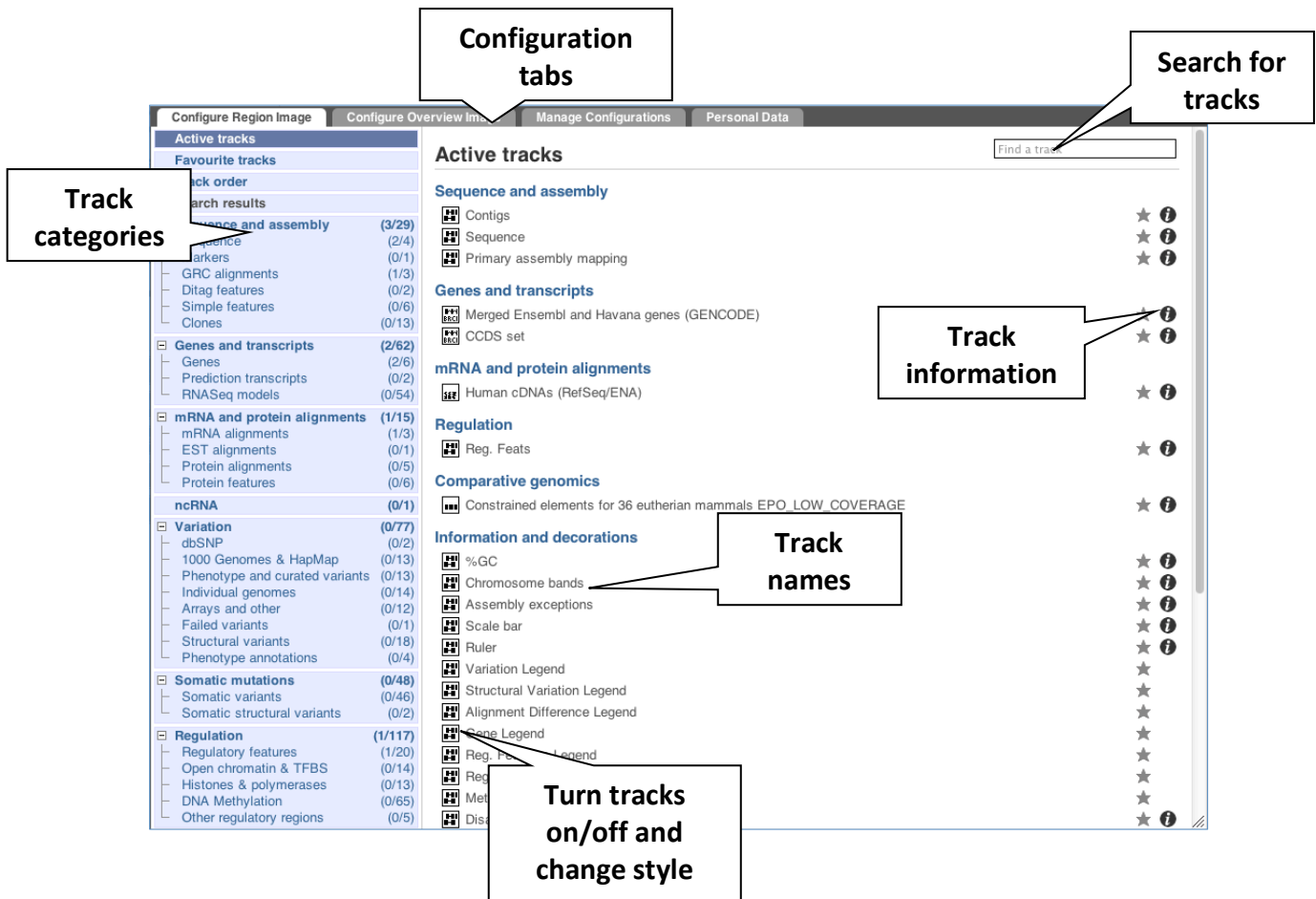


We can edit what we see on this page by clicking on the blue **Configure this page** menu at the left.



This will open a menu that allows you to change the image.

You can put some tracks on in different styles; more details are in this FAQ: <http://www.ensembl.org/Help/Faq?id=335>.

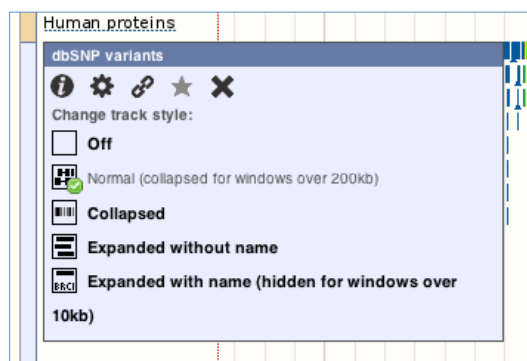


Let's add some tracks to this image. Add:

- Human proteins – Labels
- dbSNP variants – Normal
- 1000 Genomes – AMR – Collapsed

Now click on the tick in the top left hand to save and close the menu. Alternatively, click anywhere outside of the menu. We can now see the tracks in the image.

We can also change the way the tracks appear by hovering over the track name then the cog wheel to open a menu. We can move tracks around by clicking and dragging on the bar to the left of the track name.



Now that you've got the view how you want it, you might like to show something you've found to a colleague or collaborator. Click on the [Share this page](#) button to generate a link. Email the link to someone else, so that they can see the same view as you, including all the tracks you've added. These links contain the Ensembl release number, so if a new release or even assembly comes out, your link will just take you to the archive site for the release it was made on.



To return this to the default view, go to [Configure this page](#) and select [Reset configuration](#) at the bottom of the menu.

Exercises: The Region in Detail view

Exercise 5 - Exploring a genomic region in human

(a) Go to the region from 32,448,000 to 33,198,000 bp on human chromosome 13. On which cytogenetic band is this region located? How many contigs make up this portion of the assembly (contigs are contiguous stretches of DNA sequence that have been assembled solely based on direct sequencing information)?

(b) Zoom in on the *BRCA2* gene.

(c) Are there any Tilepath clones that contain the complete *BRCA2* gene?

(d) Create a [Share](#) link for this display. Email it to yourself and open the link.

(e) Export the genomic sequence of the region you are looking at in FASTA format.

(f) Turn off all tracks you added to the [Region in detail](#) page.

Exercise 6 – Exploring patches and haplotypes in human

(a) Go to the region 6:112294691-112624977 in human. What is the green highlighted region? (Tip: you can search for help terms in the Ensembl search boxes.)

(b) Can you see the patches in the chromosome view? Drag out a box to jump to a region containing the patch labelled HG27_patch. What are the coordinates of the patch?

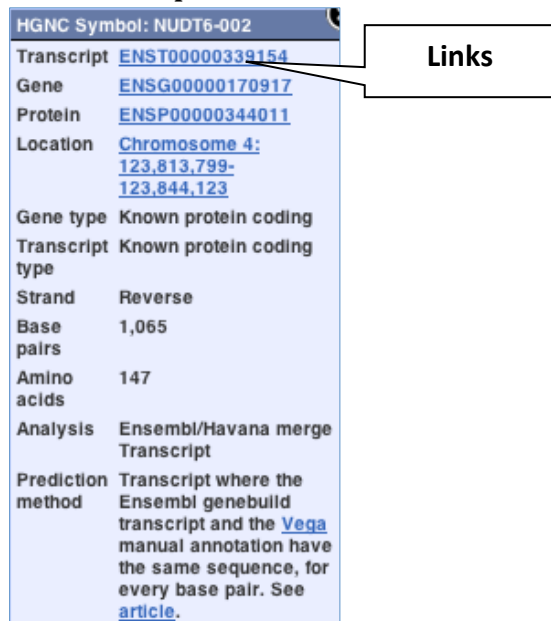
(c) Can you compare this patch with the reference? What has changed between this patch and the sequence it replaced?

(d) Go back to the previous view and scroll to the right in the 1Mb view until you reach a red highlighted region. What is this?

Genes and transcripts

Demo: The gene tab

If you click on any one of the transcripts in the Region in detail image, a pop-up menu will appear, allowing you to jump directly to that gene or transcript.



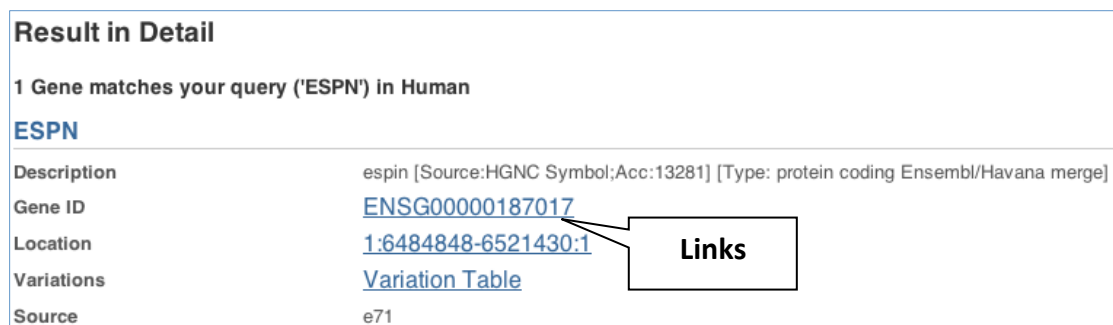
HGNC Symbol: NUDT6-002

Transcript	ENST00000339154
Gene	ENSG00000170917
Protein	ENSP00000344011
Location	Chromosome 4: 123,813,799-123,844,123
Gene type	Known protein coding
Transcript type	Known protein coding type
Strand	Reverse
Base pairs	1,065
Amino acids	147
Analysis	Ensembl/Havana merge Transcript
Prediction method	Transcript where the Ensembl genebuild transcript and the Vega manual annotation have the same sequence, for every base pair. See article .

Another way to go to a gene of interest is to search directly for it.

We're going to look at the human *ESPN* gene. This gene encodes a multifunctional actin-bundling protein with a major role in mediating sensory transduction in various mechanosensory and chemosensory cells. Mutations in this gene are associated with deafness (<http://tinyurl.com/espn-ncbi-gene>).

From ensembl.org, type *ESPN* into the search bar and click the Go button. Click on Gene and Human to find the hits.



Result in Detail

1 Gene matches your query ('ESPN') in Human

ESPN

Description	espin [Source:HGNC Symbol;Acc:13281] [Type: protein coding Ensembl/Havana merge]
Gene ID	ENSG00000187017
Location	1:6484848-6521430:1
Variations	Variation Table
Source	e71

Click on the **gene name** or **Ensembl ID**. The **Gene tab** should open:

Gene tab

Option: Open table of transcripts

ESPN-001 transcript. Click for info

Blue bar is the genome

Forward-stranded transcripts

Reverse-stranded transcripts

Gene views

Forward-stranded transcripts

Reverse-stranded transcripts

Gene summary

Gene: ESPN ENSG00000187017

Description: [ESPN](#) [Source:HGNC Symbol;Acc:13281]

Location: [Chromosome 1: 6,484,848-6,521,430](#) forward strand.

INSDC coordinates: chromosome:GRCh37:CM000663.1:6484848:6521430:1

Transcripts: This gene has 10 transcripts (splice variants) [Show transcript table](#)

Gene summary

Name: [ESPN](#) (HGNC Symbol)

Synonyms: DFNB36 [To view all Ensembl genes linked to the name [click here](#).]

CCDS: This gene is a member of the Human CCDS set: [CCDS70](#)

Ensembl version: ENSG00000187017.10

Gene type: Known protein coding

Prediction Method: Annotation for this gene includes both automatic annotation from Ensembl and [Havana](#) manual curation, see [article](#).

Alternative genes: This gene corresponds to the following database identifiers:
Havana gene: [OTTHUMG00000000753](#) (version 5)

Go to [Region in Detail](#) for more tracks and navigation options (e.g. zooming)

Genes (Merged E...)

Contigs

Genes (Merged E...)

Let's walk through some of the links in the left hand navigation column. How can we view the genomic sequence? Click [Sequence](#) at the left of the page.

Human (GRCh37) ▾

Gene-based displays

- Gene summary
- Splice variants (10)
- Transcript comparison
- Supporting evidence
- Sequence
- External references
- Regulation
- Expression

Most recent human genome assembly
GRCh37 = hg19

Click Sequence

Marked-up sequence ⓘ

Key

Exons All exons in this region ESPN exons

```
>chromosome:GRCh37:1:6484248:6522030:1
AGCGCACTAGTGGTTCCTGCTCCTGCTCCTGCCCTCCCCGCTGGAACCTCTG
TTCTCGGATCTGGAGGGACCCTGGAAGGCAGGGCTCTTTGCAATCTCCGGGGAT
CCAGAGCCCTTCAGGGACGTGGCAGGGCTGCTCCTGCCTCAGGGCCGTTGTCT
CCTCACCCCGCCTGGAATACCCTTCTCGCCGCTCAAACCCAGCCCCACGGCACCTCCTCA
GAGACCTTTCCTGTCCGCCACGCGGTCCCGACAATCACATCCCACATCACCTCTGGAATT
GCGTCGCGGGCGCCTGGAACCGCAGTTAGCGGGCACTGGGCAGATGAATGAATT
TGCCTGGACGGCTCTCCAATTCGAACCCAGTTTTGTGCTCCTGCGGGTCTCA
CGTGAGGCAAATTAGGAGAGAAGCCCTGGGCACCTTGCCCCAGTCGCACGAGT
GCGTCGCGGGCGGGGGCGGGCGGGGAACCTCGGGCGGAGGCTGCGGGGCGGGGCGGGGCGGG
GTGGGGGCGGGCCGAGTCTTAAGCCGGCGTCCGCGGGCTCCGGCCCCAGAGCGGGCGG
AGCGGAGCGCCAGGCAGCGCGGAGCGGAGGCCAGGCCACAGCCGCTCCGCTCCCGGCC
CGCAGATCCCCGACGGCCGACCGCGGGCTCCTCTGGCCCAGCAAGAACAGTGCATGGCG
TCCTGGGGAAGGCGCTGAGTGCAGGAGTCGCGGCGCCGACGCGGCACCATGGCC
CAGGCGCTGCAGGCGGCGGGCAGGGCGAGCTGGACGTGCTGAGGTGCTGCA
GGCCTCCTGGGGCCCTCGCTGCGCGACCCGCTGGACGCGTGCCTCGTGCACCAACGCGGCC
CGCGCTGGGAAGCTGCACTGTCTGCGCTTCTGGTGGAGGAAGCCGCCCTCCCCGCCGCG
GCCCGCGCCCGCAACGGCGCCACACCGGCCACGACGCTCCGCCACCGGCCACCTCGCC
TGCCTGCAGTGGCTGCTGTCGAGGGCGGCTGCAGAGTGCAGGTGGGTCCGCGCGGTTCCG
CCAGGGGCACTGAGGCTTCTCCTCAGGACAGAGTCTTGCCCCAGAGTCCCCCGGGGCTC
AAGGATGGGTGGGGTTTGGCACCTCCTGGCCCAGCTGAACCCTGCACGGAGCTCCTCCA
```

Upstream sequence

Exon of an overlapping gene

ESPN Exon

The sequence is shown in FASTA format. Take a look at the FASTA header:

name of genome assembly

chromosome

base pair start

base pair end

forward strand (-1 is reverse)

```
>chromosome:GRCh37:1:6484248:6522030:1
AGCGCACTAGTGGTTCCCGTCCCTGCCTTCCCTGCCCTCCCCGCTGGAACCTCTGGGGGCAG
TTCTCGGATCTGGAGGGACCCTGGAAGGCAGGGCTCTTTGCAATCTCCGGGGATTTCGAC
CCAGAGCCCTTCAGGGACGTGGCAGGGCTGCTCCTGCCTCAGGGCCGTTGTCCTCGTGCT
```

Exons are highlighted within the genomic sequence. Variations can be added with the [Configure this page](#) link found at the left. Click on it now.

