

Visualisation tools for next-generation sequencing

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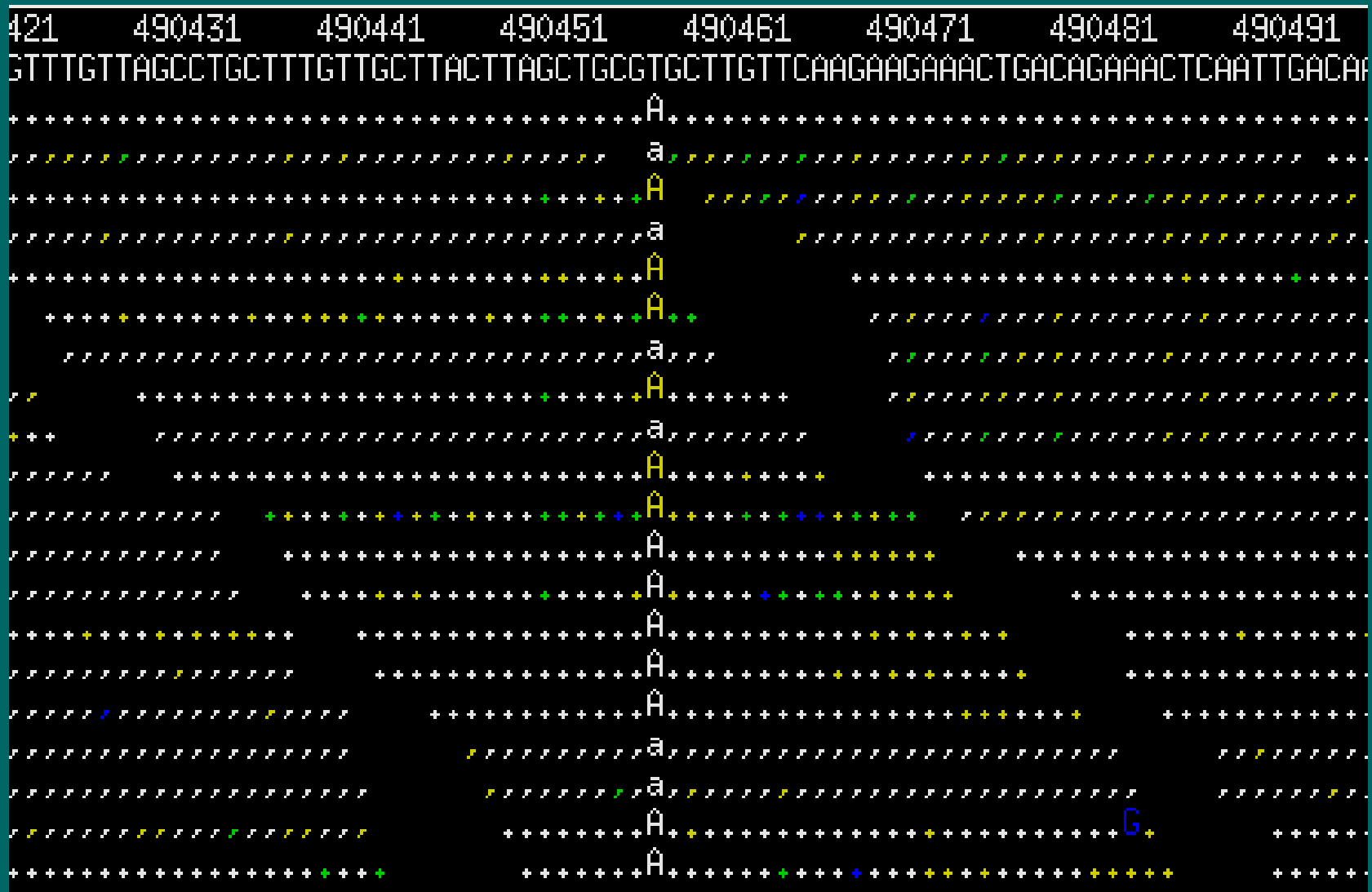
EMBL-EBI



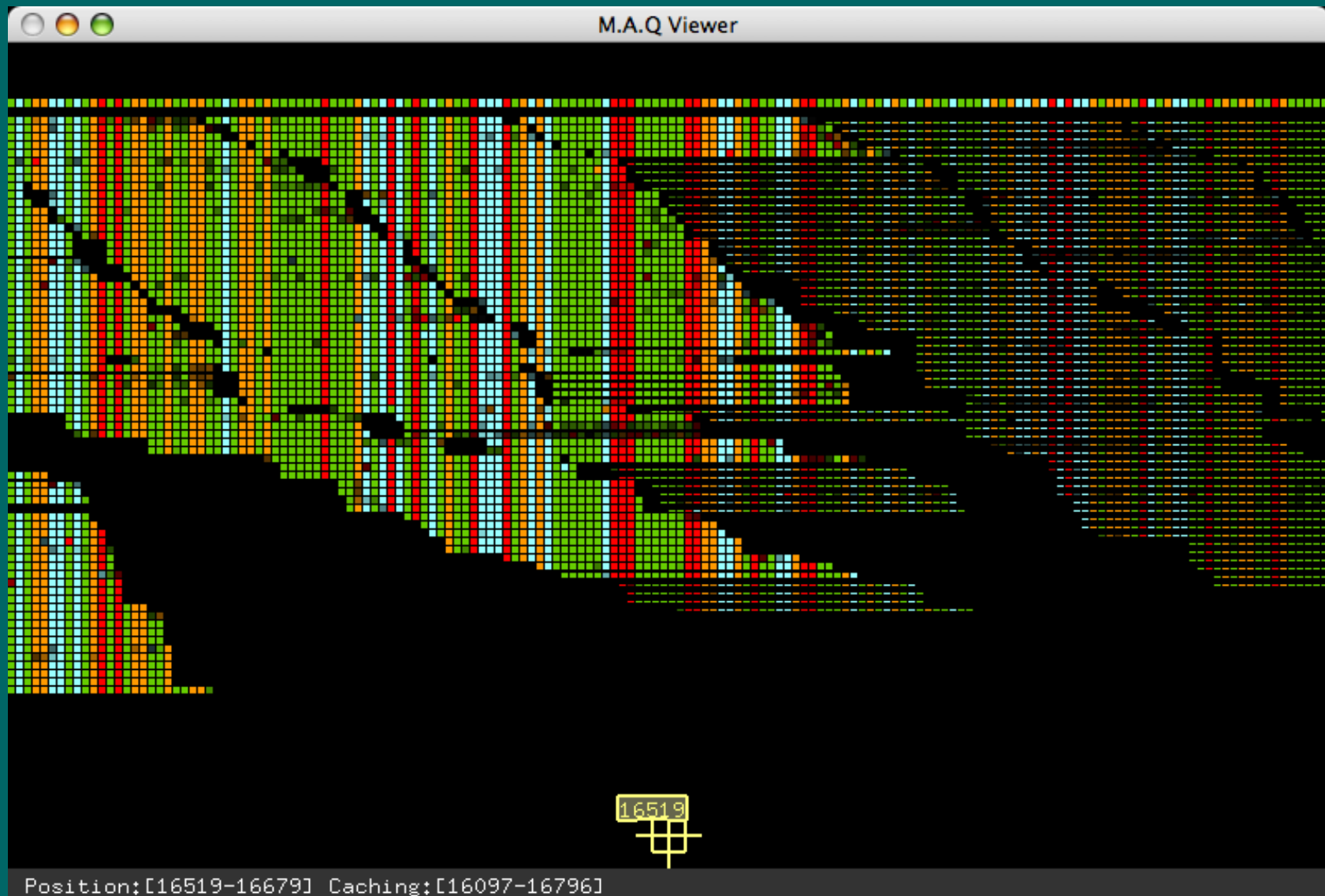
Outline

- Exploring and checking alignment with alignment viewers
- Using genome browsers
- Getting an overview over the whole data with Hilbert curve visualization
- Displaying peaks alongside feature annotation with the “GenomeGraph” package

Alignment Viewers: SAMtools tview



Alignment viewers: MaqView



Alignment Viewer: MapView

The screenshot displays the MapView 3.0 interface for a paired-end alignment on chromosome 12. The main window shows a sequence alignment with a vertical blue line indicating the current position. A popup window provides details for the variant at position 20697: Reference Position: 20697, Coverage SUM: 27, A: 2, T: 14, G: 11, C: 0, N: 0, and Variant Frequency: 0.4400.

Fast Positioning
Col: 20642
Row: 1

SNP Detection
Solexa quality score
Quality Score >= 20
Variant Frequency >= 0.4000
Coverage SUM >= 12

SNP Detection
 No quality control (FAST MODE)
Total: 18,356
Current: 22

Structure Variation
Distance >= 5000
Total: 15,376

Selection Details
Reference Position: 20697 Coverage SUM:27 A:2 T:14 G:11 C:0 N:0 Variant Frequency:0.4400

Statistics ON: 11,863,620 (A:3,386,857 T:3,379,903 G:2,545,229 C:2,551,630 N:1) 2,567,678 [+] 1,284,493 [-] 1,283,185 [contains 'N'] 40,791

Status: 1 - 35 | 20642 - 20752 | Row: --- Col: ---

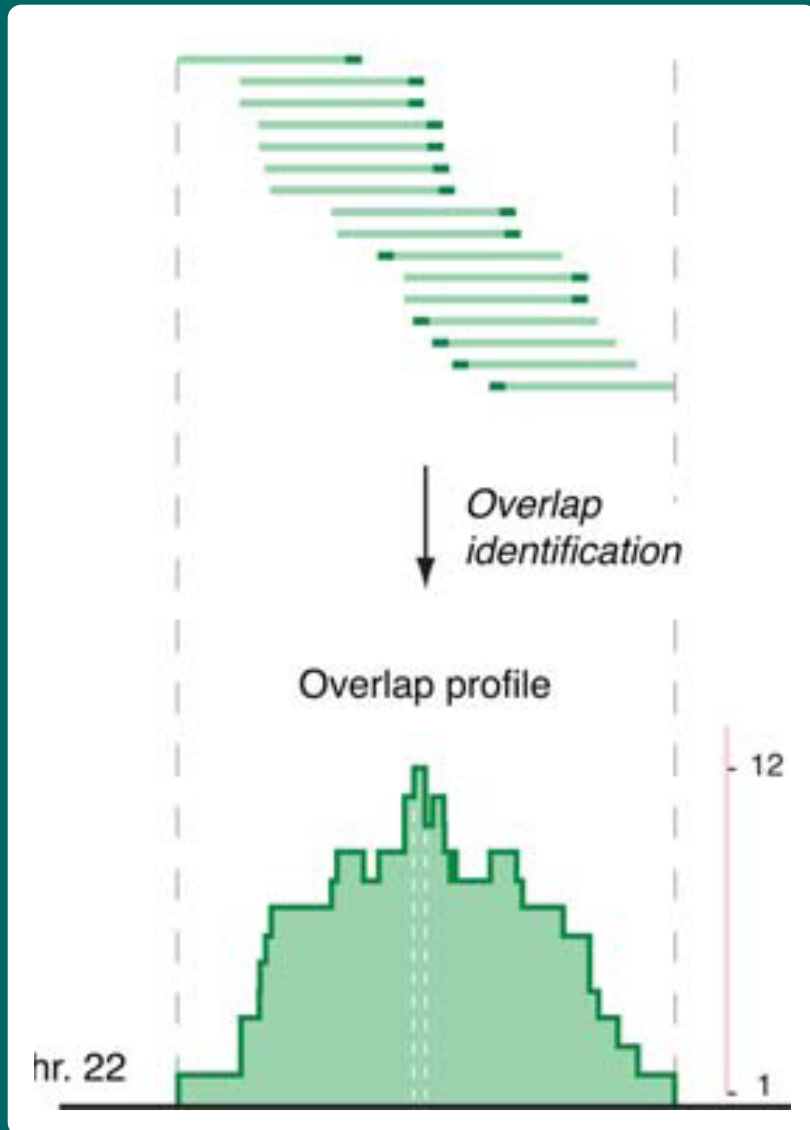
SAMtools pileup format

```

I      25514   G      G      42      0      25      5      .....^:.
I      25515   T      T      42      0      25      5      .....
I      25516   A      G      48      48     25      7      GGGGG^:G^:g
I      25517   G      G      51      0      25      8      ...../ ^: /
I      25518   T      T      60      0      25     11     ...../ / ^: . ^: / ^: /
I      25519   T      T      60      0      25     11     ...../ / / /
I      25520   G      G      60      0      25     11     ...../ / / /
I      25521   T      T      60      0      25     11     ...../ / / /
I      25522   A      A      60      0      25     11     ...../ / / /
I      25523   A      A      72      0      25     15     ...../ / / / ^: . ^: / ^: / ^: .
I      25524   C      C      72      0      25     15     ...../ / / / / .
I      25525   C      C      56      0      24     18     ...../ / / / / / ^: / ^: ! . ^: T
I      25526   A      A      81      0      24     18     ...../ / / / / / / .
I      25527   A      A      56      0      24     18     ...../ / / / / / / .G
          CCCCC
          CC?CC
          CCCCC5
          CCCCC1?
          CCCCC3A<:;
          CCCCC>A@AA
          CCCACC>A@<A
          CCCCC?ACAA
          CCCCC>ACAA
          CCCCC;ACAAC??C
          CCCCC6<<A?C=9C
          CCCCC>ACA?C=AC<
          CCCCC>ACAACAACA
          CCCCC?ACAA@A?CA
    
```

