A brief overview of the S4 class system

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Outline

The S4 class system

- The *S4 class system* is a set of facilities provided in R for OO programming.
- Implemented in the *methods* package.
- On a fresh *R* session:

  ```
  > sessionInfo()

  ...
  attached base packages:
  [1] stats     graphics  grDevices utils     datasets
  [6] methods   base
  ```
- R also supports an older class system: the *S3 class system*.

A different world

The syntax

> `foo(x, ...)`

not:

> `x.foo(...)`

like in other OO programming languages.

The central concepts

- ► The core components: *classes*[1], *generic functions* and *methods*
- ► The glue: *method dispatch* (supports *simple* and *multiple* dispatch)

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[1]also called *formal classes*, to distinguish them from the S3 classes aka *old style classes*

The result

```
> ls('package:methods')
  [1] "@<-"                    "addNextMethod"
  [3] "allGenerics"            "allNames"
  [5] "Arith"                  "as"
  [7] "as<-"                   "asMethodDefinition"
...
[199] "testVirtual"            "traceOff"
[201] "traceOn"                "tryNew"
[203] "trySilent"              "unRematchDefinition"
[205] "validObject"           "validSlotNames"
```

- Rich, complex, can be intimidating
- The classes and methods we implement in our packages can be hard to document, especially when the class hierarchy is complicated and multiple dispatch is used

S4 in Bioconductor

- Heavily used. In BioC 2.7: 1383 classes and 8397 methods defined in 200 packages! (out of 419)
- Top 4: 94 classes in *flowCore* and *IRanges* (tie), 72 classes in *Biostrings*, 68 classes in *rsbml*, ...
- For the end-user: it's mostly transparent. But when something goes wrong, error messages issued by the S4 class system can be hard to understand. Also it can be hard to find the documentation for a specific method.
- Most Bioconductor packages use only a subset of the S4 capabilities (covers 99.99% of our needs)

Outline

Where do S4 objects come from?

From a dataset

```
> library(graph)
> data(apopGraph)
> apopGraph

A graphNEL graph with directed edges
Number of Nodes = 50
Number of Edges = 59
```

From using *the* constructor

```
> library(IRanges)
> IRanges(start=c(101, 25), end=c(110, 80))

IRanges of length 2
    start end width
[1]   101 110    10
[2]    25  80    56
```

From a coercion

```
> library(Matrix)
> m <- matrix(3:-4, nrow=2)
> as(m, "Matrix")

2 x 4 Matrix of class "dgeMatrix"
     [,1] [,2] [,3] [,4]
[1,]    3    1   -1   -3
[2,]    2    0   -2   -4
```

From using a specialized high-level constructor

```
> library(GenomicFeatures)
> makeTranscriptDbFromUCSC("sacCer2", tablename="ensGene")

TranscriptDb object:
| Db type: TranscriptDb
| Data source: UCSC
| Genome: sacCer2
| UCSC Table: ensGene
...
```

From using a high-level I/O function

```
> library(ShortRead)
> lane1 <- readFastq("path/to/my/data/", pattern="s_1_sequence.txt")
> lane1

class: ShortReadQ
length: 256 reads; width: 36 cycles
```

Inside an S4 object

```
> sread(lane1)

  A DNAStringSet instance of length 256
      width seq
  [1]    36 GGACTTTGTAGGATACCCTCGCTTTCCTTCTCCTGT
  [2]    36 GATTTCTTACCTATTAGTGGTTGAACAGCATCGGAC
  [3]    36 GCGGTGGTCTATAGTGTTATTAATATCAATTTGGGT
  [4]    36 GTTACCATGATGTTATTTCTTCATTTGGAGGTAAAA
  ...    ... ...
[253]    36 GTTTTACAGACACCTAAAGCTACATCGTCAACGTTA
[254]    36 GATGAACTAAGTCAACCTCAGCACTAACCTTGCGAG
[255]    36 GTTTGGTTCGCTTTGAGTCTTCTTCGGTTCCGACTA
[256]    36 GCAATCTGCCGACCACTCGCGATTCAATCATGACTT
```

How to manipulate S4 objects?

Low-level: getters and setters

```
> ir <- IRanges(start=c(101, 25), end=c(110, 80))
> width(ir)

[1] 10 56

> width(ir) <- width(ir) - 5
> ir

IRanges of length 2
    start end width
[1]   101 105     5
[2]    25  75    51
```

High-level: plenty of specialized methods

```
> qa1 <- qa(lane1, lane="lane1")
> class(qa1)

[1] "ShortReadQQA"
attr(,"package")
[1] "ShortRead"
```

How to find the right man page?