Configure Page | Manage Configurations | Personal Data

Display options

Save as...

Load configuration

Reset configuration

Display options

5' Flanking sequence (upstream): 600 * (Maximum of 1000000)

3' Flanking sequence (downstream): 600 * (Maximum of 1000000)

Number of base pairs per row: 60 bps

Additional exons to display: Core exons

Orientation of additional exons: Display exons in both orientation

Show variations: Yes and show links **Show variations**

Filter variations by consequence type: No filter, 3 prime UTR variant, 5 prime UTR variant, Coding sequence variant, Downstream gene variant

Line numbering: Relative to this sequence **Turn on line numbers**

Display pop-up information on mouseover: Yes

Fields marked * are required

Once you have selected changes (in this example, [Show variations](#) and [Line numbering](#)) click at the top right.

21181 CCAAGGAGTAGACCCCTCCCTTCCAGGTGACCCTGCCCTCTAGACACACAAAGCCTCCA 21240 [21240: rs139332523](#)

21241 GTGCTTCCCCTCCAAACCGGAGTGCCTGGTCTTCCCCAGTAAGTGTGGGCTGGGGCAG 21300

21301 GGTAGGGCCAGGGAGGGGAAGCCAGCACCCAGGGGCCACAGCAGGTGTACCAAGTGG 21360

21361 TGCCCGGAGCCACCTTGGCCCTCGGCAAGTTGTTCCAGGTGGTGGAGAGTCTCAGTCT 21420

21421 TGGGGGACAGCCTGTGTCATGCTCCAAATCTGGCCCTTCTTCTGCCTCCCTAGGCTCT 21480 [21424: rs139332523](#)

21481 **TTCCAGCGCTAGAGCTGCAGACATACAGAGCTACATGGACATGCTGAACCCGAGCTGGG** 21540 [21532: rs139332523](#)

21541 **CCTGCCTYRGGGCAYSATTGGGAAGMCCA**YACCCCAACCACCCCAACCAGCTTCCCCC 21600 [21548: rs143439475](#)

21601 **GCCACCCCGCCCCAGGCACCCA**ACTGCCCAACCCCACTGGCTACCCA**K**CTCCCAA 21660 [21653: TMP_ESP_1_65](#)

21661 **GCCTCCTGTGGACCACAGGCAGCTGR**CATCTACATGCAGACCAAGAACA**AACTCCGCCA** 21720 [21670: rs150033799](#)

21721 **CGTGGAGACAGAGGCCCTCAAGAAGGAG**TAGTGAGCCCTCACCCCTGCCTGCCTCCA 21780 [21766: rs201919218](#)

21781 GCA**SGGG**ACT**SGG**CTGATGGGGCCAGTGAGGCCAAAGCCCTGGCCTCACTAGTGGGCA 21840 [21784: TMP_ESP_1_65](#)

21841 TAGGGTGGGGATCCCTGGGTCCATGGCATGTTCAAGAGTCAAAGCTCCTGGCGAGTCCCA 21900

Links to the variation tab

Let's look at where our gene is expressed. Click on [Expression](#) in the left-hand menu.

Expression

Expression data is available for the following tissues:

Tissue	All data	RNASeq gene models	Intron-spanning reads	RNASeq alignments
Adipose	View in location	Models built using Human adipose total RNA, lot 05060581, caucasian female, throat cancer, Illumina Human Bodymap 2.0 Data	Y	Y
Adrenal	View in location	Models built using Human adrenal total RNA, lot 0812003, caucasian male, cerebral vascular accident, Illumina Human Bodymap 2.0 Data	Y	Y
Blood	View in location	Models built using Human white blood cell, caucasian male, healthy, Illumina Human Bodymap 2.0 Data	Y	Y
Brain	View in location	Models built using Human brain total RNA, lot 03070051, caucasian female, copd, Illumina Human Bodymap 2.0 Data	Y	Y

Hover over the column titles for a pop-up definition.

Can our gene be found in other databases? Go up the left-hand menu to [External references](#):

External references

This gene corresponds to the following database identifiers:

External database	Database identifier
HGNC Symbol	ESPN espin [view all locations]
EntrezGene	ESPN espin [view all locations]
MIM disease	DEAFNESS, AUTOSOMAL RECESSIVE 36, [#609006] DEAFNESS, AUTOSOMAL RECESSIVE 36, WITH OR WITHOUT VESTIBULAR INVOLVEMENT; [view all locations]
UniProtKB Gene Name	ESPN [view all locations]
Orphanet	Autosomal recessive nonsyndromic sensorineural deafness type DFNB Autosomal recessive nonsyndromic sensorineural deafness type DFNB [view all locations]
WikiGene	ESPN espin [view all locations]
MIM gene	ESPN, MOUSE, HOMOLOG OF [*606351] ESPN, MOUSE, HOMOLOG OF; ESPN [view all locations]
UniGene	Hs.652319 [Target %id: 99; Query %id: 94] Transcribed locus [view all locations] Hs.744222 [Target %id: 99; Query %id: 99] Espin [view all locations]
ArrayExpress	ENSG00000187017 [view all locations]

This contains links to the gene in other projects, such as Uniprot.

To find out more about the individual transcripts of this gene, click on [Transcript comparison](#) in the left-hand menu.

You must now choose the transcripts you'd like to see, click on the blue **Select transcripts** button.

Let's select all the **protein-coding transcripts**, then close the menu.

Demo: The transcript tab

Let's now explore one splice isoform. Click on **Show transcript table** at the top.

Click on the ID for the largest one, ESPN-001 (**ENST00000377828**).

Show/hide columns		Filter					
Name	Transcript ID	Length (bp)	Protein ID	Length (aa)	Biotype	CDS incomplete	CCDS
ESPN-005	ENST00000478323	270	ENSP00000466437	28	Protein coding	3'	-
ESPN-007	ENST00000434576	750	ENSP00000413621	188	Protein coding	5'	-
ESPN-002	ENST00000418286	641	ENSP00000401793	214	Protein coding	5' and 3'	-
ESPN-001	ENST00000377828	3531	ENSP00000367059	854	Protein coding	-	CCDS70
ESPN-009	ENST00000461727	1869	ENSP00000465308	288	Protein coding	-	-
ESPN-201	ENST00000416731	1665	ENSP00000399239	288	Protein coding	-	-
ESPN-004	ENST00000475228	813	No protein product	-	Processed transcript	-	-
ESPN-008	ENST00000468561	664	No protein product	-	Processed transcript	-	-
ESPN-006	ENST00000475479	360	No protein product	-	Processed transcript	-	-
ESPN-003	ENST00000477679	885	No protein product	-	Retained intron	-	-

You are now in the Transcript tab for ESPN-001. The left hand navigation column provides several options for the transcript ESPN-001. Click on the [Exons](#) link, circled in the image below.

The image shows a navigation menu on the left with the following items:

- Transcript-based displays
 - Transcript summary
 - Supporting evidence (20)
 - Sequence
 - Exons (13)** (highlighted with a callout: "Click Exons")
 - cDNA
 - Protein
 - External References
 - General identifiers (21)
 - Oligo probes (49)

The main area shows a sequence viewer with the following color-coded annotations:

- Green:** flanking sequence (indicated by a callout pointing to the top line of the sequence)
- Purple:** UTR (indicated by a callout pointing to the first few lines of the sequence)
- Grey:** coding sequence (indicated by a callout pointing to the middle lines of the sequence)
- Blue:** introns (indicated by a callout pointing to the bottom lines of the sequence)

You may want to change the display (for example, to show more flanking sequence, or to show full introns). In order to do so click on [Configure this page](#) and change the display options accordingly.

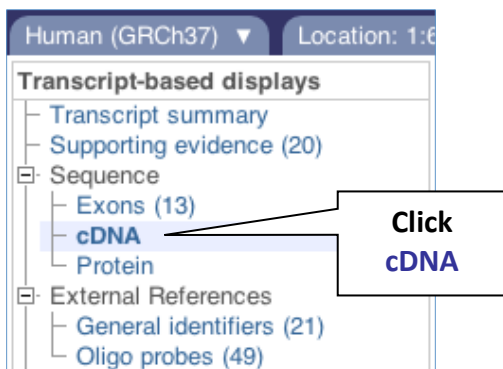
Display options

Flanking sequence at either end of transcript:	50
Number of base pairs per row:	60 bps
Intron base pairs to show at splice sites:	25
Show full intronic sequence:	<input checked="" type="checkbox"/>
Show exons only:	<input type="checkbox"/>
Line numbering:	None
Show variations:	In exons only
Filter variations by consequence type:	<ul style="list-style-type: none"> No filter 3 prime UTR variant 5 prime UTR variant Coding sequence variant Downstream gene variant

If you would like to export the sequence, including the colours, click [Download view as RTF](#). A Rich Text Format document will be generated that can be opened in word processor such as MS Word.



Now click on the [cDNA](#) link to see the spliced transcript sequence.



```

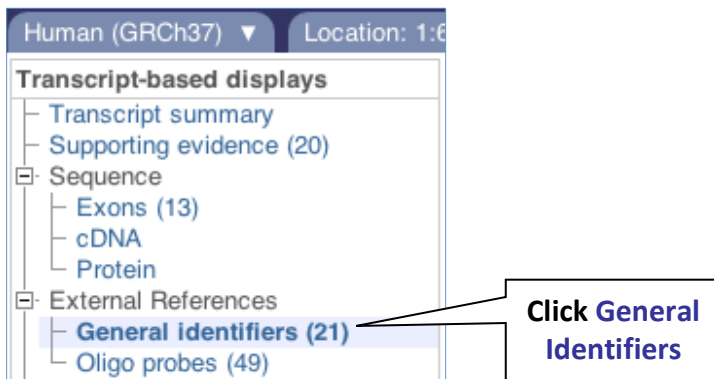
481 GCCACAGTCTTGCATCTGCTGCCCGCTTGGCCACCCCGAGGTGGTGAACCTGGCTCTTG
313 GCCACAGTCTTGCATCTGGCTGCCCGCTTCGGCCACCCCGAGGTGGTGAACCTGGCTCTTG
105 -A--T--V--L--H--L--A--A--R--F--G--H--P--E--V--V--N--W--L--L--

541 CATCATGGCGGTGGGGAACCCACCGCCGCCACAGACATGGGCGCCCTGCCTATCCACTAC
373 CATCATGGCGGTGGGACCCACCGCCGCCACAGACATGGGCGCCCTGCCTATCCACTAC
125 -H--H--G--G--G--D--P--T--A--A--T--D--M--G--A--L--P--I--H--Y--

```

UnTranslated Regions (UTRs) are highlighted in dark yellow, codons are highlighted in light yellow, and exon sequence is shown in black or blue letters to show exon divides. Sequence variants are represented by highlighted nucleotides and clickable IUPAC codes are above the sequence.

Next, follow the [General identifiers](#) link at the left.



This page shows information from other databases such as RefSeq, EntrezGene, OMIM, UniProtKB, and others, that matches to the Ensembl transcript and protein

General identifiers

This transcript corresponds to the following database identifiers:

External database	Database identifier
HGNC Symbol	ESPN espin [view all locations]
UniParc	UPI000013D2B6 [view all locations]
CCDS	CCDS70.1 [view all locations]
UniProtKB/Swiss-Prot	ESPN_HUMAN [align] Espn [view all locations]
RefSeq peptide	NP_113663.2 [Target %Id: 100; Query %Id: 100] [align] espin [view all locations]
RefSeq mRNA	NM_031475.2 [align] [view all locations]
UCSC Stable ID	uc001amy.3 [view all locations]
Human Protein Atlas	HPA028674 [view all locations] HPA028674 [view all locations]
European Nucleotide Archive	AF134401 [align] [view all locations] AL031848 [align] [view all locations] AL136880 [align] [view all locations] AL158217 [align] [view all locations] AY203958 [align] [view all locations] CH471130 [align] [view all locations]
HGNC transcript name	ESPN-001 espin [view all locations]
INSDC protein ID	AAD24480.1 [align] [view all locations] AAP34481.1 [align] [view all locations] CAB66814.1 [align] [view all locations] CAI19773.1 [align] [view all locations] CAI22163.1 [align] [view all locations] EAW71537.1 [align] [view all locations]

Click on [Ontology](#) table to see GO terms from the Gene Ontology consortium. www.geneontology.org

Ontology table ⓘ

- GO: Biological process
- GO: Cellular component
- GO: Molecular function

Descendants of GO: Biological process

Accession	Term	Evidence	Annotation Source	GOSlim Accessions	GOSlim Terms
GO:0007605	sensory perception of sound	IEA		GO:0008150 GO:0050877	biological_process neurological system process
GO:0007626	locomotory behavior	IEA		GO:0008150	biological_process
GO:0030046	parallel actin filament bundle assembly	IEA		GO:0007010 GO:0022607 GO:0008150	cytoskeleton organization cellular component assembly biological_process
GO:0051017	actin filament bundle assembly	IEA	[from Mus_musculus ENSMUSP00000030785]	GO:0007010 GO:0022607 GO:0008150	cytoskeleton organization cellular component assembly biological_process
GO:0051491	positive regulation of filopodium assembly	IEA		GO:0008150	biological_process
GO:0051494	negative regulation of cytoskeleton organization	IEA		GO:0008150	biological_process

Click on the ⓘ to see a guide to the three-letter Evidence codes.

Now click on [Protein summary](#) to view domains from Pfam, PROSITE, Superfamily, InterPro, and more.

Genetic Variation

- Variation table
- Variation image
- Population comparison
- Comparison image

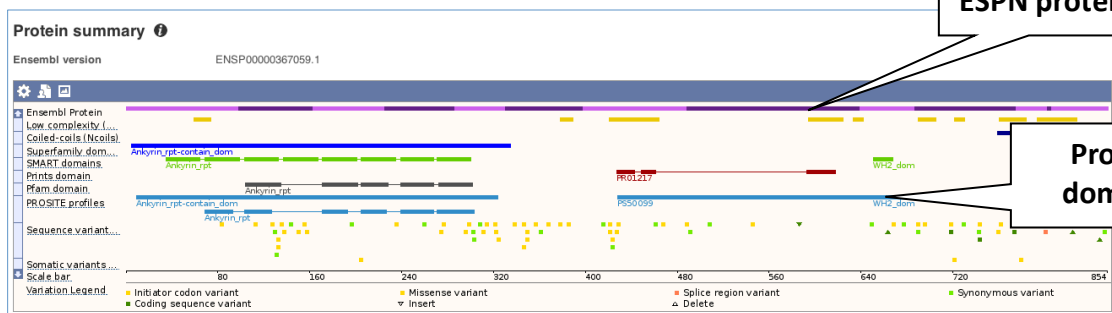
Protein Information

- Protein summary**
- Domains & features (40)
- Variations (139)

Click Protein Summary

Ensembl ESPN protein

Protein domains



Clicking on [Domains & features](#) shows a table of this information.

Domains & features ⓘ

Domains

Show **All** entries Show/hide columns Filter

Domain type	Start	End	Description	Accession	InterPro
Prints	427	443	-	PR01217	-
Prosite_profiles	428	730	-	PS50099	-
Prints	449	461	-	PR01217	-
Prints	593	618	-	PR01217	-
Prosite_profiles	768	825	-	PS50313	-
Smart	35	64	Ankyrin_rpt	SM00248	IPR002110 [Display all genes with this domain]
Prosite_profiles	69	93	Ankyrin_rpt	PS50088	IPR002110 [Display all genes with this domain]
Smart	69	99	Ankyrin_rpt	SM00248	IPR002110 [Display all genes with this domain]

Exercises: Genes and transcripts

Exercise 7 – Exploring the human *MYH9* gene

(a) Find the human *MYH9* (myosin, heavy chain 9, non-muscle) gene, and go to the [Gene tab](#).