Fields: chromosome, position, reference base, consensus base, consensus quality, SNP quality, maximum mapping quality, coverage, base pile-up, base quality pile-up

Coverage vectors



<-- Solexa reads,
aligned to genome

<-- coverage vector

Coverage vectors

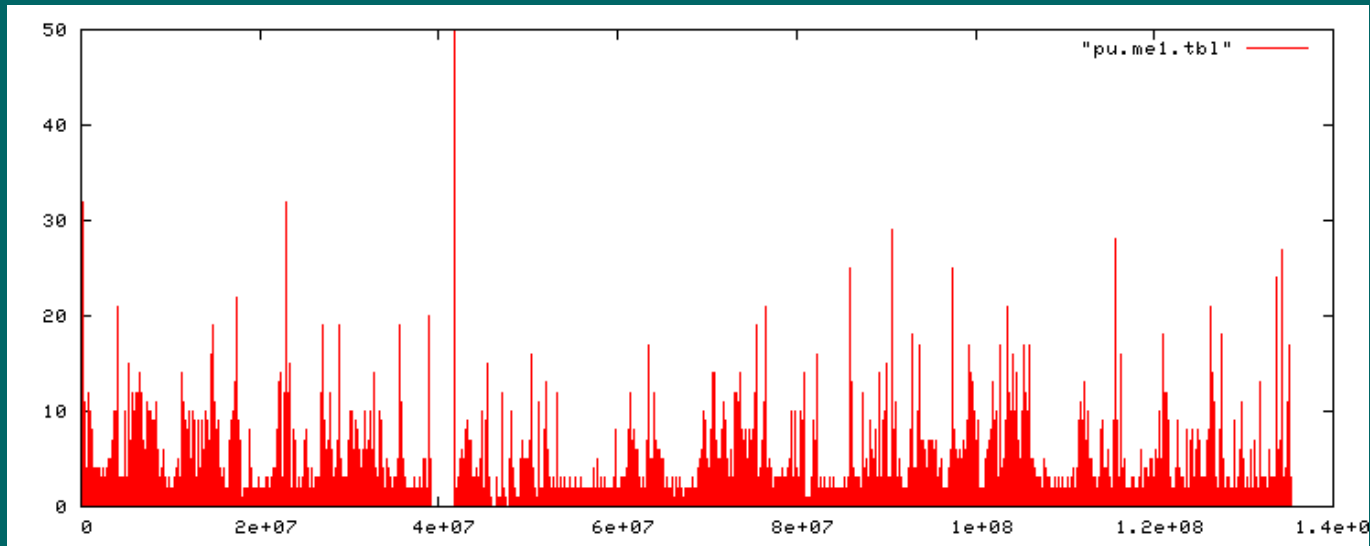
- A coverage vector (or "pile-up" vector) is an integer vector with one element per base pair in a chromosome, tallying the number of reads (or fragments) mapping onto each base pair.
- It is the essential intermediate data type in assays like ChIP-Seq or RNA-Seq
- Visualising coverage vectors is non-trivial, but essential for
 - quality control
 - hypothesis forming
 - etc.

Example: Histone modifications

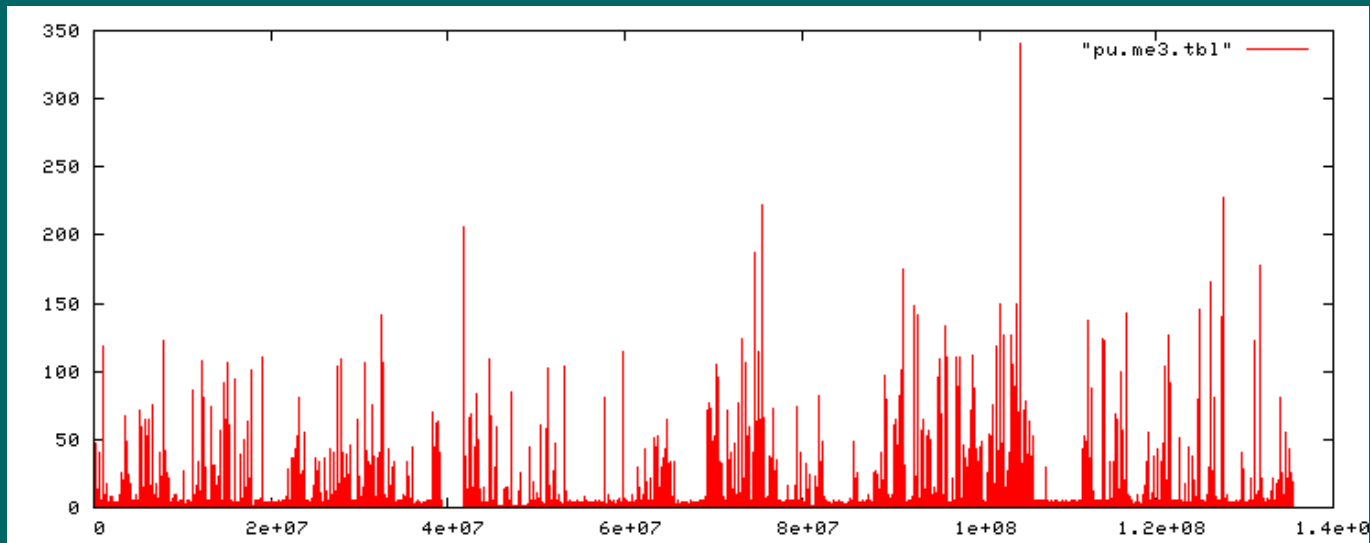
- Barski et al. (Cell, 2007) have studied histone modification in the human genome with ChIP-Seq
- I use their data for H3K4me1 and H3K4me3 as example data.
- (Each data set is from two or three Solexa lanes)

Coverage vector for a full chromosome (chr10)

H3K4me1

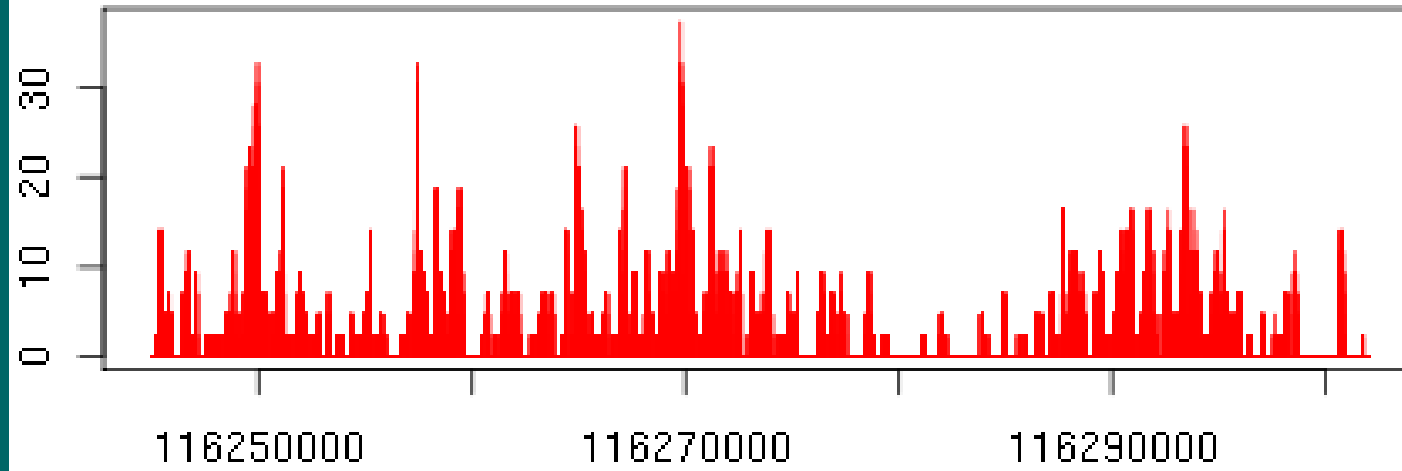


H3K4me3

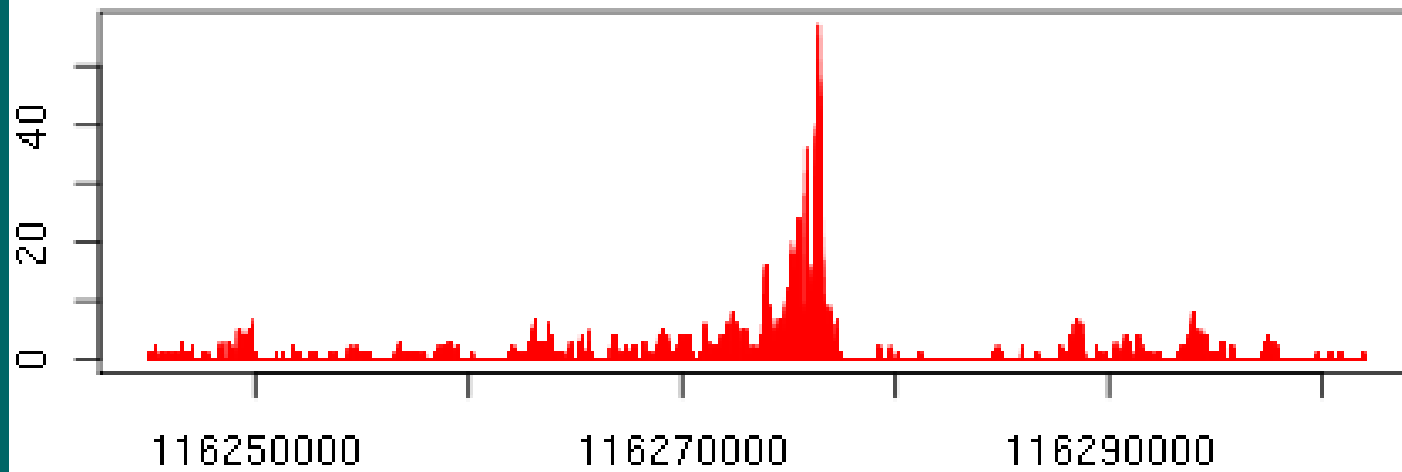


Zoom in

H3K4me1



H3K4me3



Genome browser tracks

Tracks may contain

- Features (intervals with name)
 - without score
 - with score
- vectors (continuously varying score)

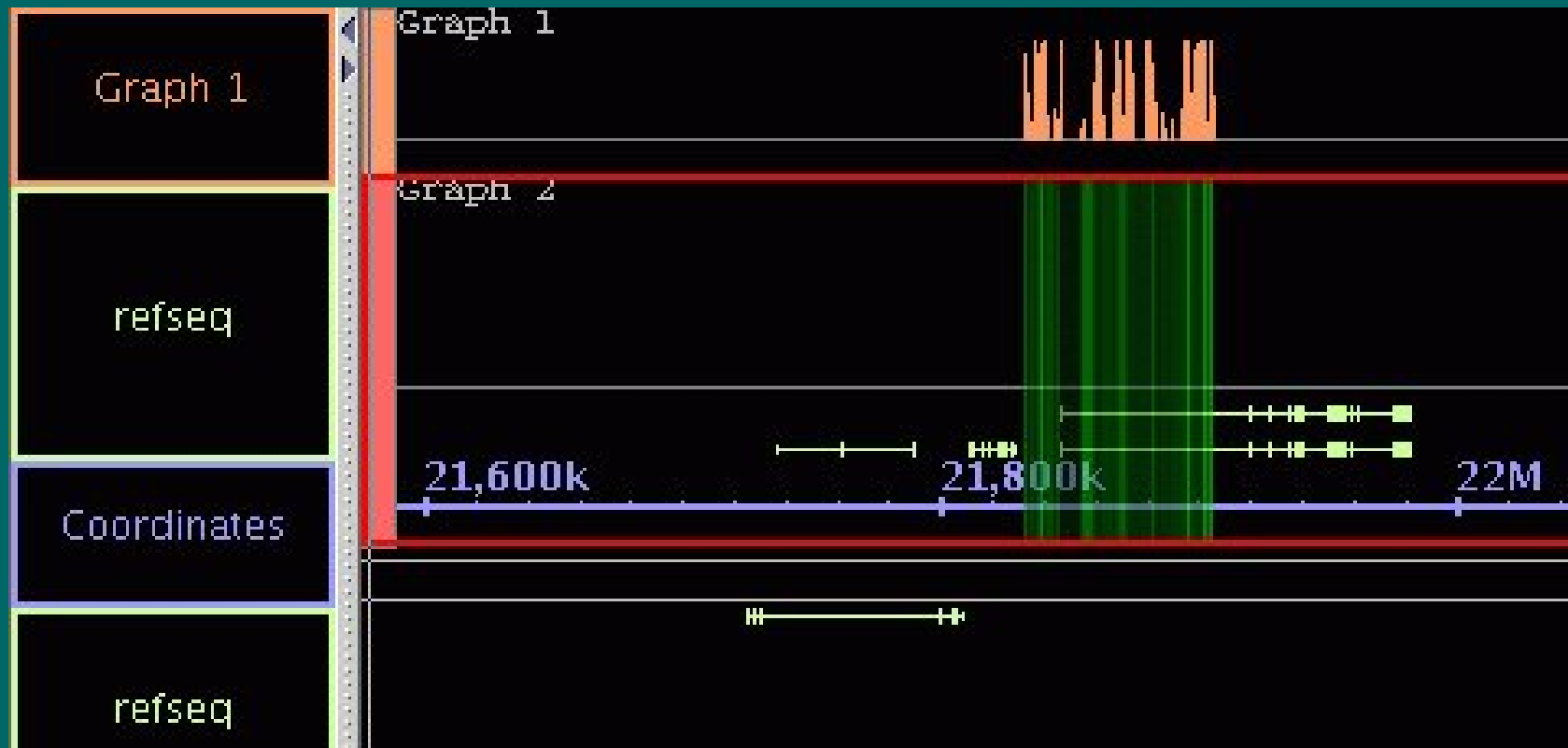
Standard formats for genome browser tracks

- BED
- GFF
- Wiggle fixedStep and variableStep

Displaying tracks alongside annotation

- Either, upload your track file to a web-base browser
 - UCSC genome browse
 - Ensembl genome browser
- or use a stand-alone browser on your desktop computer
 - Integrated Genome Browser (IGB) [Genoviz]
 - Argo Genome Browser [Broad Institute]
 - Artemis [Sanger Institute]

Displaying large amounts of data requires patience and lots of RAM. Not all tools handle it well.



rtracklayer: Bioconductor package by M. Lawrence (FHCRC)

- import and export BED, Wiggle, and GFF files
- manipulate track data and get sub-views
- directly interact with a genome browser (UCSC or Argo) to drive displaying of track data

Difference between the track formats

- Formats for feature-by-feature data:
 - BED
 - GFF
- Formats for base-by-base scores
 - Wiggle
 - BedGraph
- Wiggle has three sub-types:
 - [BED-like]
 - variableStep
 - fixedStep

Wiggle format: variableStep and fixedStep

```
browser position chr19:59304200-59310700
browser hide all
track type=wiggle_0 name="varStepTrack" description="varStep example" \
  visibility=full autoScale=off viewLimits=0.0:25.0 color=50,150,255 \
  yLineMark=11.76 yLineOnOff=on priority=10
variableStep chrom=chr19 span=150
59304701 10.0
59304901 12.5
59305401 15.0
59305601 17.5
59305901 20.0
59306081 17.5
59306301 15.0
59307871 10.0
track type=wiggle_0 name="fixedStepTrack" description="fixedStep examle"
fixedStep chrom=chr19 start=59307401 step=300 span=200
1000
 900
 800
 700
 600
 500
 400
 300
 200
 100
```

All coordinates 1-based!

bedGraph format

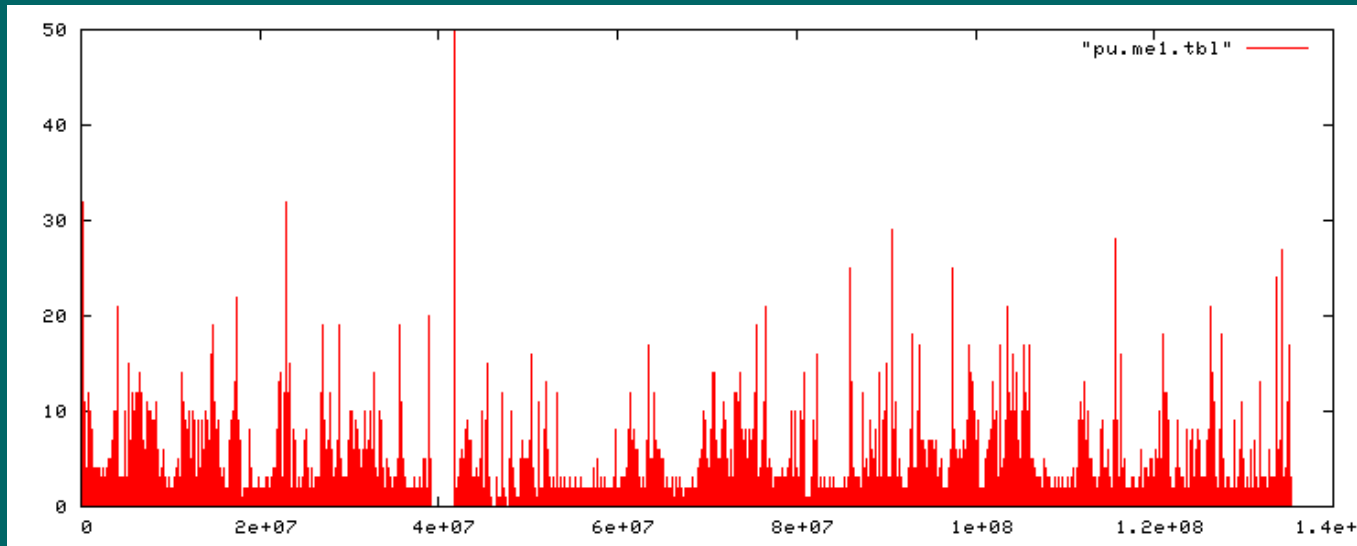
```
track type=bedGraph name="BedGraph Track"  
chr19 59302000 59302300 -1.0  
chr19 59302300 59302600 -0.75  
chr19 59302600 59302900 -0.50  
chr19 59302900 59303200 -0.25  
chr19 59303200 59303500 0.0  
chr19 59303500 59303800 0.25  
chr19 59303800 59304100 0.50  
chr19 59304100 59304400 0.75  
chr19 59304400 59304700 1.00
```

All coordinates 0-based, half-open!

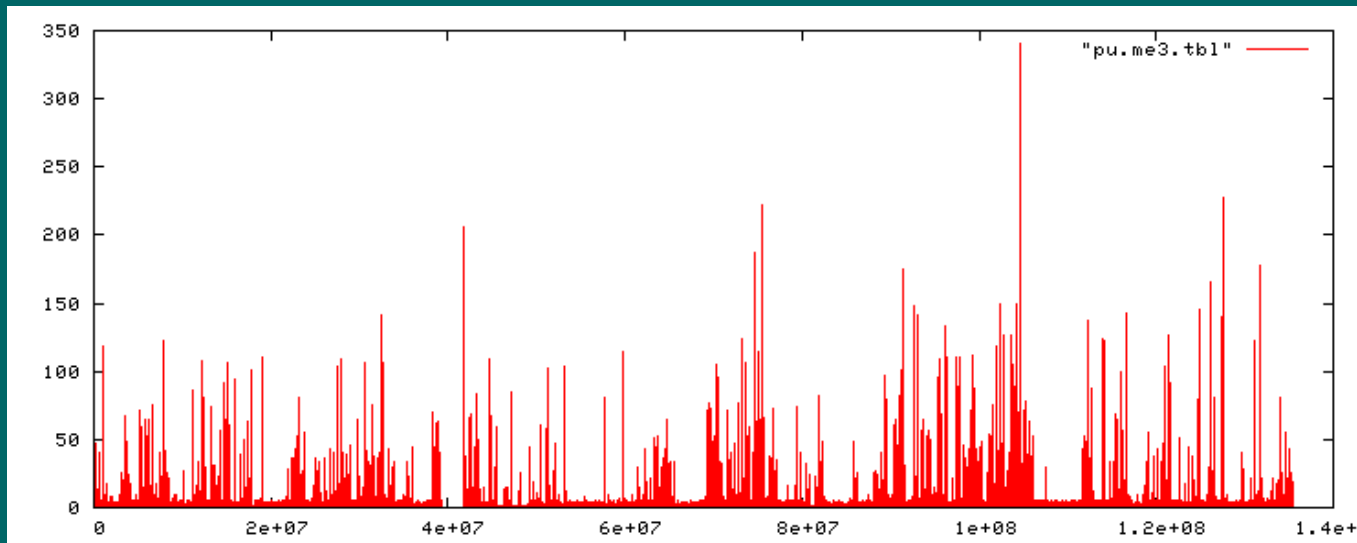
Specs: See UCSC Genome Browser web site

Back to the bird's eyes view

H3K4me1



H3K4me3



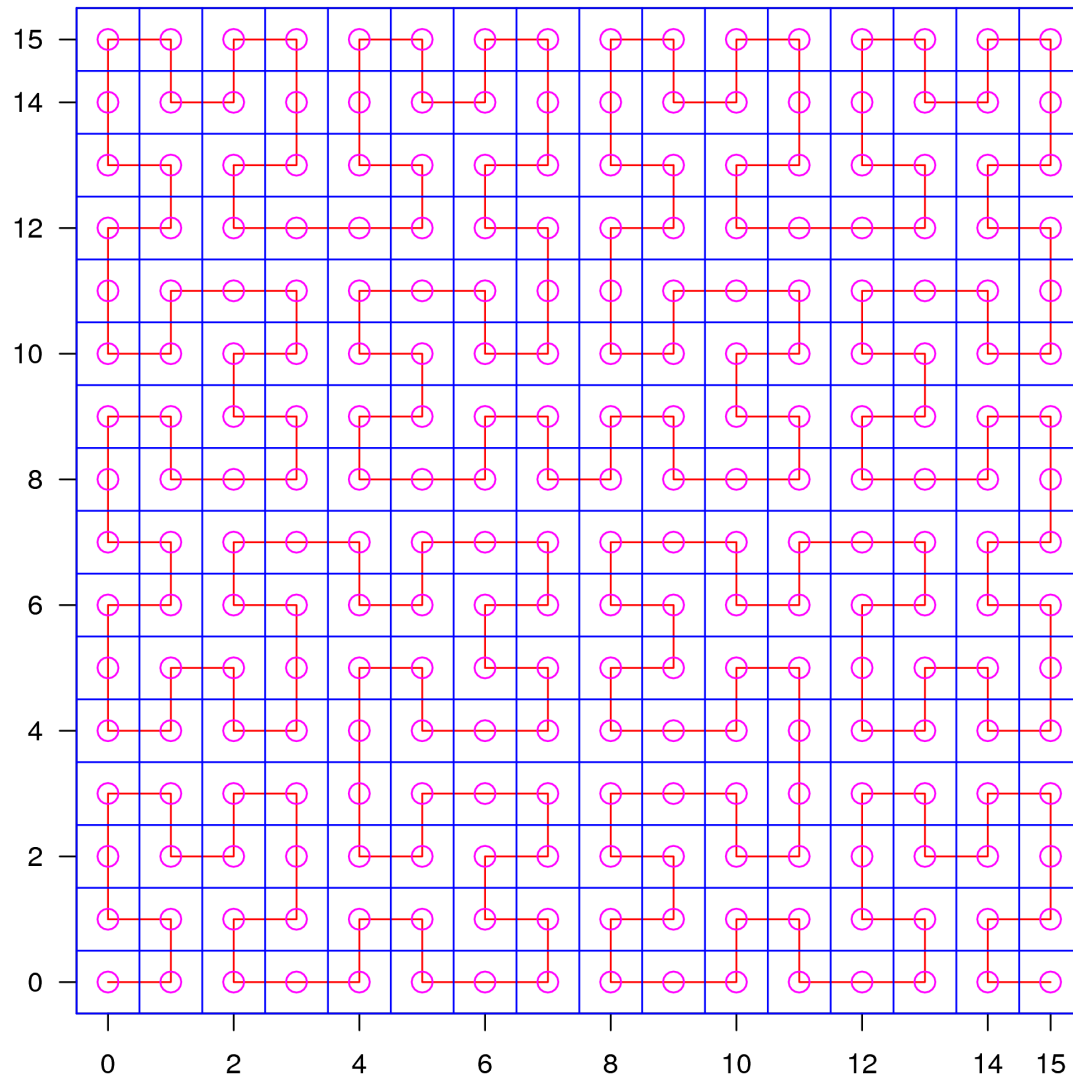
H3K4me
1

We need a way to get a general overview on the data without either not seeing any details not getting lost in them.