- On which chromosome and which strand of the genome is this gene located?
- How many transcripts (splice variants) are there?
- How many of these transcripts are protein coding?
- What is the longest transcript, and how long is the protein it encodes?
- Which transcript has a CCDS record associated with it?

? Why is the CCDS important – what does it tell us?

(b) Click on [Phenotype](#) at the left side of the page. Are there any diseases associated with this gene, according to MIM (Mendelian Inheritance in Man)?

(c) In the transcript table, click on the transcript ID for MYH9-001, and go to the [Transcript tab](#).

- How many exons does it have?

- Are any of the exons completely or partially untranslated?
- Is there an associated sequence in UniProtKB/Swiss-Prot? Have a look at the [General identifiers](#) for this transcript.
- What are some functions of MYH9-001 according to the Gene Ontology consortium? Have a look at the [Ontology table](#) for this transcript.

(d) Are there microarray (oligo) probes that can be used to monitor ENST00000216181 expression?

Exercise 8 – Finding a gene associated with a phenotype

Phenylketonuria is a genetic disorder caused by an inability to metabolise phenylalanine. This results in an accumulation of phenylalanine causing seizures and mental retardation.

(a) Search for **phenylketonuria** from the Ensembl homepage. What gene is associated with this disorder?

(b) What tissues is this gene expressed in? Is this surprising, given the gene's role in disease? What is meant by "Intron-spanning reads" and "RNASeq alignments"?

(c) How many protein coding transcripts does this gene have? View all of these in the transcript comparison view.

(d) What is the MIM disease identifier for this gene? Does Orphanet list any other disorders associated with this gene?

Exercise 9 – Exploring a plant gene (*Vitis vinifera*, grape)

Start in <http://plants.ensembl.org/index.html> and select the *Vitis vinifera* genome.

(a) What GO: biological process terms are associated with the *MADS4* gene?

(b) Go to the transcript tab for the only transcript, Vv01s0010g03900.t01. How many exons does it have? Which one is the longest? How much of that is coding?

(c) What domains can be found in the protein product of this transcript? How many different groups agree with each of these domains?

BioMart

Demo: BioMart

Follow these instructions to guide you through BioMart to answer the following query:

You have three questions about a set of human genes: *ESPN*, *MYH9*, *USH1C*, *CHD7*, *CISD2*, *THRB*, *DFNB31* (these are HGNC gene symbols. More details on the HUGO Gene Nomenclature Committee can be found on <http://www.genenames.org>)

- 1) What are the EntrezGene IDs for these genes?
- 2) Are there associated functions from the GO (gene ontology) project that might help describe their function?
- 3) What are their cDNA sequences?

Step 1: Click on *BioMart* in the top header of a www.ensembl.org page to go to: www.ensembl.org/biomart/martview

NOTE: These answers were determined using BioMart Ensembl 71.

The screenshot shows the Ensembl BioMart interface. The top navigation bar includes 'Home', 'Login / Register', 'BLAST/BLAT', 'BioMart', and 'Docs & FAQs'. Below the navigation bar, there are buttons for 'New', 'Count', and 'Results'. The main content area is divided into two sections. The left section is a sidebar with 'Dataset' (None selected), 'Filters' (None selected), and 'Attributes' (Ensembl Gene ID, Ensembl Transcript ID). The right section contains a dropdown menu for the database, currently set to '- CHOOSE DATABASE -'. A callout box labeled 'STEP 2:' points to this dropdown menu, stating: 'Choose Ensembl Genes 72 as the primary database.' Below this, the dropdown menu is shown with 'Ensembl Genes 71' selected. A second callout box labeled 'STEP 3:' points to the dropdown menu, stating: 'Choose Homo sapiens genes as the dataset.'

STEP 4:
Click **Filters** at the left.
Expand the **GENE** panel.

Dataset: Homo sapiens genes (GRCh37.p10)

Filters
[None selected]

Attributes
Ensembl Gene ID
Ensembl Transcript ID

Dataset: [None Selected]

Please restrict your query using criteria below

- GENE:
- TRANSCRIPT EVENT:
- GENE ONTOLOGY:
- EXPRESSION:
- MULTI SPECIES COMPARISONS:
- PROTEIN DOMAINS:
- VARIATION:

STEP 5:
In **ID List Limit**, paste in your gene symbols. Change the heading to read **HGNC symbol(s) [e.g. ZFY]**.

Dataset: Homo sapiens genes (GRCh37.p10)

Filters
HGNC symbol(s) [e.g. ZFY]: [ID-list specified]

Attributes
Ensembl Gene ID
Ensembl Transcript ID

Dataset: [None Selected]

Please restrict your query using criteria below

- REGION:
- GENE:
 - Limit to genes ... with ArrayExpress
 - Only
 - Excluded
- ID list limit [Max 500 advised]
 - HGNC symbol(s) [e.g. ZFY]
 - ESPN, MYH9, USH1C, CHD7, CISD2, THRB, DFNB31
 - Choose File No file chosen

STEP 6:
Click **Count** to see BioMart is reading 8 genes out of 63,253 possible *H. sapiens* genes (this number includes ncRNA genes) (note that *CHD7* has an Ensembl copy and an LRG copy (<http://www.ensembl.org/Help/Glossary?id=406>), hence 8 counts for 7 HGNC genes).

Dataset: **8 / 63253** Homo sapiens genes (GRCh37.p11)

Filters
HGNC symbol(s) [e.g. ZFY]: [ID-list specified]

STEP 7:
Click on **Attributes** to select output options (i.e. GO terms)

Results

Features
 Homologs
 Structures
 Variation
 Transcript Event
 Sequences

GENE:
 EXTERNAL:
 EXPRESSION:
 PROTEIN DOMAINS:

HGNC symbol(s) [e.g. ZFY]:
 [ID-list specified]

Attributes
 Ensembl Gene ID
 Ensembl Transcript ID

STEP 8:
Expand the **EXTERNAL** panel.

STEP 9:
Scroll down to select **EntrezGene ID** (to answer question 1)

LRG to Ensembl link transcript
 EntrezGene ID
 VEGA transcript ID(s) (OTTT)
 VEGA gene ID(s) (OTTG)
 Ensembl transcript (where OTTT shares CDS with ENST)
 HAVANA transcript (where ENST
 HAVANA transcript (where ENST
 HGNC ID(s)
 HGNC symbol

STEP 10:
Also select **HGNC symbol** to see the input gene symbols we started with.

STEP 11:
Scroll back up to select **GO term** fields (to answer question 2)

GO Term Accession
 GO Term Name
 GO Term Definition

STEP 12:
Click **Results**.

Export: all results to TSV Unique results only

Email notification to:

View: 10 rows as HTML Unique results only

Ensembl Gene ID	Ensembl Transcript ID	EntrezGene ID	HGNC symbol	GO Term Accession	GO Term Name	GO Term Definition
ENSG00000151090	ENST00000356447	7068	THRB	GO:0000122	negative regulation of transcription from RNA polymerase II promoter	"Any process that stops, prevents, or reduces the frequency, rate or extent of transcription from an RNA polymerase II promoter." [GOC:go_curators, GOC:txnOH]
ENSG00000151090	ENST00000356447	7068	THRB	GO:0006351	transcription, DNA-dependent	"The cellular synthesis of RNA on a template of DNA." [GOC:j, GOC:txnOH]
ENSG00000151090	ENST00000356447	7068	THRB	GO:0045944	positive regulation of transcription from RNA polymerase II promoter	"Any process that activates or increases the frequency, rate or extent of transcription from an RNA polymerase II promoter." [GOC:go_curators, GOC:txnOH]

Dataset 8 / 62252 Genes
 Homo sapiens genes (GRCh37.p10)
 Filters
 HGNC symbol(s) [e.g. ZFY]: [ID-list specified]
Attributes
 Ensembl Gene ID
 Ensembl Transcript ID
 EntrezGene ID
 HGNC symbol
 GO Term Accession
 GO Term Name
 GO Term Definition
 Dataset
 [None Selected]

Why are there multiple rows for one gene ID? For example, look at the first few rows.

Ensembl Gene ID	Ensembl Transcript ID	EntrezGene ID	GO Term Accession	GO Term Name	GO Term Definition	HGNC symbol
ENSG00000187017	ENST00000377828	83715	GO:0007605	sensory perception of sound	"The series of events required for an organism to receive an auditory stimulus, convert it to a molecular signal, and recognize and characterize the signal. Sonic stimuli are detected in the form of vibrations and are processed to form a sound." [GOC:ai]	ESPN
ENSG00000187017	ENST00000377828	83715	GO:0007626	locomotory behavior	"The specific movement from place to place of an organism in response to external or internal stimuli. Locomotion of a whole organism in a manner dependent upon some combination of that organism's internal state and external conditions." [GOC:dph]	ESPN
ENSG00000187017	ENST00000377828	83715	GO:0030046	parallel actin filament bundle assembly	"Assembly of actin filament bundles in which the filaments are tightly packed (approximately 10-20 nm apart) and oriented with the same polarity." [GOC:man, ISBN:0815316194]	ESPN

STEP 13:
Click *Attributes* again

The screenshot shows the Ensembl genome browser interface. On the left, the 'Attributes' sidebar is expanded, showing options like 'Ensembl Gene ID', 'Ensembl Transcript ID', and 'cDNA sequences'. In the main content area, the 'Sequences' option is selected under the 'SEQUENCES' section. Below this, there are radio button options for 'Unspliced (Transcript)', 'Unspliced (Gene)', 'Flank (Transcript)', 'Flank (Gene)', 'Flank-coding region (Transcript)', and 'Flank-coding region (Gene)'. On the right side of the main content area, there are more radio button options: '3' UTR', 'Exon sequences', 'cDNA sequences', 'Coding sequence', and 'Protein'. The 'cDNA sequences' option is selected.

STEP 14:
Select *Sequences* at the top, then expand *SEQUENCES* and choose the option *cDNA sequences* (to answer question 3).

The screenshot shows the 'Header Information' sidebar in the Ensembl genome browser. Under the 'Gene Information' section, the following options are listed with checkboxes: 'Ensembl Gene ID' (checked), 'Description' (unchecked), 'Associated Gene Name' (checked), 'Associated Gene DB' (unchecked), and 'Chromosome Name' (unchecked).

STEP 15:
Expand *Header Information* to select the *Associated Gene Name*

STEP 16:
Click [Results](#) to see the cDNA sequences in FASTA format.

STEP 17:
Change [View 10 rows](#) to [View All rows](#) so that you see the full table.
Note: Pop-up blocking must be switched off in your browser.

Export all results to FASTA Unique results only

Email notification to

View rows as FASTA Unique results only

```
>ENSG00000006611 | ENST00000000926 | USH1C
ATGGACCGAAAAAGTGGCCCGAGAATTCCGGGCTGGGATTTCTGATTGAAAATGAT
GCAGAGAAGGACTATCTCTATGATGTGCTGCGAACTGAGACCATGGACGTGGCC
GTGCTCGTGGGAGACCTGAAGCTGGTCATCAATGAAGCTCTCTGTTTGGT
GCCATTCGGCCGCTGATCCCACTGAAGCACCA
CGCTCCAGGAAGCTGAAGGAGGTGCGTCTGGA
AGTGTGCGTGGTGGCCTGGAGTTTGCTGTGG
GGTCAGGCAGACAGCGTCGGGCTCCAGGTAGG
TCCATCTCCTCCTGTACCCATGAGGAGGTGAT
TCCATCAAAGTGAGACACATCGGCCTGATCCC
ACTTGGCAGTATGTGGATCAGTTTGTGTGCGGA
TCCCCTGGAAATCGGAAAACAAGGAGAAGAA
GGCCTTGGCTGCAGCATTTCCAGCGGCCCAT
GTGAAACCTGGCTCCCTGTCTGTGAGGTGGG
GTCAATGGCGTCGACTTCTTAACCTGGATCA
AGCCCGAGCCTGACCATCTCATTGTAGCTGC
CGGGAGCGGCTGGCAGAGCGCGGCAGCGTGA
AAGCGGCTGGCGATGGAGTCCAACAAGATCCT
AGGAGAAAAGAAATGCCCAGAAGGCAGCAGA
GAACAGATTGTAGAGGAGGAAGAGAAGTTTAA
```

Note: you can use the [Go](#) button to export a file.

*What did you learn about the human genes in this exercise?
Could you learn these things from the Ensembl browser? Would it take longer?*

For more details on BioMart, have a look at these publications:

Smedley, D. *et al*
BioMart – biological queries made easy
BMC Genomics 2009 Jan 14;10:22

Kinsella, R.J. *et al*
Ensembl BioMart: a hub for data retrieval across taxonomic space.
Database (Oxford) 2011;bar030

Exercises: BioMart

Exercise 10 – Finding genes by protein domain

Find mouse proteins with transmembrane domains located on chromosome 9.

Exercise 11 – Convert IDs

BioMart is a very handy tool when you want to convert IDs from different databases. The following is a list of 29 IDs of **human proteins** from the NCBI RefSeq database

(<http://www.ncbi.nlm.nih.gov/projects/RefSeq/>):

NP_001218, NP_203125, NP_203124, NP_203126, NP_001007233,
NP_150636, NP_150635, NP_001214, NP_150637, NP_150634,
NP_150649, NP_001216, NP_116787, NP_001217, NP_127463,
NP_001220, NP_004338, NP_004337, NP_116786, NP_036246,
NP_116756, NP_116759, NP_001221, NP_203519, NP_001073594,
NP_001219, NP_001073593, NP_203520, NP_203522

Generate a list that shows to which Ensembl Gene IDs and to which HGNC symbols these RefSeq IDs correspond. Do these 29 proteins correspond to 29 genes?

Hint: For this exercise, it's easier to copy and paste the IDs from the online exercise booklet. One copy is here:

URL

Exercise 12 – Export homologues

For a list of *Ciona savignyi* Ensembl genes, export the human orthologues.

ENSCSAVG000000000002, ENSCSAVG000000000003,
ENSCSAVG000000000006, ENSCSAVG000000000007,
ENSCSAVG000000000009, ENSCSAVG000000000011

Exercise 13 – Export structural variants

You can use BioMart to query variants, not just genes.

(a) Export the study accession, source name, chromosome, sequence region start and end (in bp) of human structural variations (SV) on chromosome 1, starting at 130,408 and ending at 210,597.

(b) In a new BioMart query, find the alleles, phenotype descriptions, and associated genes for rs1801500 and rs1801368. Can you view this same information in the Ensembl browser?

Exercise 14 – Find genes associated with array probes

Forrest *et al* performed a microarray analysis of peripheral blood mononuclear cell gene expression in benzene-exposed workers (Environ Health Perspect. 2005 June; 113(6): 801–807). The microarray used was the human Affymetrix U133A/B (also called U133 plus 2) GeneChip. The top 25 up-regulated probe-sets were:

207630_s_at, 221840_at, 219228_at, 204924_at, 227613_at,
223454_at, 228962_at, 214696_at, 210732_s_at, 212370_at,
225390_s_at, 227645_at, 226652_at, 221641_s_at, 202055_at,
226743_at, 228393_s_at, 225120_at, 218515_at, 202224_at,
200614_at, 212014_x_at, 223461_at, 209835_x_at, 213315_x_at

(a) Retrieve for the genes corresponding to these probe-sets the Ensembl Gene and Transcript IDs as well as their HGNC symbols and descriptions.