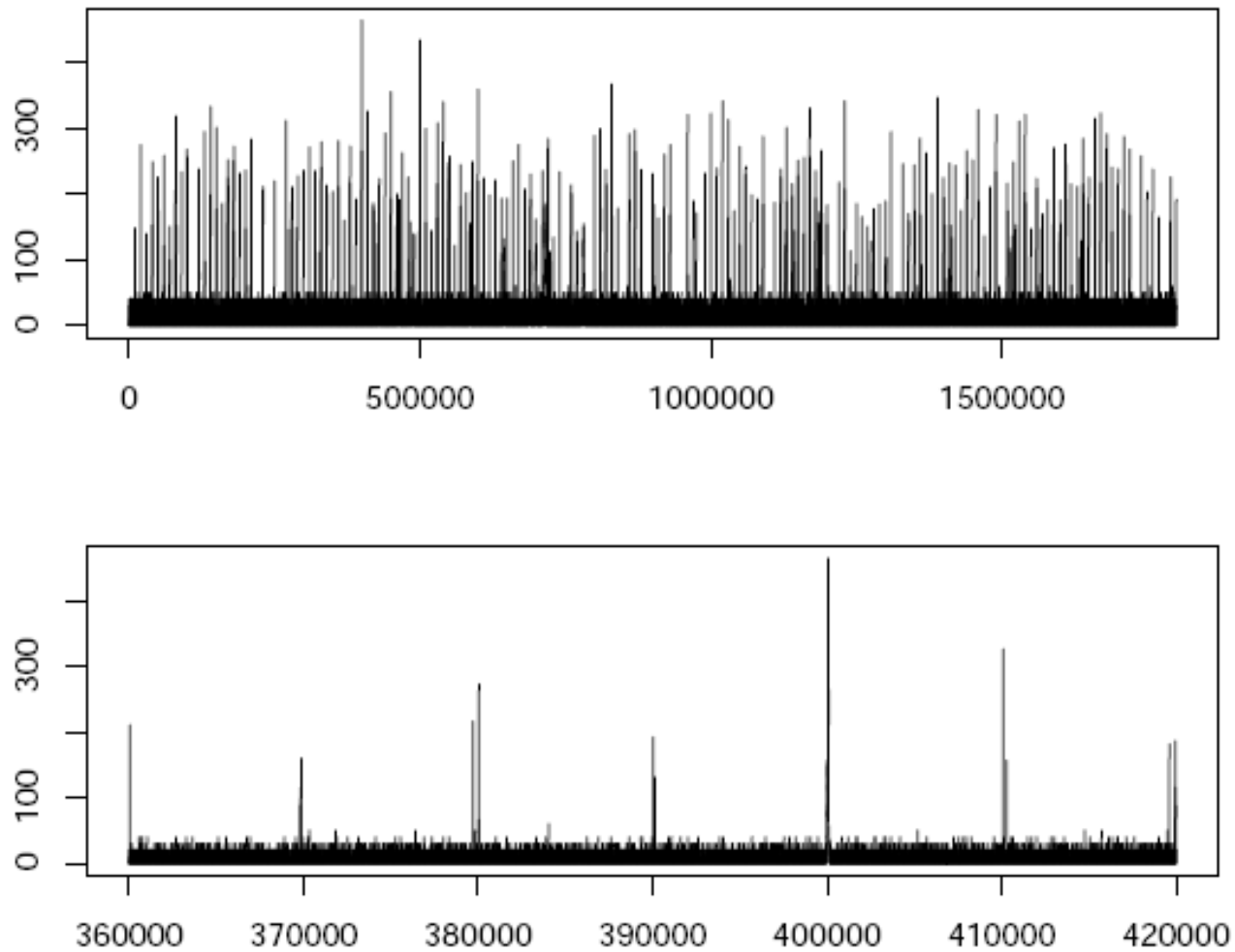
A possible solution: Hilbert curve visualisation

S. An.: “Visualisation of genomic data with the Hilbert curve”,
Bioinformatics, Vol. 25 (2009) pp. 1231-1235

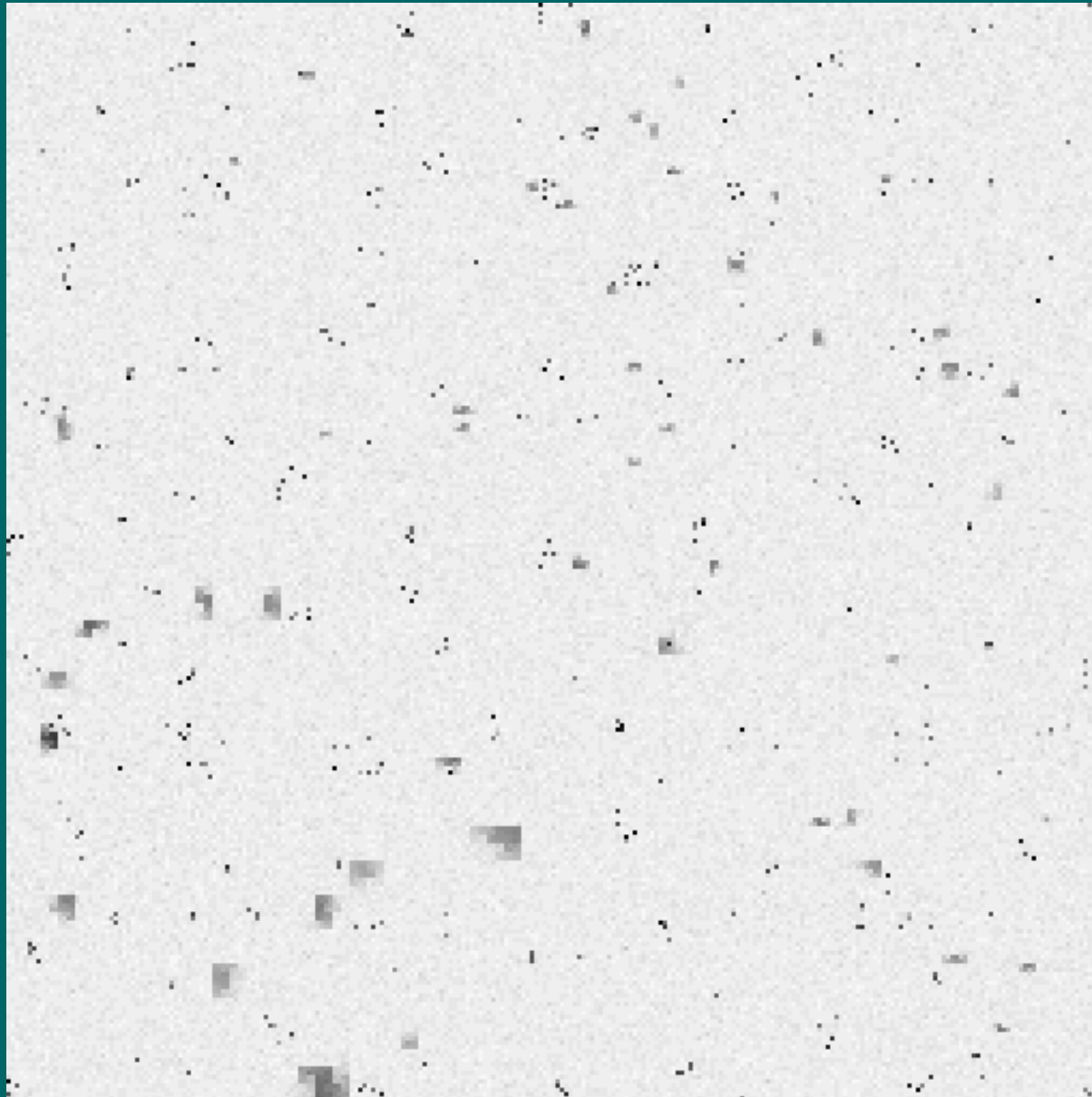
The Hilbert curve



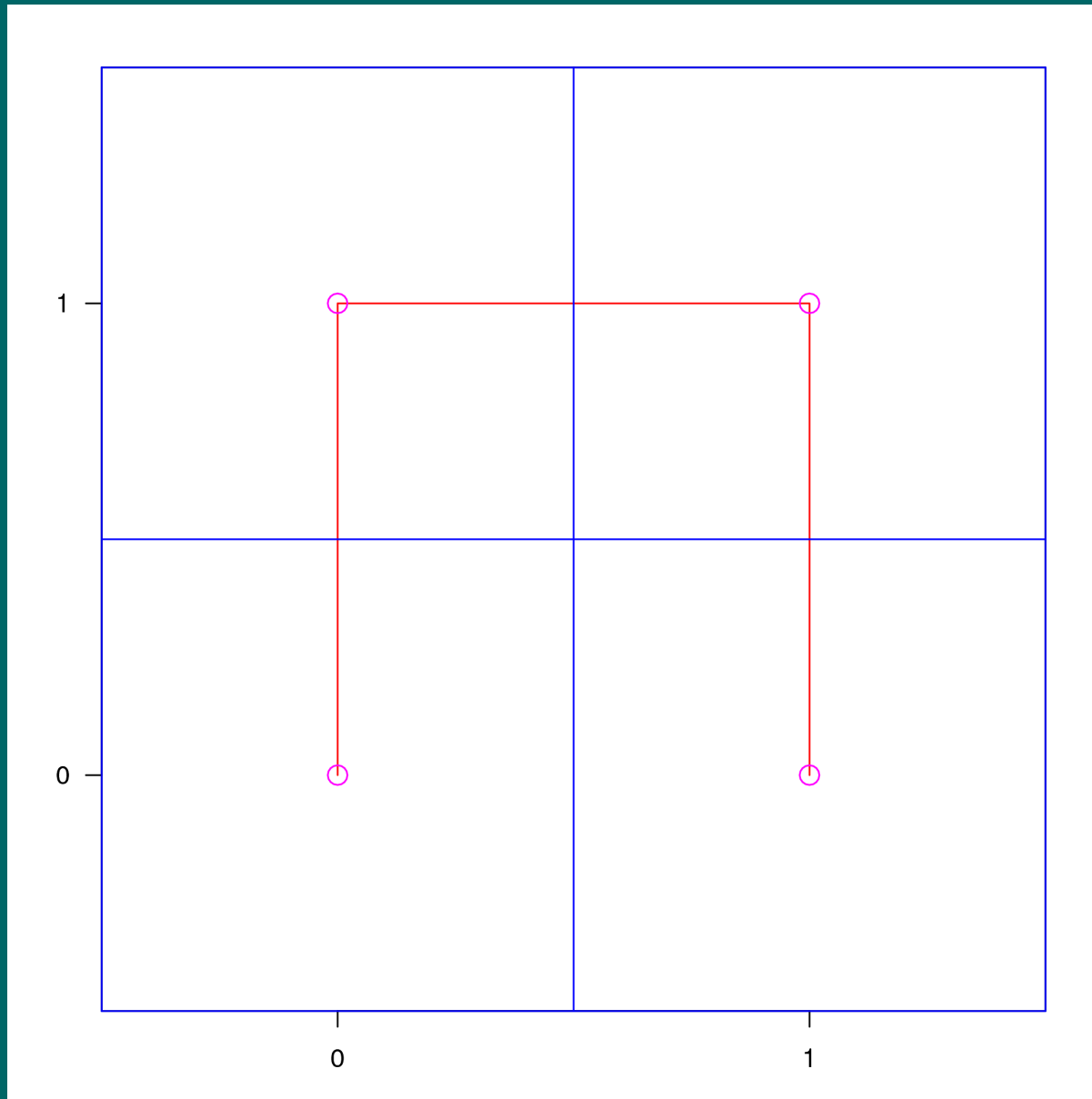
What is hidden in here?



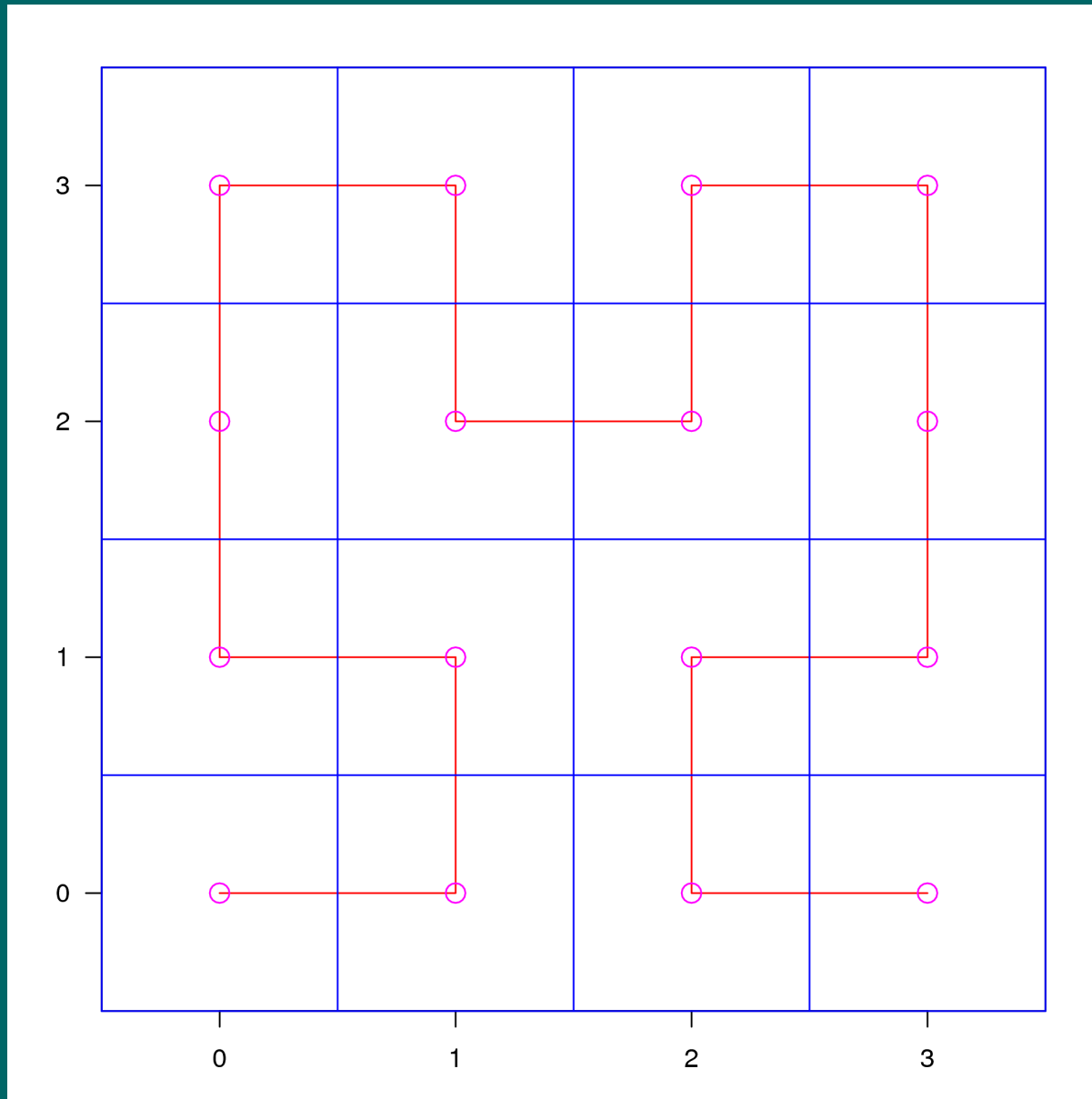
Hilbert plot of the constructed example vector



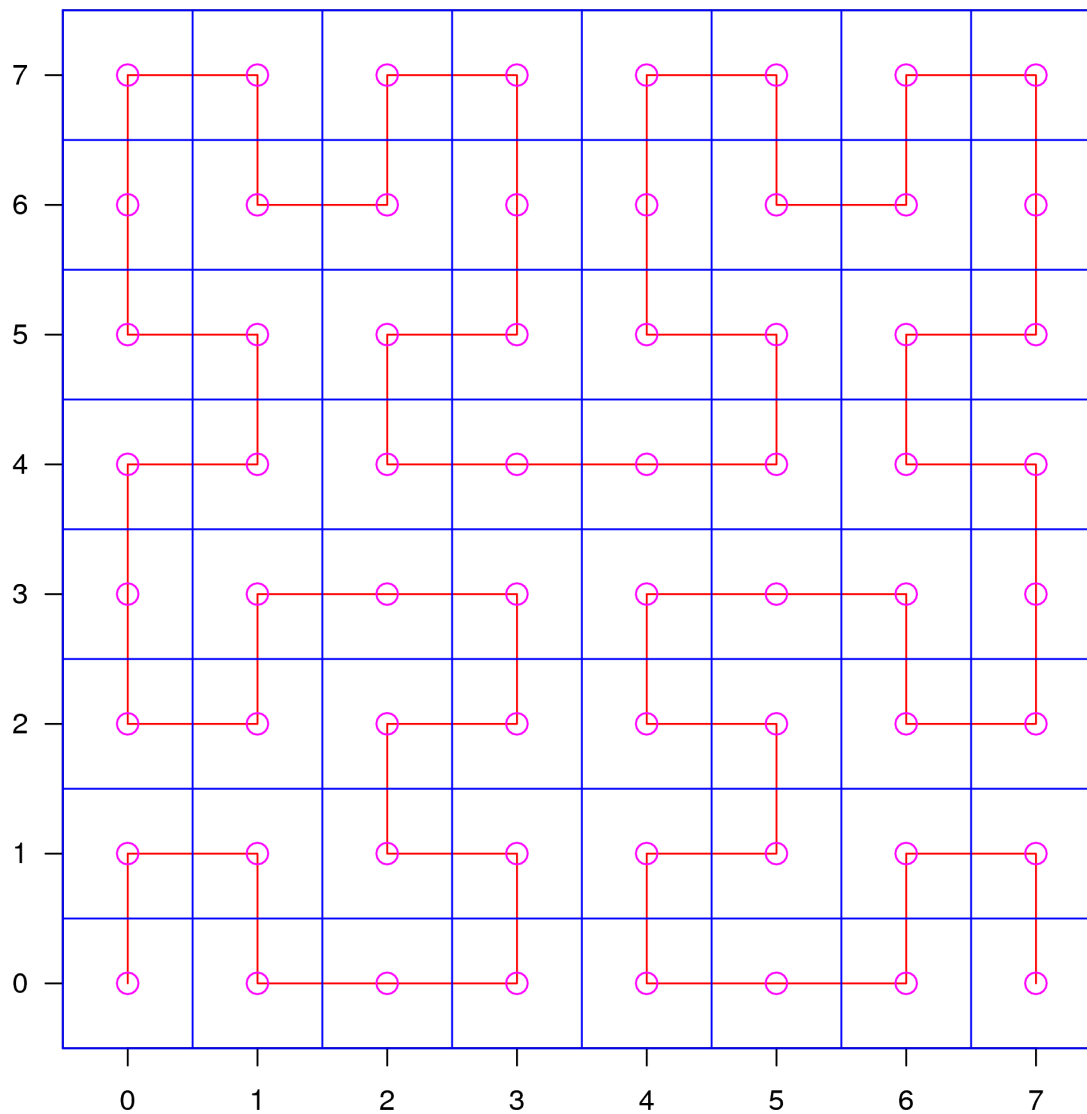
Construction of the Hilbert curve: Level 1



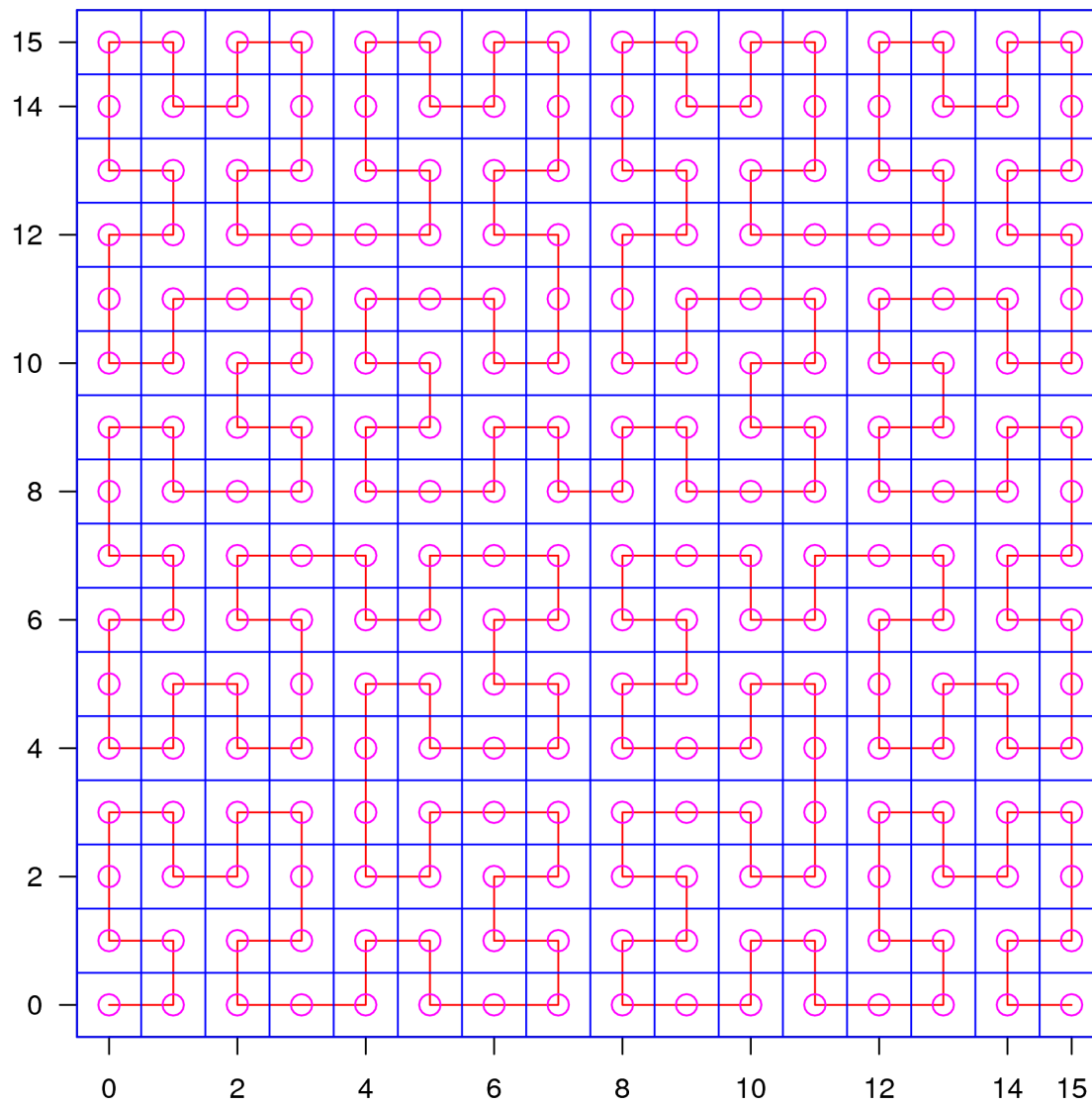
Construction of the Hilbert curve: Level 2