(b) In order to analyse these genes for possible promoter/enhancer elements, retrieve the 2000 bp upstream of the transcripts of these genes.

(c) In order to be able to study these human genes in mouse, identify their mouse orthologues. Also retrieve the genomic coordinates of these orthologues.

Exercise 15 – BioMart in Ensembl Fungi

You can use BioMart for the non-vertebrate species hosted at www.ensemblgenomes.org. Export a list of Gene IDs (from the PomBase project only) for *S. pombe* that are protein coding and located on Chromosome III.

(Start at <http://fungi.ensembl.org>)

Variation

Demo: Finding variants in Ensembl

In any of the sequence views shown in the [Gene](#) and [Transcript](#) tabs, you can view variants on the sequence. You can do this by clicking on [Configure this page](#) from any of these views.

Let's take a look at the [Gene sequence](#) view for *MCM6* in human. Search for *MCM6* and go to the [Sequence](#) view.

If you can't see variants marked on this view, click on [Configure this page](#) and select [Show variations: Yes](#) and [show links](#).

Marked-up sequence ⓘ

Key

Exons	All exons in this region	MCM6 exons	
Variations	3 prime UTR	5 prime UTR	Frameshift
	Intron	Missense	Splice region
	Synonymous	Upstream	

>chromosome:GRCh37:2:136596596:136634596:-1

1 GAACTCCTGACCTCAGGTGATCCACACGCCCTCGGCCTCCAAAAGTGCTAGGATTACAGGT 60 [40: rs1057031](#)

61 TGTGAGCCACCGCCCGCCGCAATGTCAAAATTTTCGCTGATTTTCCTCGGTCAGCACCA 120

121 CCACAATAATTTAGACATCTCTTTGACTCTTTTACA 180

181 GGGTTTTAFAACTAGACAGCAGAAGCGGCTTACTCA 240 [197: rs72972191](#);

241 TTTTCTCTCAATTAATATCAGTGGAAATGATTTAA 300 [257: rs4988140](#);

301 AGAACATTCAGATTCCAAGCTTGTCCAGCAGCCAA 360

361 AAATACCCGCGCAAAGGCAGATGGGCTTTCTCCAGA 420

421 AGGGCTTTTGGGCTGGGGCTCTTGGAGAGGCGCCG 480 [430: rs4988141](#); [45](#)

481 TTCATTGGTCAGGTTGGCGCCCTCCAGCTCCTGTGCACGATTGGTTCGGGCCGT 540 [496: rs75464314](#);

541 GCAGGTCGGAAGAGGGGGCGGCGGAAGCGGGCGCGCGCCGCAAAGCTGCAGCGTCT 600

601 **GGAAAAAGCGACTTGTGGCGTCGAGCGTGGCGYAGGCGAATCCTCGGCCACTRAGCRAA** 660 [621: rs3087349](#); [6](#)

661 **TATGGACCTGCGRCGGCAGCGGAGCCGGGCCCGCAGCCAGCACCTGGAGGTCGGCGA** 720 [670: rs201537325](#);

721 **CGAGGTGGCCGAGAAGTGCCAGAACTGTTCTCTGGACTTCTTGGWGA** 780 [765: rs3087355](#);

781 GCGCCCCGGGCTCGCCCGCCTCCGGAGCCTGCGGCGTSTGGGCCGGCGGTGCCGTGTG 840 [819: rs148582236](#);

841 **SGCGGCGGGTGTTCGGAACTGGGGTTCGGCTGTCGGGAAGCGMCTCCCCGCCGCC** 900 [841: rs138586953](#);

901 CCCAACTTTAGGCTCCGAGCCCGCGCCGCGAGGAGCCTGSGACAGGCTGGGTTTTTCT 960 [942: rs111751203](#);

961 GGAGTAAAACCATCTCGAGCCCTAAGACGCTCGTGCCAGTGTCTCAGCGCTTTGCTTCT 1020

Find out more about a variant by clicking on it.

Variation: [rs1057031](#)

Position: 2:136633962

Alleles: G/A

Types: 5_prime_UTR_variant
regulatory_region_variant

[Gene/Transcript Locations](#)

[Population Allele Frequencies](#)

You can add variants to all other sequence views in the same way.

You can go to the **Variation** tab by clicking on the **variant ID**. For now, we'll explore more ways of finding variants.

To view all the sequence variations in table form, click the **Variation table** link at the left of the gene tab.

Gene: MCM6 ENSG00000076003

Description minichromosome maintenance complex component 6 [Source:HGNC Symbol;Acc:6949]

Location [Chromosome 2: 136,597,196-136,633,996](#) reverse strand.

INSDC coordinates chromosome:GRCh37:CM000664.1:136597196:136633996:1

Transcripts This gene has 3 transcripts (splice variants) [Show transcript table](#)

Variation table ⓘ

Summary of variation consequences in ENSG00000076003 [Switch to tree view](#)

Show	All ▾	entries	Filter
Number of variant consequences	Type	Description	
0	Transcript ablation	A feature ablation whereby the deleted region includes a transcript feature (SO:0001893)	
0	Splice donor variant	A splice variant that changes the 2 base region at the 5' end of an intron (SO:0001578)	
1	Splice acceptor variant	A splice variant that changes the 2 base region at the 3' end of an intron (SO:0001574)	
0	Stop gained	A sequence variant whereby at least one base of a codon is changed, resulting in a premature stop codon, leading to a shortened transcript (SO:0001582)	
1	Frameshift variant	A sequence variant which causes a disruption of the translational reading frame, because the number of nucleotides inserted or deleted is not a multiple of three (SO:0001589)	
0	Stop lost	A sequence variant where at least one base of the terminator codon (stop) is changed, resulting in an elongated transcript (SO:0001579)	
0	Initiator codon variant	A codon variant that changes at least one base of the first codon of a transcript (SO:0001582)	

The table is divided into consequence types. You can also view the consequence types as an ontology tree. Click on **Switch to tree view** at the right hand side to change the view.

Summary of variation consequences in ENSG00000076003

- All variants | Show (2826)
 - Upstream gene variant | Show (323) | SO:0001631
 - Downstream gene variant | Show (388) | SO:0001632
 - Feature elongation | Show (64) | SO:0001907
 - Feature truncation | Show (75) | SO:0001906
 - Gene variant | Show (1976) | SO:0001564
 - Splicing variant | Show (16) | SO:0001568
 - Splice region variant | Show (16) | SO:0001630
 - Transcript variant | Show (1960) | SO:0001576
 - Nc transcript variant | Show (563) | SO:0001619
 - Intron variant | Show (1173) | SO:0001627
 - Splice site variant | Show (1) | SO:0001629
 - Splice acceptor variant | Show (1) | SO:0001574
 - UTR variant | Show (38) | SO:0001622
 - 5 prime UTR variant | Show (4) | SO:0001623
 - 3 prime UTR variant | Show (34) | SO:0001624
 - Exon variant | Show (186) | SO:0001791
 - Non coding exon variant | Show (46) | SO:0001792
 - Coding sequence variant | Show (140) | SO:0001580
 - Synonymous variant | Show (35) | SO:0001819
 - Protein altering variant | Show (104) | SO:0001818
 - Inframe variant | Show (103) | SO:0001650
 - Missense variant | Show (103) | SO:0001583
 - Frameshift variant | Show (1) | SO:0001589

Click on **Show** to expand a detailed table for any of the consequence types available.

Let's expand Missense variants.

ID	Chr: bp	Alleles	Global MAF	Class	Source	Evidence	Type	AA	AA coord	SIFT	PolyPhen	Transcript
rs200231978	2:136598413	C/T	-	SNP	dbSNP	-	Missense variant	E/K	820	0.02	0.508	ENST00000264156
rs1804699	2:136598428	G/T	-	SNP	dbSNP	-	Missense variant	P/T	815	0	0.998	ENST00000264156
rs5660827	2:136598443	A/G	0.001 (G)	SNP	dbSNP	⚡	Missense variant	Y/H	810	0.22	0.71	ENST00000264156
rs988283	2:136598455	C/T	-	SNP	dbSNP	⚡	Missense variant	E/K	806	0.14	0.104	ENST00000264156
rs148613967	2:136598457	T/C	-	SNP	dbSNP	⚡	Missense variant	Y/C	805	0.06	0.011	ENST00000264156
rs25M570243	2:136598469	C/T	-	somatic_SNP	COSMIC	-	Missense variant	G/E	801	1	0	ENST00000264156
rs138809319	2:136598471	C/G	0.001 (G)	SNP	dbSNP	⚡	Missense variant	E/D	800	0.45	0	ENST00000264156
TMP_ESP_2_136598493	2:136598493	G/A	-	SNP	ESP	-	Missense variant	A/V	793	0.24	0.004	ENST00000264156

The table contains lots of information about the variants. You can click on the IDs here to go to the Variation tab too.

Let's look at Structural Variation in the Gene Tab. You'll find it in the left-hand menu.

Structural variation

Contigs: AC011893.7 >

Genes (Merged): MCM6-001 protein coding, MCM6-003 processed transcript, MCM6-002 retained intron

Larger structural variants: Condensed into a single bar

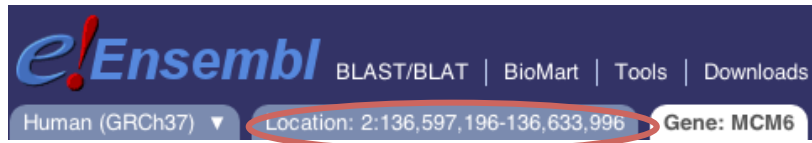
Smaller structural variants: Shown individually

Structural variants table

Name	Chr:bp	Genomic size (bp)	Class	Source Study	Study description
nsv429565	2:2784-242738129	242,735,346	CNV	DGVa:nstd11	Database of Genomic Variants Archive: W 2009 "Acquired copy number alteration in acute myeloid leukemia genomes." PMID:19651600 [remapped from build NCBI36]
esv1914561	2:25372589-222958286	-	Intrachromosomal breakpoint	DGVa:estd192	Database of Genomic Variants Archive: Catalogue of Somatic Mutations in Cancer (COSMIC) version 61
esv1914533	2:25432903-222993892	-	Intrachromosomal breakpoint	DGVa:estd192	Database of Genomic Variants Archive: Catalogue of Somatic Mutations in Cancer (COSMIC) version 61

You can click on the structural variants (SVs) in the image, or on their IDs in the table to go to the SV tab.

Let's have a look at variants in the Location tab. Click on the [Location](#) tab in the top bar.



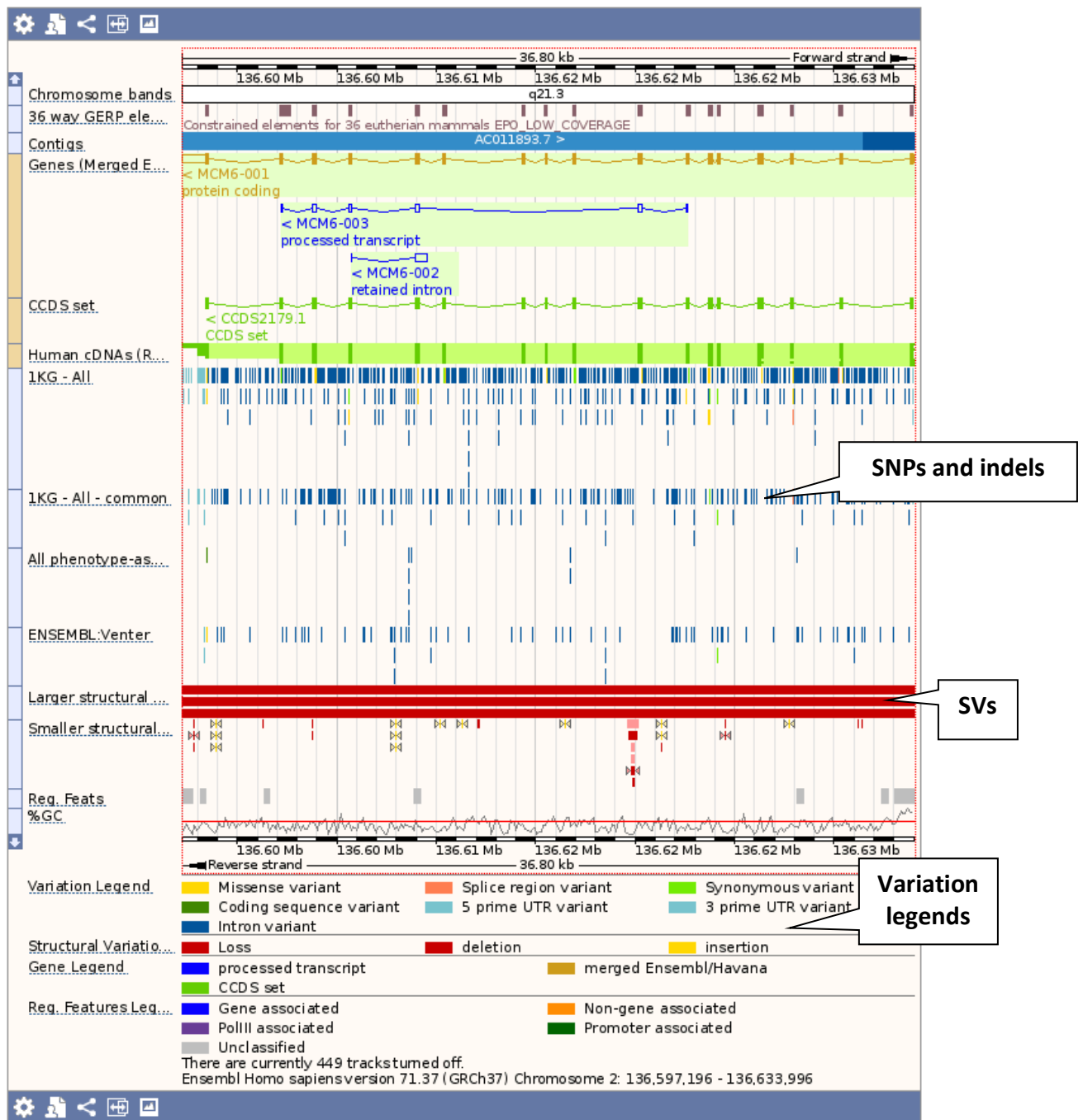
Configure this page and open [Variation](#) from the left-hand menu.