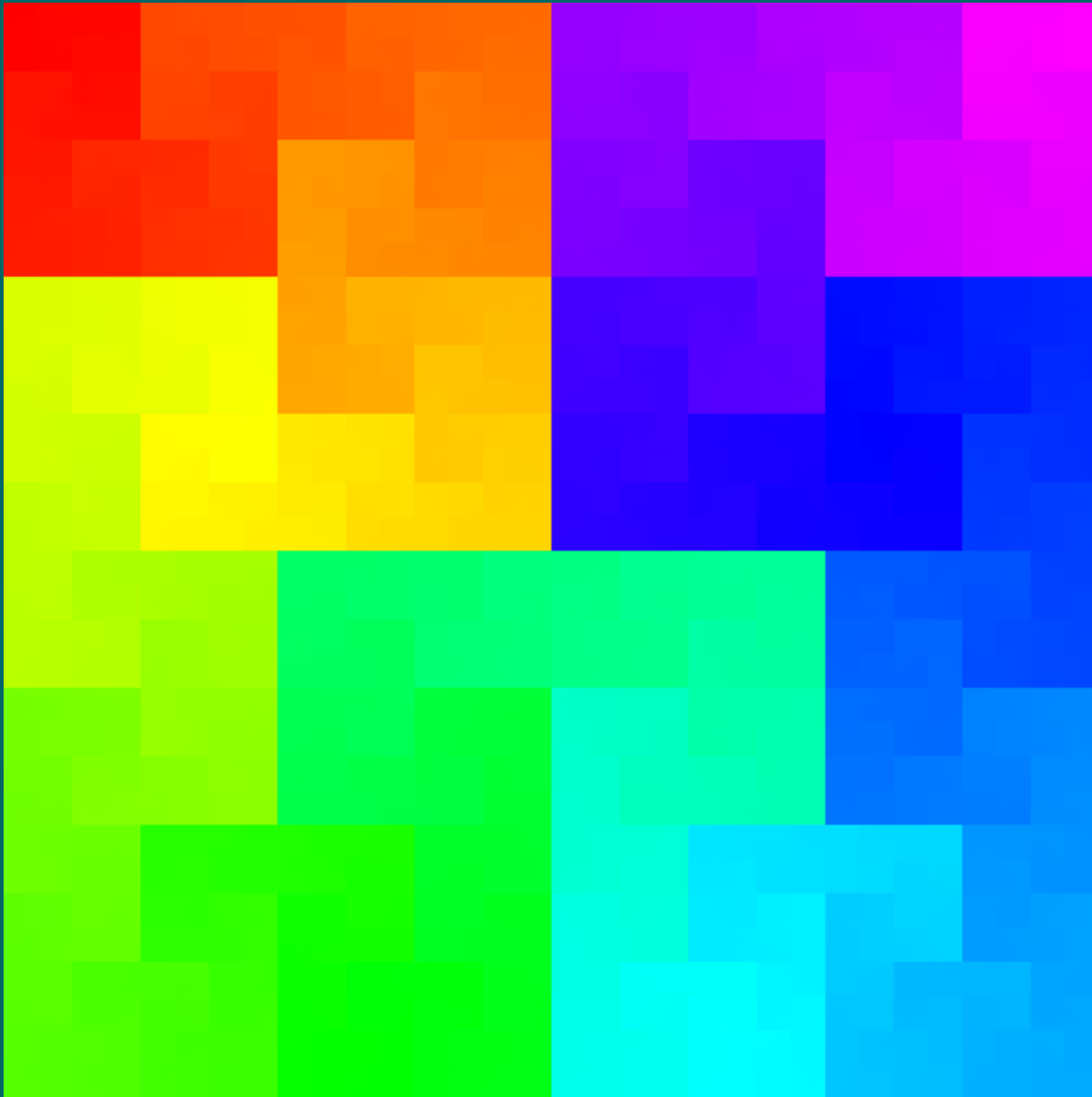
Construction of the Hilbert curve: Level 3



Construction of the Hilbert curve: Level 4

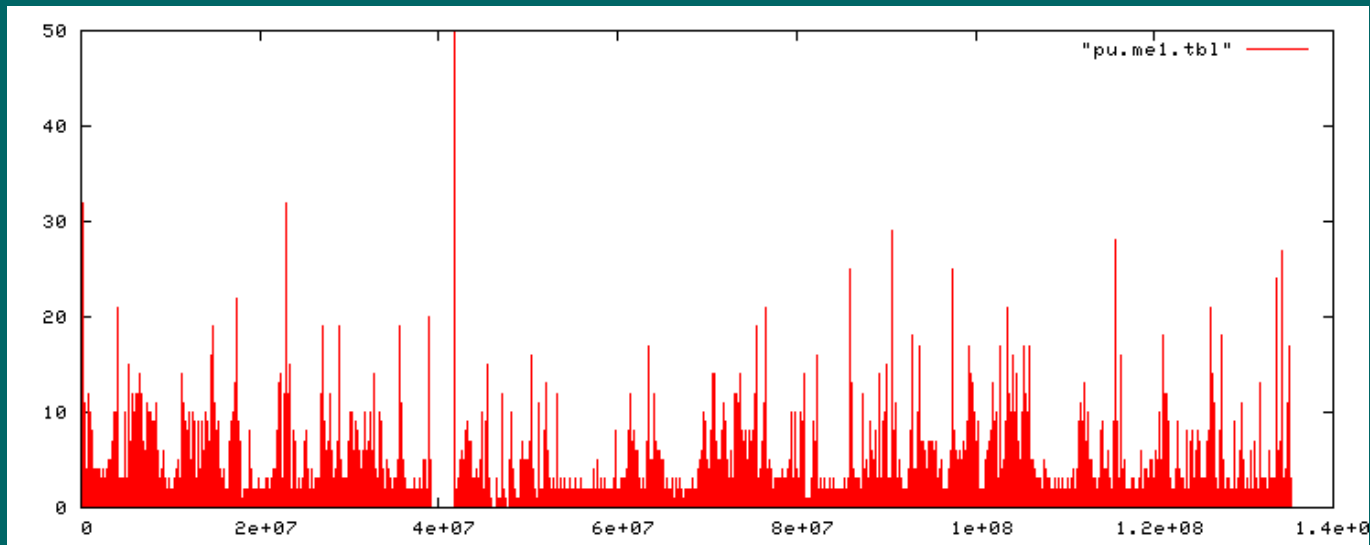


Hilbert curve: Approaching the limit

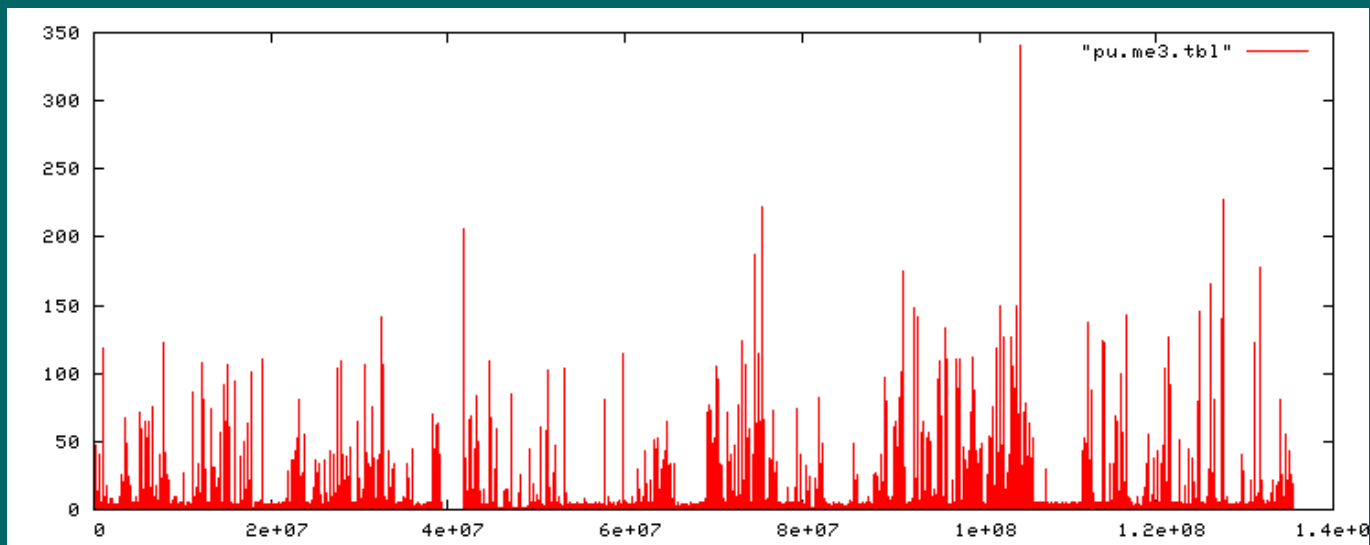


Coverage vector for a full chromosome (chr10)