Active tracks	Variation
Favourite tracks	<input type="text" value="Find a track"/>
Track order	Enable/disable all dbSNP
Search results	<input type="checkbox"/> Sequence variants (dbSNP and all other sources) ★ ⓘ
<ul style="list-style-type: none"> <input checked="" type="checkbox"/> Sequence and assembly (3/29) <ul style="list-style-type: none"> Sequence (2/4) Markers (0/1) GRC alignments (1/3) Ditag features (0/2) Simple features (0/6) Clones (0/13) <input checked="" type="checkbox"/> Genes and transcripts (2/62) <ul style="list-style-type: none"> Genes (2/6) Prediction transcripts (0/2) RNASeq models (0/54) <input checked="" type="checkbox"/> mRNA and protein alignments (1/15) <ul style="list-style-type: none"> mRNA alignments (1/3) EST alignments (0/1) Protein alignments (0/5) Protein features (0/6) <input checked="" type="checkbox"/> ncRNA (0/1) <input checked="" type="checkbox"/> Variation (0/77) <ul style="list-style-type: none"> dbSNP (0/2) 1000 Genomes & HapMap (0/13) Phenotype and curated variants (0/13) Individual genomes (0/14) Arrays and other (0/12) Failed variants (0/1) Structural variants (0/18) Phenotype annotations (0/4) 	<ul style="list-style-type: none"> <input type="checkbox"/> dbSNP variants ★ ⓘ <p>HapMap</p> <ul style="list-style-type: none"> <input type="checkbox"/> All HapMap ★ ⓘ <p>Enable/disable all 1000 Genomes</p> <ul style="list-style-type: none"> <input type="checkbox"/> 1000 Genomes - AFR ★ ⓘ <input type="checkbox"/> 1000 Genomes - AFR - common ★ ⓘ <input type="checkbox"/> 1000 Genomes - All ★ ⓘ <input type="checkbox"/> 1000 Genomes - All - common ★ ⓘ <input type="checkbox"/> 1000 Genomes - AMR ★ ⓘ <input type="checkbox"/> 1000 Genomes - AMR - common ★ ⓘ <input type="checkbox"/> 1000 Genomes - ASN ★ ⓘ <input type="checkbox"/> 1000 Genomes - ASN - common ★ ⓘ <input type="checkbox"/> 1000 Genomes - EUR ★ ⓘ <input type="checkbox"/> 1000 Genomes - EUR - common ★ ⓘ <input type="checkbox"/> 1000 Genomes - High coverage - Trios ★ ⓘ <input type="checkbox"/> 1000 Genomes - Low coverage ★ ⓘ <p>Enable/disable all Phenotype and curated variants</p> <ul style="list-style-type: none"> <input type="checkbox"/> Uniprot phenotype variants ★ ⓘ <input type="checkbox"/> OMIM phenotype variants ★ ⓘ

There are various options for turning on variants. You can turn on variants by source, by frequency, presence of a phenotype or by individual genome they were isolated from. Turn on the following sequence variants in [Normal](#).

- [1000 genomes – All](#)
- [1000 genomes – All – common](#)
- [All phenotype-associated variants](#)
- [ENSEMBL:Venter](#)

Also turn on [Larger](#) and [Smaller Structural variants \(all sources\)](#) in [Expanded](#).



Click on a variant to find out more information. It may be easier to see the individual variants if you zoom in.

Let's zoom in on the region **2:136607850-136609811** by typing it into the **Location** box.

Change the track style on one of the SNP variant tracks to **Expanded with name**. Click on the variant **rs4988235** to open a pop-up, then click on the ID to open the **Variation** tab.

Variation displays

- Explore this variation
- Genomic context
- Genes and regulation (3)
- Flanking sequence
- Population genetics
- Individual genotypes (1869)
- Linkage disequilibrium
- Phenotype Data (1)
- Phylogenetic Context (6)
- Citations (4)
- External Data
- SNPedia
- LOVD

rs4988235 SNP

Variant information

Original source: Variants (including SNPs and indels) imported from dbSNP (released in dbSNP)

Alleles: Reference/Alternative: G/A | Ancestral: G | Ambiguity code: none

Location: Chromosome 2:136608646 (forward strand) | [View in location tab](#)

Co-located with HGMD-PUBLIC [CR024269](#)

Evidence status:

Clinical significance: HASH(0x172b850) (from dbSNP) | [View explanation](#)

Synonyms: None currently in the database

HGVS names: This variation has 4 HGVS names - click the plus to show

- [2:g.136608646G>A](#)
- [ENST00000264156.2:c.1917+326C>T](#)
- [ENST00000492091.1:n.343+326C>T](#)
- [ENST00000483902.1:n.544+326C>T](#)

Genotyping chips: This variation has assays on 4 chips - click the plus to show

Variation icons: These go to the same places as the links on the left

- Genomic context
- Genes and regulation
- Population genetics
- Individual genotypes
- Linkage disequilibrium
- Phenotype data
- Citations
- Phylogenetic context
- Flanking sequence

The icons show you what information is available for this variant. Click on [Genes and regulation](#), or follow the link at the left.

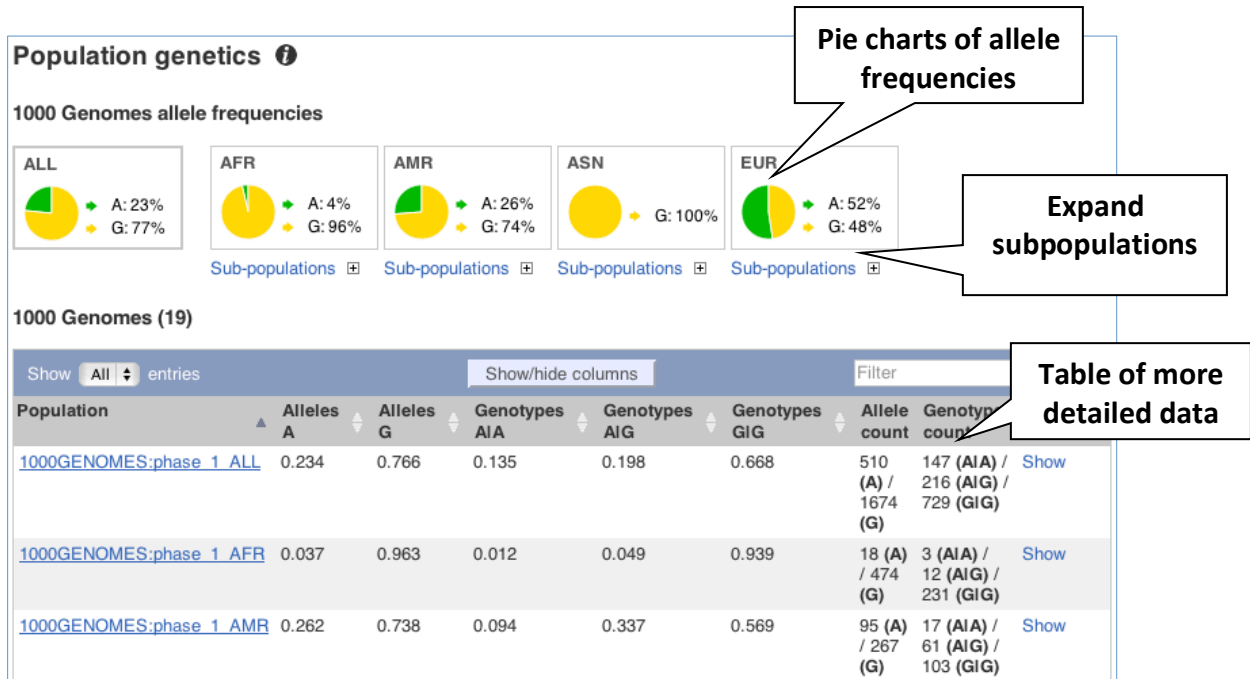
Genes and regulation

Gene and Transcript consequences

Gene	Transcript (strand)	Allele (transcript allele)	Type	Position in transcript	Position in CDS	Position in protein	Amino acid	Codons	SIFT	PolyPhen	Detail
ENSG00000076003 HGNC: MCM6	ENST00000264156 (-) biotype: protein_coding	A (T)	Intron variant	-	-	-	-	-	-	-	Show
ENSG00000076003 HGNC: MCM6	ENST00000483902 (-) biotype: retained_intron	A (T)	NC transcript variant, intron variant	-	-	-	-	-	-	-	Show
ENSG00000076003 HGNC: MCM6	ENST00000492091 (-) biotype: processed_transcript	A (T)	NC transcript variant, intron variant	-	-	-	-	-	-	-	Show

This variant is found in three transcripts of the *MCM6* gene. It has not been associated with any regulatory features or motifs.

Let's look at population genetics. Either click on [Explore this variant](#) in the left hand menu then click on the [Population genetics](#) icon, or click on [Population genetics](#) in the left-hand menu.



These data are mostly from the **1000 genomes** and **HapMap** projects in human.

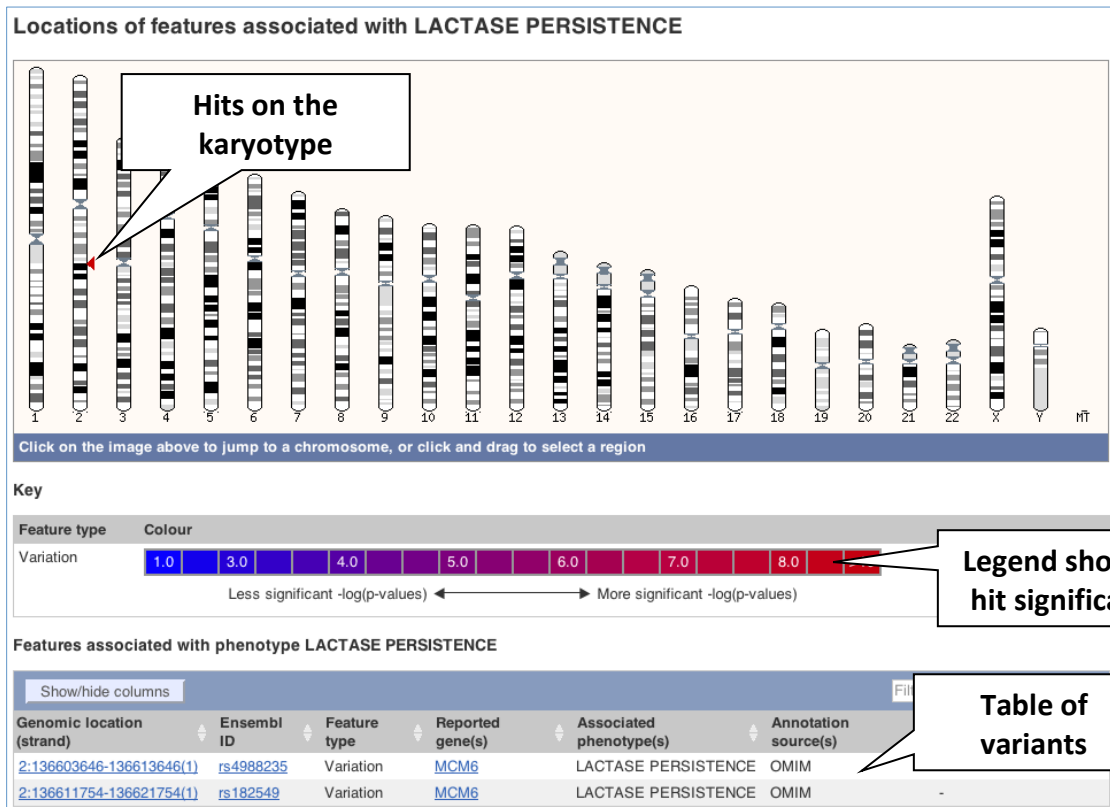
There are big differences in allele frequencies between populations. Let's have a look at the phenotypes associated with this variant to see if they are known to be specific to certain human populations. Either click on [Explore this variant](#) in the left hand menu then click on the [Phenotype data](#) icon, or click on [Phenotype Data](#) in the left-hand menu.

Phenotype Data ⓘ

Disease/Trait	Source(s)	Study	Reported gene(s)	Associated variant(s)	Most associated allele	P value
LACTASE PERSISTENCE [View on Karyotype]	OMIM	MIM:601806	MCM6	rs4988235	0001	

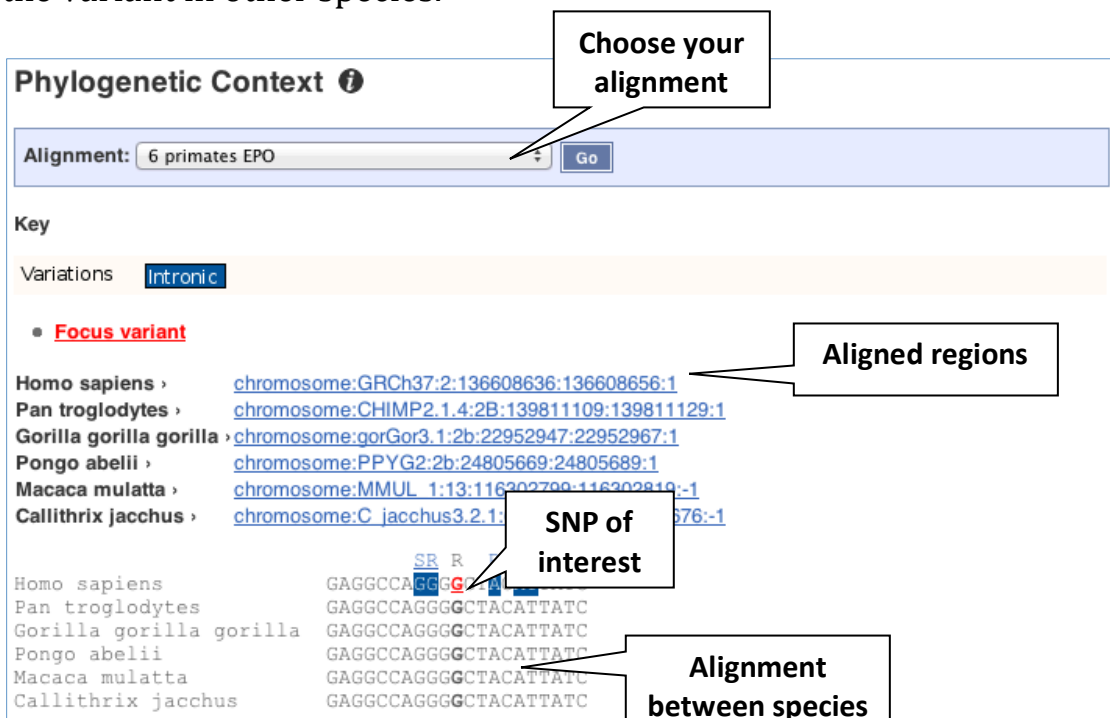
This variant is associated with lactase persistence, which is known to be common in European populations, and rare in Asian populations, exactly as we saw in the allele frequencies in these populations.

Are there other variants in the genome that also cause lactase persistence? Click on [View on Karyotype](#) to find out.



Two variants are known to be associated with this phenotype. Both are found with the *MCM6* gene.

Click back to the [Variation Tab](#). Click on [Phylogenetic context](#) to see the variant in other species.



The variant is not marked in the other species. This means that the variant arose in humans.

Another way to look at variation is using the **Resequencing** view. Click on the **Location** tab, then choose **Resequencing** from the left-hand menu.

Resequencing Alignments

Location: 2:136607850-136609811 Go Gene: Go

Key

Variations **Intronic**

Other features **All exons**

- Basepairs in secondary individuals matching the reference individual are replaced by dots
- ~ No resequencing coverage at this position

Homo_sapiens > chromosome:GRCh37:2:136607850-136609811.1

Homo_sapiens	1	TCTCCCTCTCGTTTTTTTTTAAATTCACACTTATATTTAGGTTCTGGGGGTACACATGCAGGTTTGTACATAGTAAACTGTGTCTATGGGGATTAGTATATAGATTATTTTCATCAC	120
VENTER	1	120
WATSON	1	120
Homo_sapiens	121	CCACTCTATTTTACATGCTTAATTAATACAGGAGGGCTCTCATTCAATTTTTGTCCCTACAGGTGTTCCTTAATAATTGGG	240
VENTER	121	240
WATSON	121	240
Homo_sapiens	241	TGAATCAAGAATATTAATAACCTAGTGTGAAATTTGAAAATTTAGTGGAAAAAGCCAAAAATTAACCTGTGGGATAAAAAGT	360
VENTER	241	360
WATSON	241	360
Homo_sapiens	361	AGTGATTGAAGTCGGAACAGTGGTTACTGAGAGAGGGCTAAGACTGAAAAGAGGCAGGGGTTGGAACTTTCTGAAATTATGGAAATGTTTTGGATCTTGAGCTGGGTATTCACA	480
VENTER	361	480
WATSON	361	480

This view is used to look at sequence between individuals. Craig Venter and James Watson are shown by default. You can change the individuals shown by clicking on [Configure this page](#).

It can also be used to look at different mouse strains or dog breeds. Can you find our variant rs4988235 in this view? Is the alternate allele present in James Watson or Craig Venter?