H3K4me1

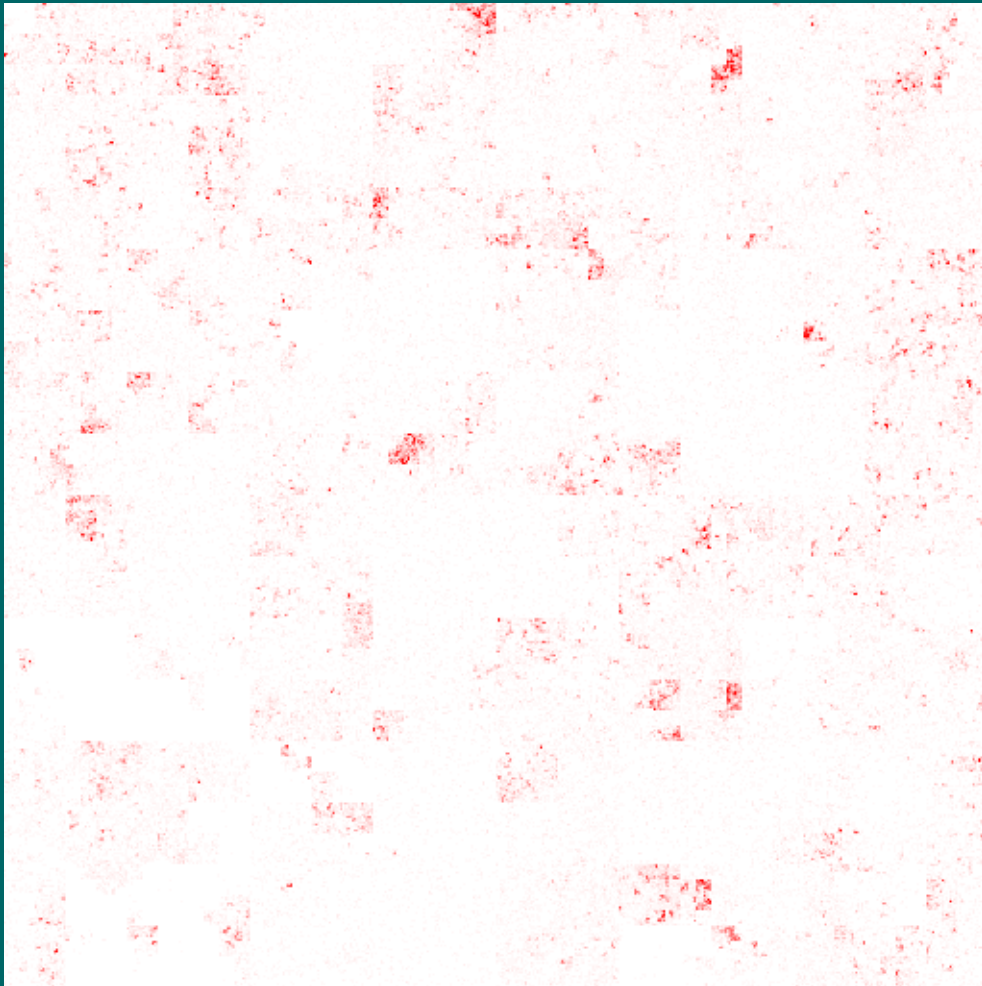


H3K4me3

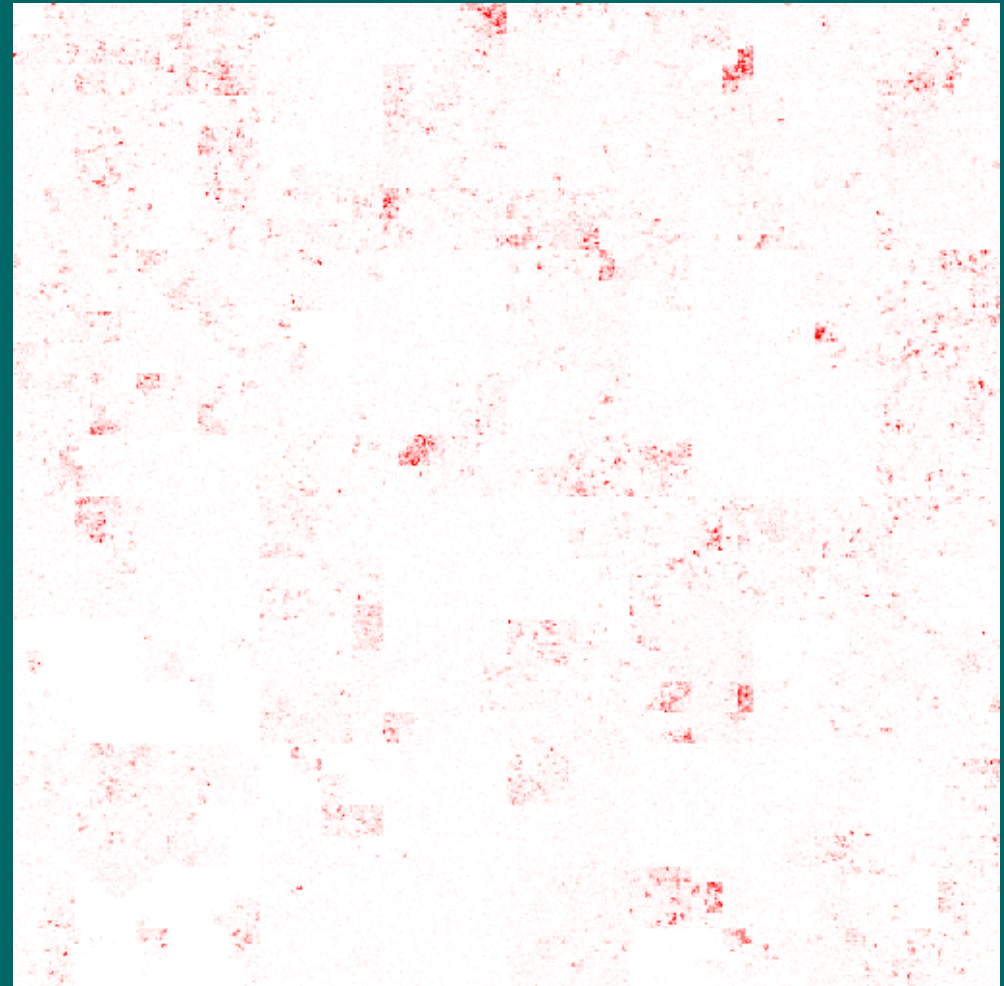


chrom.
10

Hilbert plot of the coverage vectors

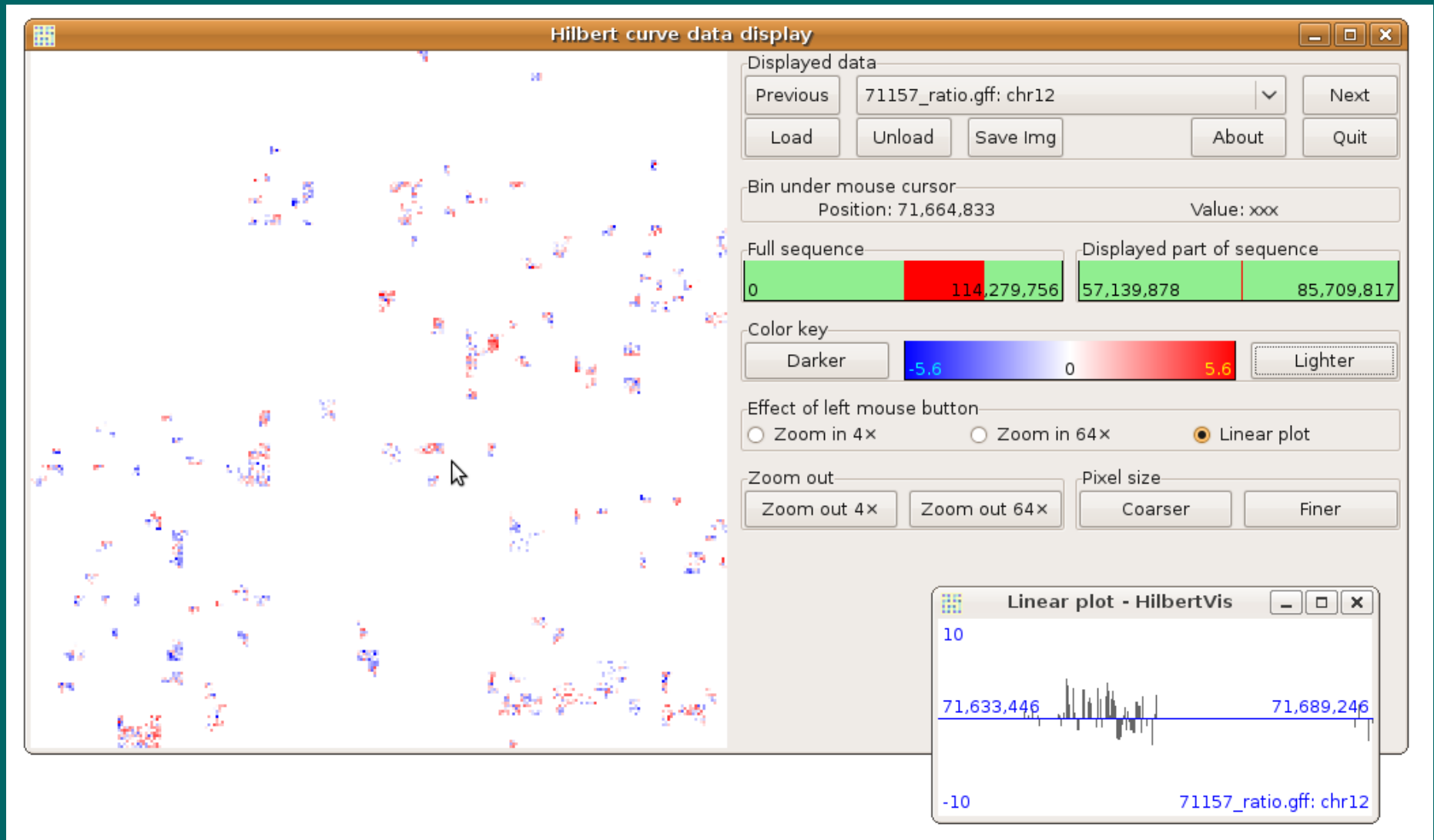


H3K4me1
(mono-methylation)



H3K4me3
(tri-methylation)

HilbertVis



HilbertVis

- stand-alone tool to display GFF, Wiggle, Maq map

<http://www.ebi.ac.uk/huber-srv/hilbert/>

(or Google for “hilbertvis”)

- R package to display any long R vector

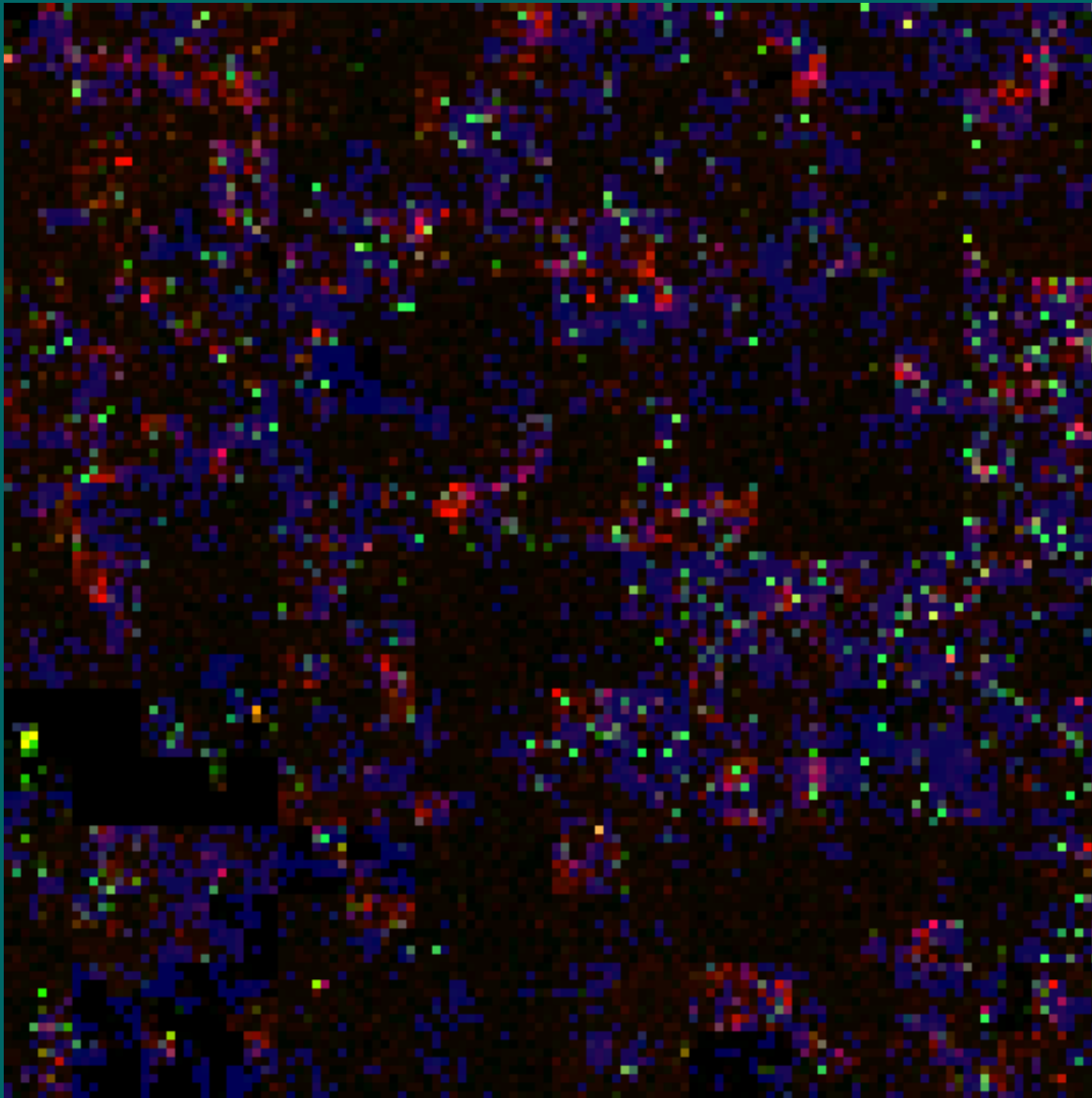
- either via commands for batch processing

Bioconductor package “HilbertVis”

- or via GUI for exploring

Bioconductor package “HilbertVisGUI”