Exercises: Finding variants in Ensembl

Exercise 16 – Human population genetics and phenotype data

The SNP rs1738074 in the 5' UTR of the human *TAGAP* gene has been identified as a genetic risk factor for a few diseases.

(a) In which transcripts is this SNP found?

(b) What is the least frequent genotype for this SNP in the Yoruba (YRI) population from the HapMap set?

(c) What is the ancestral allele? Is it conserved in the 36 eutherian mammals?

(d) With which diseases is this SNP associated? Are there any known risk (or associated) alleles?

Exercise 17 – Exploring a SNP in human

The missense variation rs1801133 in the human *MTHFR* gene has been linked to elevated levels of homocysteine, an amino acid whose plasma concentration seems to be associated with the risk of cardiovascular diseases, neural tube defects, and loss of cognitive function. This SNP is also referred to as ‘A222V’, ‘Ala222Val’ as well as other HGVS names.

(a) Find the page with information for rs1801133.

(Note: a bug in the current release means that these alleles are erroneously reported as G/A/CT/CT. Please ignore these extra CT alleles.)

(b) Is rs1801133 a Missense variation in all transcripts of the *MTHFR* gene?

(c) Why are the alleles for this variation in Ensembl given as G/A and not as C/T, as in dbSNP and literature?

(http://www.ncbi.nlm.nih.gov/projects/SNP/snp_ref.cgi?rs=1801133)

(d) What is the major allele in rs1801133?

(e) In which paper is the association between rs1801133 and homocysteine levels described?

(f) According to the data imported from dbSNP, the ancestral allele for rs1801133 is G. Ancestral alleles in dbSNP are based on a comparison between human and chimp. Does the sequence at this same position in four other primates, i.e. gorilla, orangutan, macaque and marmoset, confirm that the ancestral allele is G?

(g) Were both alleles of rs1801133 already present in Neanderthal?

To answer this question, have a look at the individual reads at its genomic position in the Neanderthal Genome Browser

(<http://neandertal.ensemblgenomes.org/>).

Exercise 18 – Structural variation in human

In the paper ‘The influence of *CCL3L1* gene-containing segmental duplications on HIV-1/AIDS susceptibility’ (Gonzalez *et al* Science. 2005 Mar 4; 307(5704):1434-40) it is shown that a higher copy number of the *CCL3L1* (Chemokine (C-C motif) ligand 3-like 1) gene is associated with lower susceptibility to HIV infection.

- (a) Find the human *CCL3L1* gene.
- (b) Have any CNVs been annotated for this gene? Note: In Ensembl, CNVs are classified as structural variants.

Exercise 19 – Exploring a SNP in mouse

Madsen *et al* in the paper ‘Altered metabolic signature in pre-diabetic NOD mice’ (PloS One. 2012; 7(4): e35445) have described several regulatory and coding SNPs, some of them in genes residing within the previously defined *insulin dependent diabetes (IDD)* regions. The authors describe that one of the identified SNPs in the murine *Xdh* gene (rs29522348) would lead to an amino acid substitution and could be damaging as predicted as by SIFT (<http://sift.jcvi.org/>).

- (a) Which chromosome and coordinates in the SNP located?
- (b) What is the HGVS recommendation nomenclature for this SNP?
- (c) Why does Ensembl put the C allele first (C/T)?
- (d) Are there differences between the genotypes reported in NOD/LTJ and BALB/cByJ?

Demo: The Variant Effect Predictor (VEP)

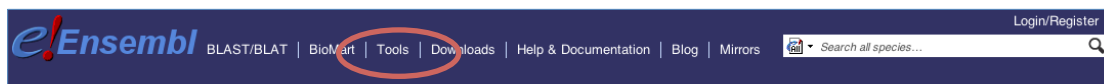
We have analysed a samples from a patient with a genetic disorder. The patient presents with facial and limb deformities, mental

retardation and gastrointestinal reflux. Our genotyping has identified a mutation that may be responsible for the phenotype:
An A->G mutation on chromosome 5 at 37,017,205 on the + strand.


We will use the **Ensembl VEP** to determine:


- Has my variant already been annotated in Ensembl?
- What genes are affected by my variant?
- Does my variant result in a protein change?

From any page in Ensembl click on **Tools** in the top bar.



This provides a table showing the tools that are available in Ensembl. There's a short description of what each of the tools do. All are available as both an online tool, and code that you can download. We're going to using the Variant Effect Predictor online tool.

Name	Description	Online tool	Download code
 Variant Effect Predictor	(Formerly SNP Effect Predictor). Analyse your own variants and predict the functional consequences of known and unknown variants via our Variant Effect Predictor (VEP) tool. You can do this online (max 750 variants), or download the script. Accepted formats include a simple tab-delimited file, VCF and other formats. Uploaded tracks can be viewed on Location pages. See also full documentation .		
Assembly converter	Map your data to the current assembly. Accepted file formats: GFF , GTF , BED , PSL . N.B. Export is currently in GFF only		
ID History converter	Convert a set of Ensembl IDs from a previous release into their current equivalents (max 30 ids).		
Ensembl Virtual Machine	VirtualBox virtual Machine with Ubuntu desktop and pre-configured with the latest Ensembl API plus Variant Effect Predictor (VEP). NB: download is >1 GB . See also full documentation .	-	

Click on the spanner  beside the **Variant Effect Predictor (VEP)**.

This will open up a dialogue box. This allows us to input data on our variant.

The screenshot shows the Variant Effect Predictor (Ve!P) web interface. On the left is a navigation menu with options like 'Add your data', 'Attach DAS', 'Manage Data', and 'Features on Karyotype'. The main area is titled 'Variant Effect Predictor:' and contains a description of the tool, a 'Ve!P' logo, and a note about the 750-variant limit. Below this is the 'Input file' section with fields for 'Species' (set to 'Human (Homo sapiens): GRCh37'), 'Name for this data (optional)', and 'Paste data:'. The 'Paste data:' field contains the text: '1 881907 881906 -/C +', '5 140532 140532 T/C +'. There are also 'Upload file:' and 'or provide file URL:' options, and an 'Input file format:' dropdown set to 'Ensembl default'. At the bottom is an 'Options' section with radio buttons for 'Transcript database to use: Ensembl transcripts' (selected) and 'RefSeq and other transcripts'. Three callout boxes with arrows point to the 'Name for this data' field, the 'Paste data' field, and the 'Upload file' button, containing the text: 'Give your data a name', 'Put your data in here.', and 'You can also upload a file.' respectively.

The data is in the format:

Chromosome Start End alleles (reference/mutation) strand

Delete the writing already in the [Paste data](#) box and type in:

5 37017205 37017205 A/G +

Scroll down to see some of the options we can also choose.

Options

Transcript database to use: Ensembl transcripts RefSeq and other transcripts

Get regulatory region consequences (human and mouse only):

Type of consequences to display:

Check for existing co-located variants:

Get 1000 Genomes global allele frequency for existing variants:

Return results for variants in coding regions only:

Show HGNC Identifier for genes where available:

Show Ensembl protein identifiers where available:

Show HGVS identifiers for variants where available:

Missense SNP predictions (human only)

SIFT predictions:

PolyPhen predictions:

Frequency filtering of existing variants (human only)

Filter variants by frequency:

NB: Enabling frequency filtering may be slow for large datasets. The default options will filter out common variants found by the 1000 Genomes project.

Filter:

Choose which database to map your variant to.

Find out if variants already exist in our database.

Choose to see scores for protein changes.

Choose to only see common or rare variants

Select **Prediction and Score for SIFT predictions** and **PolyPhen predictions**. These are algorithms that predict how deleterious a mutation will be on a protein.

When you've selected everything you need, scroll right to the bottom and click **Next**.

Click **HTML** to view your results with clickable links.

Uploaded Variation	Location	Allele	Gene	Feature	Feature type	Consequence	Position in cDNA	Position in CDS	Position in protein	Amino acid change	Codon change	Co-located Variation	Extra
5_37017205_A/G	5:37017205	G	ENSG00000164190 ENST00000448238	Transcript	missense_variant		5329	4861	1621	R/G	Aga/Gga	rs62654861	PolyPhen=probably_damaging(1); SIFT=deleterious(0); GMAF=-
5_37017205_A/G	5:37017205	G	ENSG00000164190 ENST00000282516	Transcript	missense_variant		5360	4861	1621	R/G	Aga/Gga	rs62654861	PolyPhen=probably_damaging(0.994); SIFT=deleterious(0); GMAF=-

Our mutation affects two transcripts of one gene

Our mutation causes an amino acid change

Our mutation is already in the Ensembl database

Exercise: The Variant Effect Predictor (VEP)

Exercise 20 – VEP

Resequencing of the genomic region of the human *CFTR* (cystic fibrosis transmembrane conductance regulator (ATP-binding cassette sub-family C, member 7) gene (ENSG00000001626) has revealed the following variants (alleles defined in the forward strand):

- G/A at 7:117,171,039
- T/C at 7:117,171,092
- T/C at 7:117,171,122

(a) Use the VEP tool in Ensembl and choose the options to see SIFT and PolyPhen predictions. Do these variants result in a change in the proteins encoded by any of the Ensembl genes? Which gene? Have the variants already been found?

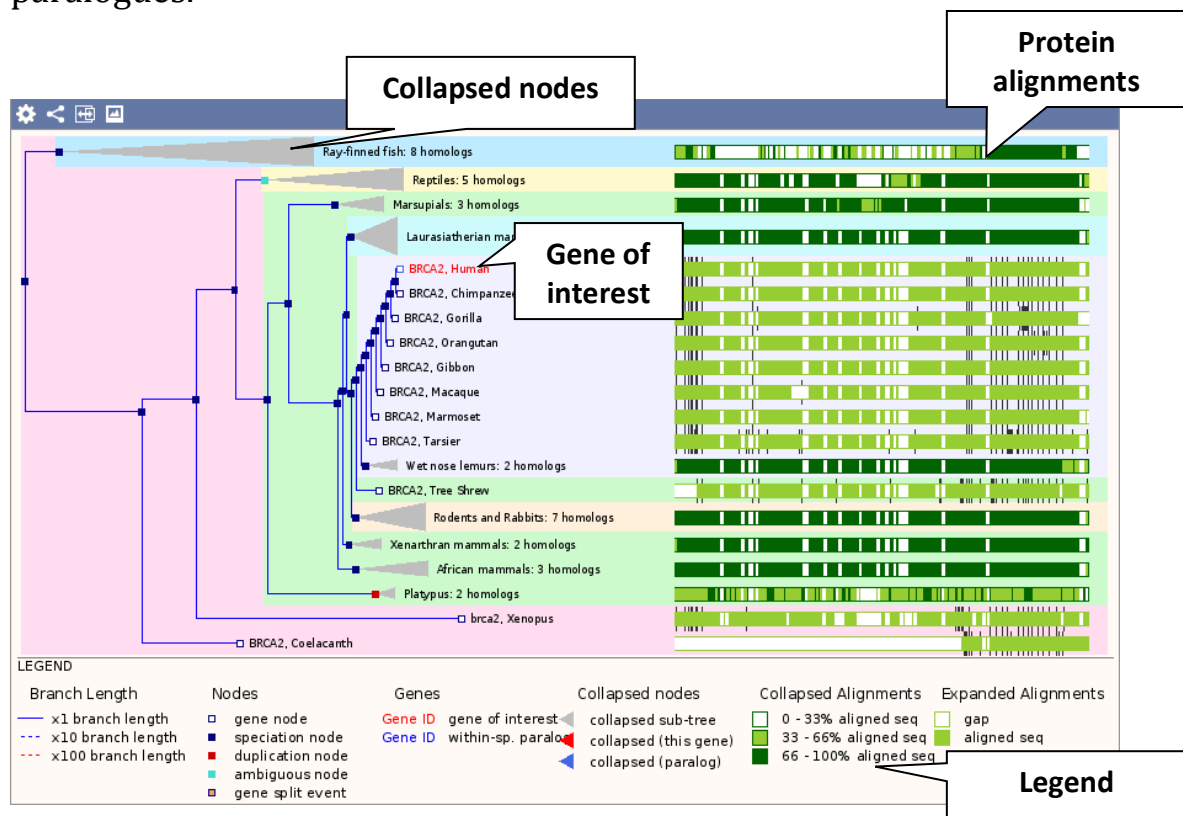
(b) Go to [Region in detail](#) for *CFTR*. Do you see the VEP track?

Comparative genomics

Demo: Gene trees and homologues

Let's look at the homologues of human *BRCA2*. Search for the gene and go to the **Gene** tab.

Click on [Gene tree \(image\)](#), which will display the current gene in the context of a phylogenetic tree used to determine orthologues and paralogues.



Funnels indicate collapsed nodes. We can expand them by clicking on the node and selecting [Expand this sub-tree](#) from the pop-up menu.

Taxon: Marsupials ~148 MYA (Marsupialia)

Gene Count 3

Branch Length 0.131357

Bootstrap 100

Type Speciation

Support phymi_nt,nj_ds,phymi_aa,nj_dn,nj_mm

Image [expand this sub-tree](#)

Image [expand all sub-trees](#)

Image [collapse other nodes](#)

Comparison [Jump to Region Comparison view](#)

View Sub-tree [Alignment: FASTA](#)

View Sub-tree [Tree: New Hampshire](#)

View Sub-tree [Expand for Jalview](#)

We can look at homologues in the [Orthologues](#) and [Paralogues](#) pages, which can be accessed from the left-hand menu. The numbers of orthologues or paralogues available are indicated in brackets alongside the name. If there are none, then the name will be greyed out. [Paralogues](#) is greyed out for *BRCA2* indicating that there are no paralogues available.

Click on [Orthologues](#) to see the 56 orthologues available.

Orthologues ⓘ

[View sequence alignments of all orthologues.](#)

Summary of orthologues of this gene

Click on 'Show' to display the orthologues for one or more groups, or click on 'Configure this page' to choose a custom list of species.

Species set	Show details	1:1	1:many	many:many	No orthologues
Primates Humans and other primates	<input type="checkbox"/>	9	0	0	0
Rodents Rodents, rabbits and related species	<input checked="" type="checkbox"/>	8	0	0	0
Laurasiatheria Carnivores, ungulates and insectivores	<input type="checkbox"/>	12	1	0	0
Placental Mammals All placental mammals	<input type="checkbox"/>	34	1	0	0
Sauropsida Reptiles	<input type="checkbox"/>	5	0	0	0
Amphibia Amphibians	<input type="checkbox"/>	9	0	0	0
Other vertebrates Including invertebrates	<input type="checkbox"/>	52	4		

Orthologue types

Choose a taxon of interest

Information on orthologues

Species	Type	dN/dS	Ensembl identifier & gene name	Compare	Location	Target %id	Query %id
Guinea Pig (<i>Cavia porcellus</i>)	1-to-1	0.43930	ENSCPOG00000005153 BRCA2 breast cancer 2, early onset [Source:HGNC SymbolAcc:1101]	<ul style="list-style-type: none"> Region Comparison Alignment (protein) Alignment (cDNA) Gene Tree (image) 	scaffold_6:33778275-33853154:1	65	63
Kangaroo rat (<i>Dipodomys ordii</i>)	1-to-1	n/a	ENSNDORG00000007049 Brca2 breast cancer 2 Gene [Source:MGI SymbolAcc:MGI:109337]	<ul style="list-style-type: none"> Region Comparison Alignment (protein) Alignment (cDNA) Gene Tree (image) 	GeneScaffold_1038:358641-411759:1	41	41
Mouse (<i>Mus musculus</i>)	1-to-1	0.41716	ENSMUSG00000041147 Brca2 No description	<ul style="list-style-type: none"> Region Comparison Alignment (protein) Alignment (cDNA) Gene Tree (image) 	5:150522630-150569746:1	56	54

Choose to see only **Rodent** orthologues by selecting the box. The table below will now only show details of rodent orthologues. Let's look at mouse.