Three-colour Hilbert plot



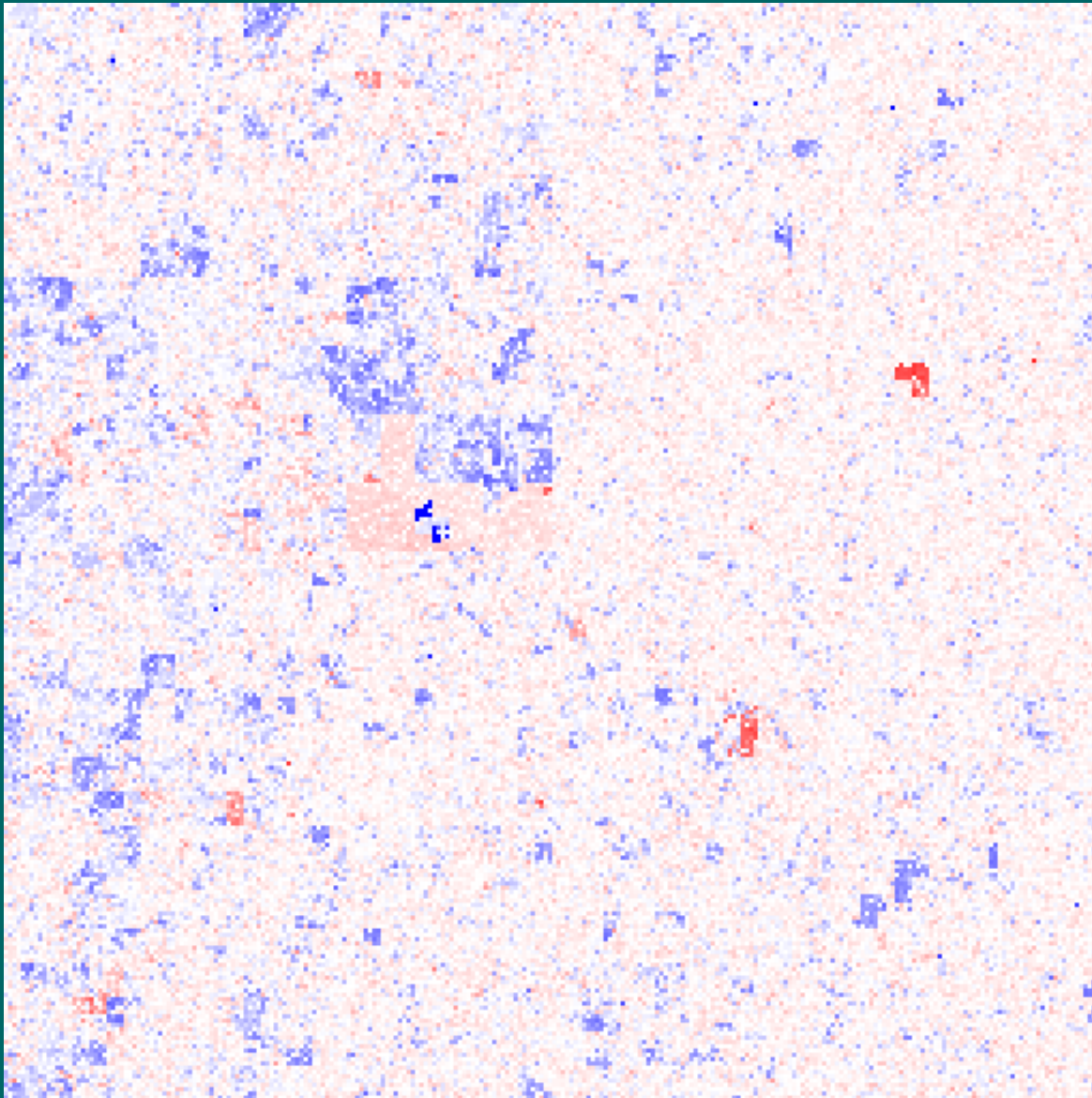
Overlay of the previous plots and exon density

red:
mono-methylation

green:
tri-methylation

blue:
exons

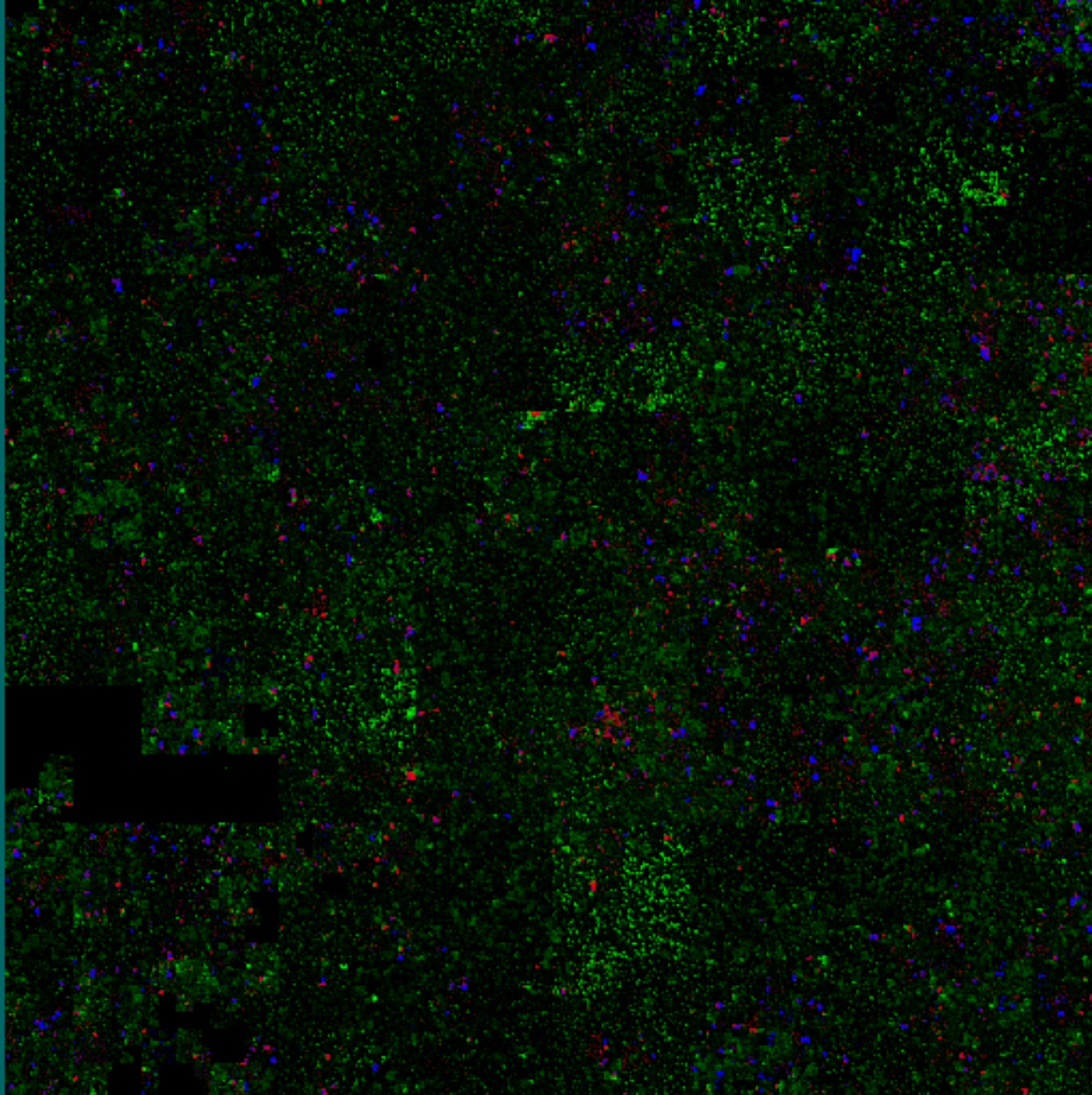
Other uses: Array-CGH



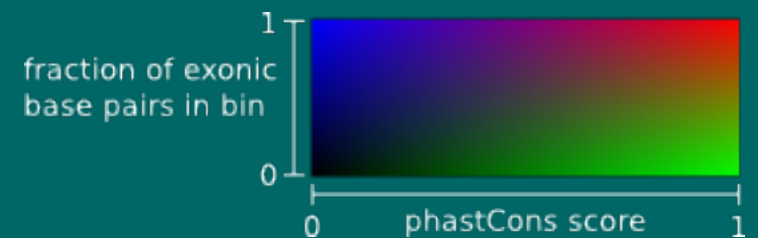
Log fold-changes
between two
Arabidopsis eco-types,
chromosome 2

[Data courtesy of
M. Seiffert, IPK Gatersleben

Other uses: Conservation scores



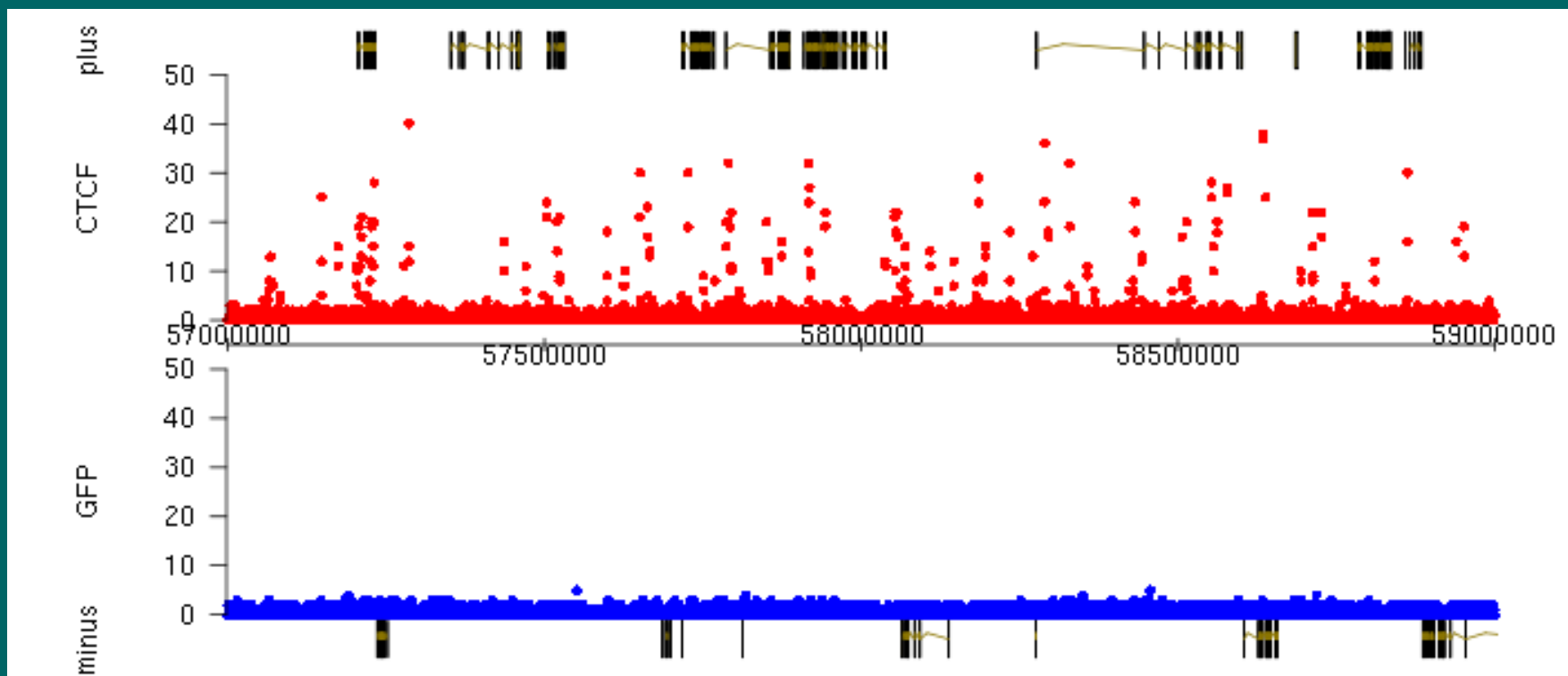
Human chromosome 10:
Comparing phastCons
conservation scores with
exon density



GenomeGraphs

GenomeGraphs: Bioconductor package by S. Durrinck, UCB

- Load gene models from Ensembl via BiomaRt and plots them, alongside experimental data



GenomeGraphs: Code for sample plot

```
library(GenomeGraphs)
library(HilbertVis)

mart <- useMart("ensembl", dataset = "mmusculus_gene_ensembl")

start <- 57000000
end <- 59000000

plusStrand <- makeGeneRegion( chromosome = 10,
                               start = start, end = end, strand = "+", biomart = mart )

minusStrand <- makeGeneRegion( chromosome = 10,
                               start = start, end = end, strand = "-", biomart = mart )

genomeAxis <- makeGenomeAxis( )
```



GenomeGraphs: Code for sample plot, cont'd

```
track.ctcf <- makeBaseTrack(  
  base = seq( start, end, length.out = 10000 ),  
  value = shrinkVector(  
    as.vector( cov.ctcf$chr10[start:end] ), 10000 ),  
  dp = DisplayPars( lwd = 0.5, color="red", ylim=c(0, 50) ) )  
  
track.gfp <- makeBaseTrack(  
  base = seq( start, end, length.out = 10000 ),  
  value = shrinkVector(  
    as.vector( cov.gfp$chr10[start:end] ), 10000 ),  
  dp = DisplayPars( lwd = 0.5, color="blue", ylim=c(0, 50) ) )  
  
gdPlot( list( `plus`=plusStrand, `CTCF`=track.ctcf,  
  genomeAxis, `GFP`=track.gfp, `minus`=minusStrand ) )
```

*