Mouse (<i>Mus musculus</i>)	1-to-1	0.41716	ENSMUSG00000041147 Brca2 No description	<ul style="list-style-type: none"> Region Comparison Alignment (protein) Alignment (cDNA) Gene Tree (image) 	5:150522630-150569746:1	56	54
----------------------------------	--------	---------	---	---	---	----	----

Links from the orthologue allow you to go to alignments of the orthologous proteins and cDNAs. Click on [Alignment \(protein\)](#) for the mouse orthologue.

Orthologue alignment ⓘ

Orthologue type: 1 to 1 orthologue

Species	Gene ID	Peptide ID	Peptide length	% identity	Genomic location
Homo sapiens	ENSG00000139618	ENSP00000439902	3418 aa	50 %	13:32889611-32973805
Mus musculus	ENSMUSG00000041147	ENSMUSP00000038576	3329 aa	52 %	5:150522630-150569746

CLUSTAL W(1.81) multiple sequence alignment

```

ENSP00000439902/1-3418      MPIGSKERPTFFEIFKTRCNKADLGPIISLNWFEELSSEAPPYNSEPAEEF      KNNNYEPN
ENSMUSP00000038576/1-3329  MPVEYKRRPTFWEIFKARCSTADLGPIISLNWFEELSSEAPPYNSEPPPESEYKPHGYEPQ
**:  * .****:****:*. .*****:*****:*****:*****:*****:*****:
      89902/1-3418      LFKTPQRKP-SYNQLASTPIIFKEQGLTLPLYQSPVKELDKFKLDLGRNV-PNSR--HKS
      00038576/1-3329  LFKTPQRNPP-YHQFASTPIMFKERSQTLPLDQSPFREL-----GKV-VAS--SKHKT
*****:*  *:****:****:.. **** **.:**      * : ..      **:

```

Information on orthologue pair

Alignment in Clustal W format

Protein IDs

Exercises: Gene trees and homologues

Exercise 21 - Orthologues, paralogues and genetrees for the human *BRAF* gene.

(a) How many orthologues are predicted for this gene in primates? Note the Target %id and Query %id.

How much sequence identity does the *Tarsius syrichta* protein have to the human one? Click on the [Alignment](#) link next to the [Ensembl identifier](#) column to view a protein alignment in Clustal format.

(b) Go to the orthologue in marmoset. Is there a genomic alignment between marmoset and human? Is there a gene for both species in this region?

Exercise 22 – Zebrafish orthologues

Go to www.ensembl.org to find the *dbh* gene on the zebrafish genome.

(a) Go to the [Location](#) page for this gene. View the [Alignments \(image\)](#) and [Alignments \(text\)](#) for the 5 teleost fish. Which fish genomes are represented in the alignment? Do all the fish show a gene in these alignments?

(b) Export the alignments (as Clustal).

(c) Click on the [Region in detail](#) link at the left and turn on the tracks for [multiple alignments](#) and [conservation score](#) for the [5 teleost fish EPO](#) by configuring the page.

What is the difference between the [5 teleost fish EPO multiple alignment](#) track and the [Constrained elements](#) already turned on by default? Which regions of the gene, do most of the constrained element blocks match up to?

Can you find more information on how the constrained elements track was generated?

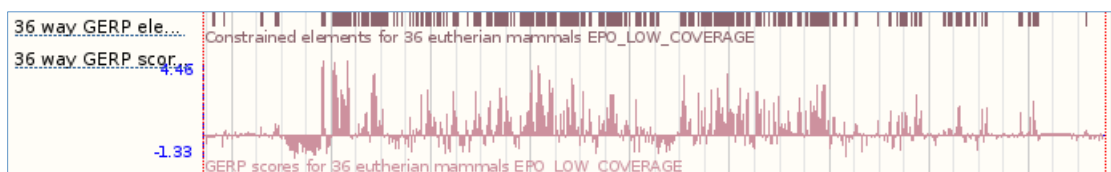
Demo: Whole genome alignments

Let's look at some of the comparative genomics views in the Location tab. Go to the region [2:176914144-177094980](#) in human, which contains the *HoxD* cluster which is involved in limb development and is highly conserved between species.

In the **Region in detail** view, we can already see the [Constrained elements for 36 eutherian mammals EPO_LOW_COVERAGE](#) track by default. This track indicates regions of high conservation between species, considered to be “constrained” by evolution.



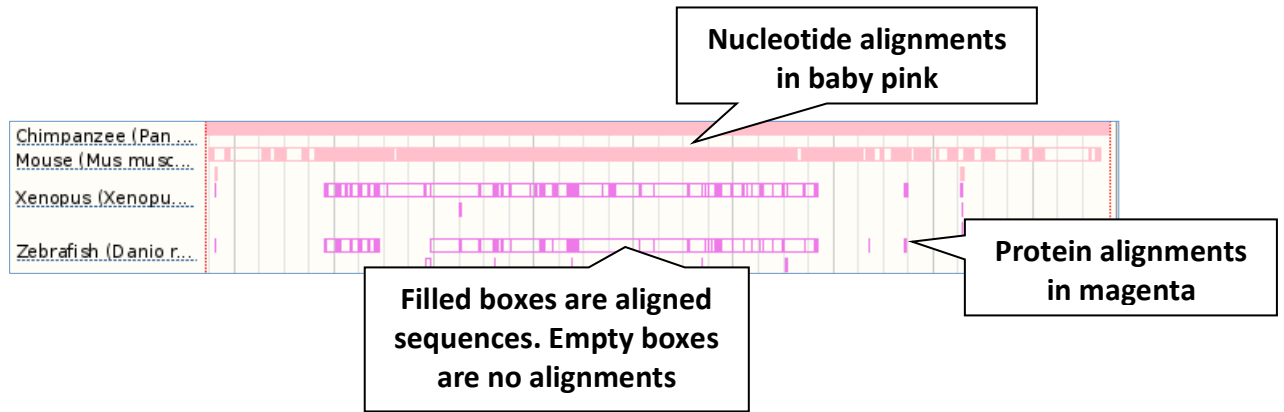
This track has a matching conservation score track. Click on [Configure this page](#), then [Comparative genomics](#) and turn on the track for [Conservation score for 36 eutherian mammals EPO_LOW_COVERAGE](#). Save and close the menu.



You can now see the conservation scores that were used to determine the peaks indicated in the constrained elements track.

We can also look at individual species comparative genomics tracks in this view by clicking on [Configure this page](#).

Select BLASTz/LASTz alignments from the left-hand menu to choose alignments between closely related species. Turn on the alignments for Mouse and Chimpanzee in Normal. Go to Translated blat alignments and turn on alignments with Zebrafish and Xenopus in Normal. Save and close the menu.



The alignment is greatest between closely related species.

We can also look at the alignment between species or groups of species as text. Click on [Alignments \(text\)](#) in the left hand menu.

Alignments (text)

Choose an alignment from the drop-down

Multiple alignments

Pairwise alignments

Alignment: -- Select an alignment --

Location: 6 primates EPO
13 eutherian mammals EPO
20 amniota vertebrates Pecan
36 eutherian mammals EPO LOW COVERAGE

Gene: Pairwise alignments

Key: Alpaca (Vicugna pacos) - blastz
Anole lizard (Anolis carolinensis) - translated blat
Armadillo (Dasypus novemcinctus) - blastz
Bushbaby (Otolemur garnettii) - lastz
Cat (Felis catus) - lastz
Chicken (Gallus gallus) - lastz
Chicken (Gallus gallus) - translated blat
Chimpanzee (Pan troglodytes) - lastz
Chinese softshell turtle (Pelodiscus sinensis) - lastz
Ciona intestinalis - translated blat
Ciona savignyi - translated blat
Cod (Gadus morhua) - translated blat
Coelacanth (Latimeria chalumnae) - translated blat
Cow (Bos taurus) - lastz
Dog (Canis lupus familiaris) - lastz
Dolphin (Tursiops truncatus) - blastz
Elephant (Loxodonta africana) - blastz

Homo sapiens: AATTACCCAGTCTCAGATAGTGTCTTTATATCAGTGT
Homo sapiens: TGATTA AAAAGGTA AATTTTATGTAACGAATATTTTA
Homo sapiens: GGCTCAGCCTGTAATCCCAGCACTTTGGGAGGCCGA
Homo sapiens: AAAAGTACAAAAAATTAGCCGGGCGTGGTGGCGGGCG
Homo sapiens: GCCACTGCACTCCAGCCTGGGCGACAGAGCGAGACTC
Homo sapiens: CTCTGGTTGACAGTCCTAGCTGATCTTGACCTFCCAG
Homo sapiens: GCTGCTAGGTTTCAGGATAATCTGTCTACAGCAATA
Homo sapiens: GGAGTGCTTGGACAGGCTTTGGGAAAAATTTGAGAAAT
Homo sapiens: TGGTAAACATACTTGACATACAAAGGCCAATTCAGTC

Select [Mouse](#) from the alignments list then click [Go](#).

You will see a list of the regions aligned, followed by the sequence alignment. Exons are shown in red.

This can also be viewed graphically. Click on [Alignments \(image\)](#) in the left-hand menu.

Alignment:

Location:

Gene:

Human region

Human

20.00 kb 40.00 kb 60.00 kb 80.00 kb 100.00 kb 120.00 kb 140.00 kb 187.32 kb

Genes (Merged E...)

HOXD10 > HOXD11 > HOXD9 > HOXD12 > HOXD8 > HOXD4 > HOXD13 > HOXD3 > AC009336.1 > MIR10B > AC009336.24 > HOXD1 >

Contigs

AC009336.13 > AC016739.5 <

Genes (Merged E...)

< EVX2 < HOXD-AS2 < HOXD-AS1 < AC016739.2

Gene Legend

processed transcript merged Ensembl/Havana RNA gene

Mouse

Genes (Merged E...)

Gm20440 > Gm14396 > Hoxd12 > Hoxd8 > Hoxd4 > Gm14424 > Hoxd13 > Hoxd10 > Hoxd3 > Gm14421 > Hoxd11 > Hoxd9 > Mir10b > Gm17511 >

Contigs

< Evx2 < 1700109F18Rik < 6720416L17Rik < Gm17511

Gene Legend

protein coding merged Ensembl/Havana processed transcript RNA gene

AlignSlice Legend

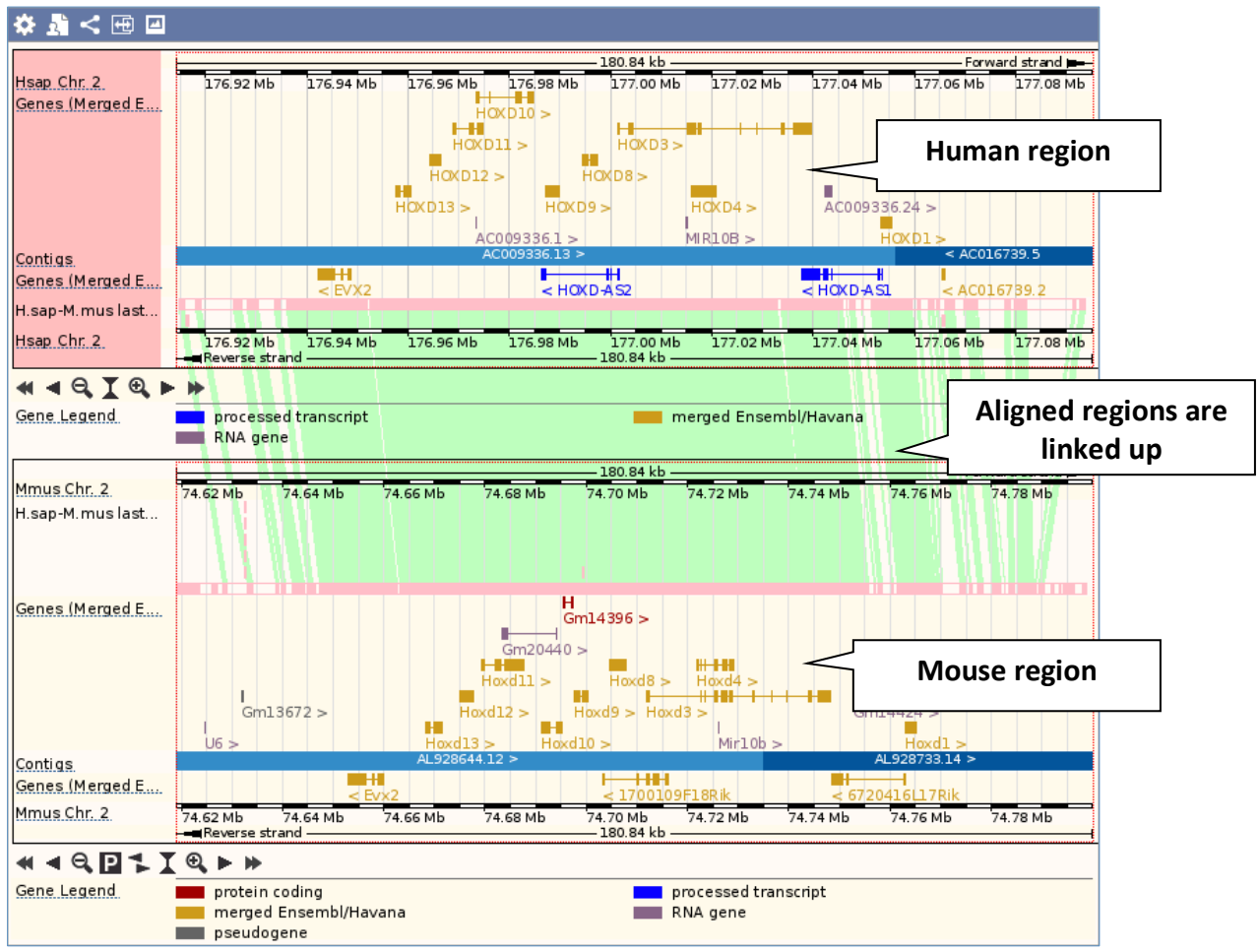
Breakpoint between different chromosomes Gap between two underlying slices

Mouse is already selected (from text view)

Mouse region, rearranged to align with human

In both alignment views the contig the compared species is rearranged to align to the species of interest. To compare with both contigs in their natural order, go to [Region comparison](#).

To add species to this view, click on the blue [Select species or regions](#) button. Choose [Mouse](#) from the list then close the menu.



We can view large scale syntenic regions from our chromosome of interest. Click on [Synteny](#) in the left hand menu.

Synteny between Human chromosome 2 and Mouse

Change Species: Mouse Go
Change Chromosome: 2 Go

Human chromosome

Mouse chromosome with syntenic region

Syntenic regions

Region of interest

Table of syntenic genes

◀ 15 upstream genes Navigate homology

Show/hide columns Filter

Homo sapiens genes	Location		Mus musculus homologues	Location	
EVX2 (ENSG00000174279)	2:176942200-176948641	→	Evx2 (ENSMUSG00000001815)	2:74652997-74659861	Region Comparison
HOXD13 (ENSG00000128714)	2:176957619-176960666	→	Hoxd13 (ENSMUSG00000001819)	2:74668310-74671599	Region Comparison

Exercises: Whole genome alignments

Exercise 23 – Synteny

Go to www.ensembl.org

Find the Rhodopsin (*RHO*) gene for Human. Go to the [Location](#) tab.

(a) Click [Synteny](#) at the left. Are there any syntenic regions in dog? If so, which chromosomes are shown in this view?

(b) Stay in the [Synteny](#) view. Is there a homologue in dog for human *RHO*? Are there more genes in this syntenic block with homologues?

Exercise 24 – Whole genome alignments

- (a) Find the Ensembl *BRCA2* (Breast cancer type 2 susceptibility protein) gene for human and go to the [Region in detail](#) page.
- (b) Turn on the [BLASTZ alignment](#) tracks for chicken, chimp, mouse and platypus and the [Translated BLAT alignment](#) tracks for anole lizard and zebrafish. Does the degree of conservation between human and the various other species reflect their evolutionary relationship? Which parts of the *BRCA2* gene seem to be the most conserved? Did you expect this?
- (c) Have a look at the [Conservation score](#) and [Constrained elements](#) tracks for the set of 36 mammals and the set of 19 vertebrates. Do these tracks confirm what you already saw in the tracks with pairwise alignment data?
- (d) Retrieve the genomic alignment for a constrained element. Highlight the bases that match in >50% of the species in the alignment.
- (e) Retrieve the genomic alignment for the *BRCA2* gene for primates. Highlight the bases that match in >50% of the species in the alignment.

Regulation

Demo: Raw ChIPSeq data

We're going to add some regulation data to the **Region in detail** view. We'll start at the human region **11:2012486-2030153**, which contains the imprinted *H19* gene.

Add regulation tracks using [Configure this page](#). First, we're going to add ChIP-seq data for histone modifications and polymerase binding. Click on [Histones & polymerases](#) under [Regulation](#) in the left-hand menu.

Regulation

Histone modifications & RNA polymerases ⓘ

Filter by: All classes (dropdown)
Enter cell or evidence type (input)

Key: On (dark blue), Off (light blue), No Data (grey), Filtered: On (dark green), Off (light green)

Cell lines: CD4, GM06990, GM12878, H1ESC, HMEC, HSMN, HUVEC, HeLa-S3, HepG2, IMR90, K562, NH-A, NHEK

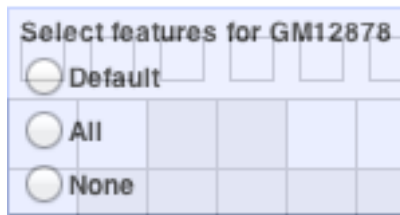
Track style: [Enable/disable all](#)

	CD4	GM06990	GM12878	H1ESC	HMEC	HSMN	HUVEC	HeLa-S3	HepG2	IMR90	K562	NH-A	NHEK
PolII	On	On	On	On	On	On	On	On	On	On	On	On	On
PolIII	On	On	On	On	On	On	On	On	On	On	On	On	On
H2AK5ac	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H2AK9ac	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H2AZ	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H4K20ac	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H4K27ac	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H3K4me1	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H2BK20ac	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H2BK5ac	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H2BK5me1	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H3K14ac	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H3K18ac	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off
H3K23ac	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off	Off

Callouts: Show tutorial, Add tutorial labels to help use this view, Legend, Cell lines, Choose track styles, Select boxes, Histone modifications

You can turn on a single track by clicking on the box in the matrix. Note that certain tracks are selected for all cell lines by default (PolII, PolIII, H3K27me3, H3K36me3, H3K4me3, H3K9me3). These will appear in the Region in detail view only if you specify a track style for the cell lines.

Turn on all the tracks for [GM12878](#). Hover over the cell line name then select [All](#).

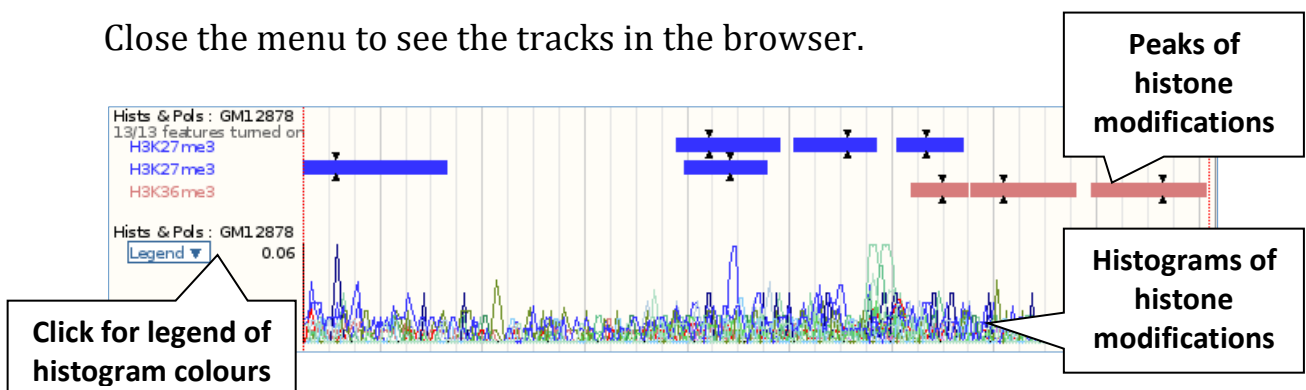


Now choose the track style for the tracks you've switched on. Click on the track style box for GM12878 and select [Both](#).



There is a similar matrix for [Open chromatin & TFBS](#). Use this to turn on all tracks for [GM12878](#) in [Both](#).

Close the menu to see the tracks in the browser.

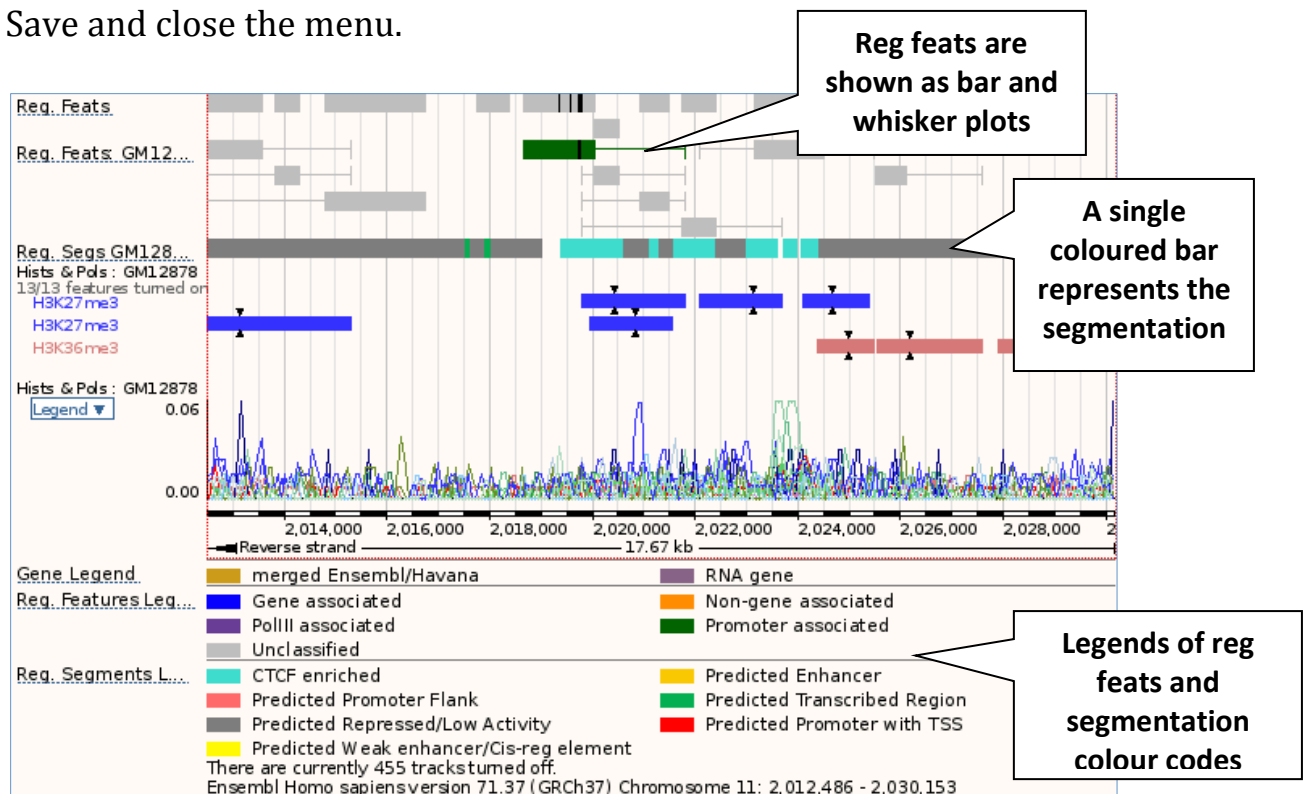


Demo: Regulatory features and segmentation

These data are used to construct the **Reg-feats** and **Segmentation features**. The merged Reg-feats are switched on in the Region in detail view by default.

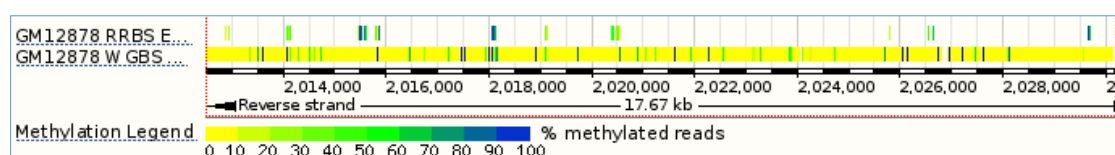
Click on [Configure this page](#). Then select [Regulatory features](#). Turn on the [Reg. Feats: GM12878](#) and [Reg. Segs: GM12878](#) tracks.

Save and close the menu.



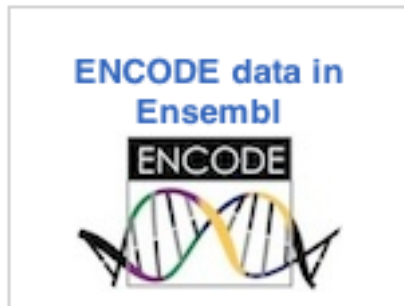
Can you see correlations between the different kinds of regulatory data representation?

You can also add methylation data using [Configure this page](#). Find it under [DNA methylation](#) and turn on [GM12878 RRBS ENCODE](#) and [GM12878 WGBS ENCODE](#).



Our regulatory data incorporates the ENCODE data. To see the raw ENCODE data and the ENCODE segmentation, you need to add the ENCODE hub.

From ensembl.org, click on the [ENCODE icon](#).



This page contains information about the ENCODE data and how it is incorporated into Ensembl.

Add the ENCODE hub by clicking on the [Link to add the ENCODE track hub](#).

This will take you directly to the matrices for adding ENCODE data to the Region in detail view. The ENCODE matrices work in the same way as the Open chromatin & TFBS and Histones & polymerases matrices, except that some have multiple options (indicated by numbers within the boxes).

Exercises: Regulation

Exercise 25 – Gene regulation: Human *STX7*

(a) Find the Location tab ([Region in detail page](#)) for the *STX7* gene. Are there regulatory features in this gene region? If so, where in the gene do they appear?

(b) Click [Configure this page](#) and on the [Regulatory features](#) menu in the left hand side. Turn on Segmentation features for [HUVEC](#), [HeLa-S3](#), and [HepG2](#) cell types. Do any of these cells show predicted enhancer regions in the *STX7* region?

(c) Use [Configure this page](#) to add supporting data indicating open chromatin for HeLa-S3 cells. Are there sites enriched for marks of

open chromatin (DNase1 and FAIRE) in HeLa cells at the 5' end of *STX7*?

(d) Configure this page once again to add histone modification supporting data for the same cell type as above (e.g.HeLa-S3). Which ones are present at the 5' end of *STX7*?

(e) Is there any data to support methylated CpG sites in this region (5' end) of *STX7* in B-cells?

(f) Create a [Share](#) link for this display. Email it to yourself then open the link.

Exercise 26 – Regulatory features in human

The *HLA-DRB1* and *HLA-DQA1* genes are part of the human major histocompatibility complex class II (MHC-II) region and are located about 44 kb from each other on chromosome 6. In the paper 'The human major histocompatibility complex class II *HLA-DRB1* and *HLA-DQA1* genes are separated by a CTCF-binding enhancer-blocking element' (Majumder *et al* J Biol Chem. 2006 Jul 7;281(27):18435-43) a region of high acetylation located in the intergenic sequences between *HLA-DRB1* and *HLA-DQA1* is described. This region, termed XL9, coincided with sequences that bound the insulator protein CCCTC-binding factor (CTCF). Majumder *et al* hypothesise that the XL9 region may have evolved to separate the transcriptional units of the *HLA-DR* and *HLA-DQ* genes.

(a) Go to the region from 32,540,000 to 32,620,000 bp on human chromosome 6

(b) Is there a regulatory feature annotated in the intergenic region between the *HLA-DRB1* and *HLA-DQA1* genes that has CTCF binding supporting data as (part of) its core evidence?

(c) Has the CTCF binding detected at this position been observed in all cell/tissue types analysed?

(d) Have a look at the [Regulatory supporting evidence - Histones & Polymerases](#) configuration matrix. For which cell/tissue type are the most histone acetylation data sets available? In this cell/tissue type,

is the region that shows CTCF binding also a region of high acetylation, as found by Majumder *et al*?

Quick Guide to Databases and Projects

Here is a list of databases and projects you will come across in these exercises. Google any of these to learn more. Projects include many species, unless otherwise noted.

Other help:

The Ensembl Glossary: <http://www.ensembl.org/Help/Glossary>

Ensembl FAQs:

<http://www.ensembl.org/Help/Faq>

SEQUENCES

EMBL-Bank, NCBI GenBank, DDBJ – Contain nucleic acid sequences deposited by submitters such as wet-lab biologists and gene sequencing projects. These three databases are synchronised with each other every day, so the same sequences should be found in each.

CCDS – coding sequences that are agreed upon by Ensembl, VEGA-Havana, UCSC, and NCBI. (*human and mouse*).

NCBI Entrez Gene – NCBI's gene collection

NCBI RefSeq – NCBI's collection of 'reference sequences', includes genomic DNA, transcripts and proteins. NM stands for 'Known mRNA' (eg NM_005476) and NP (eg NP_005467) are 'Known proteins'.

UniProtKB – the "Protein knowledgebase", a comprehensive set of protein sequences. Divided into two parts: Swiss-Prot and TrEMBL

UniProt Swiss-Prot – the manually annotated, reviewed protein sequences in the UniProtKB. High quality.

UniProt TrEMBL – the automatically annotated, unreviewed set of proteins (EMBL-Bank translated). Varying quality.

VEGA – Vertebrate Genome Annotation, a selection of manually-curated genes, transcripts, and proteins. (*human, mouse, zebrafish, gorilla, wallaby, pig, and dog*).

VEGA-HAVANA – The main contributor to the VEGA project, located at the Wellcome Trust Sanger Institute, Hinxton, UK.

GENE NAMES

HGNC – HUGO Gene Nomenclature Committee, a project assigning a unique and meaningful name and symbol to every human gene. (*Human*).

ZFIN – The Zebrafish Model Organism Database. Gene names are only one part of this project. (*Z-fish*).

PROTEIN SIGNATURES

InterPro – A collection of domains, motifs, and other protein signatures. Protein signature records are extensive, and combine information from individual projects such as UniProt, along with other databases such as SMART, PFAM and PROSITE (explained below).

PFAM – A collection of protein families

PROSITE – A collection of protein domains, families, and functional sites.

SMART – A collection of evolutionarily conserved protein domains.

OTHER PROJECTS

NCBI dbSNP – A collection of sequence polymorphisms; mainly single nucleotide polymorphisms, along with insertion-deletions.

NCBI OMIM – Online Mendelian Inheritance in Man – a resource showing phenotypes and diseases related to genes (*human*).