- `class?graphNEL` or equivalently `?`graphNEL-class`` for accessing the man page of a class
- `?qa` for accessing the man page of a generic function
- The man page for a generic might also document some or all of the methods for this generic. The *See Also:* section might give a clue. Also using `showMethods()` can be useful:

  ```
  > showMethods("qa")

  Function: qa (package ShortRead)
  dirPath="character"
  dirPath="list"
  dirPath="ShortReadQ"
  dirPath="SolexaPath"
  ```

- `?`qa,ShortReadQ-method`` to access the man page for a particular method (might be the same man page as for the generic)
- In doubt: `??qa` will search the man pages of all the installed packages and return the list of man pages that contain the string `qa`

Inspecting objects and discovering methods

- ▶ class() and showClass()

  ```
  > class(lane1)

  [1] "ShortReadQ"
  attr(,"package")
  [1] "ShortRead"

  > showClass("ShortReadQ")

  Class "ShortReadQ" [package "ShortRead"]

  Slots:

  Name:      quality      sread        id
  Class: QualityScore DNAStringSet  BStringSet

  Extends:
  Class "ShortRead", directly
  Class ".ShortReadBase", by class "ShortRead", distance 2

  Known Subclasses: "AlignedRead"
  ```

- ▶ str() for compact display of the content of an object
- ▶ showMethods() to discover methods
- ▶ selectMethod() to see the code

Outline

Class definition and constructor

Class definition

```
> setClass("SNPLocations",
+     representation(
+        genome="character",   # a single string
+        snpid="character",    # a character vector of length N
+        chrom="character",    # a character vector of length N
+        pos="integer"         # an integer vector of length N
+     )
+ )

[1] "SNPLocations"
```

Constructor

```
> SNPLocations <- function(genome, snpid, chrom, pos)
+     new("SNPLocations", genome=genome, snpid=snpid, chrom=chrom, pos=pos)

> snplocs <- SNPLocations("hg19",
+              c("rs0001", "rs0002"),
+              c("chr1", "chrX"),
+              c(224033L, 1266886L))
```

Getters

Defining the `length` method

```
> setMethod("length", "SNPLocations", function(x) length(x@snpid))
> length(snplocs)  # just testing
[1] 2
```

Defining the slot getters

```
> setGeneric("genome", function(x) standardGeneric("genome"))
> setMethod("genome", "SNPLocations", function(x) x@genome)

> setGeneric("snpid", function(x) standardGeneric("snpid"))
> setMethod("snpid", "SNPLocations", function(x) x@snpid)

> setGeneric("chrom", function(x) standardGeneric("chrom"))
> setMethod("chrom", "SNPLocations", function(x) x@chrom)

> setGeneric("pos", function(x) standardGeneric("pos"))
> setMethod("pos", "SNPLocations", function(x) x@pos)

> genome(snplocs)  # just testing
[1] "hg19"
> snpid(snplocs)   # just testing
[1] "rs0001" "rs0002"
```

Defining the `show` method

```
> setMethod("show", "SNPLocations",
+     function(object)
+         cat(class(object), "instance with", length(object),
+             "SNPs on genome", genome(object), "\n")
+ )

> snplocs  # just testing

SNPLocations instance with 2 SNPs on genome hg19
```

Defining the *validity method*

```
> setValidity("SNPLocations",
+     function(object) {
+         if (!is.character(genome(object)) ||
+             length(genome(object)) != 1 || is.na(genome(object)))
+             return("'genome' slot must be a single string")
+         slot_lengths <- c(length(snpid(object)),
+                           length(chrom(object)),
+                           length(pos(object)))
+         if (length(unique(slot_lengths)) != 1)
+             return("lengths of slots 'snpid', 'chrom' and 'pos' differ")
+         TRUE
+     }
+ )

> snplocs@chrom <- LETTERS[1:3]  # a very bad idea!
> validObject(snplocs)

Error in validObject(snplocs) :
  invalid class "SNPLocations" object: lengths of slots 'snpid', 'chrom'
  and 'pos' differ
```

Defining slot setters

```
> setGeneric("chrom<-", function(x, value) standardGeneric("chrom<-"))
> setReplaceMethod("chrom", "SNPLocations",
+     function(x, value) {x@chrom <- value; validObject(x); x})

> chrom(snplocs) <- LETTERS[1:2]  # repair currently broken object

> chrom(snplocs) <- LETTERS[1:3]  # try to break it again

Error in validObject(x) :
  invalid class "SNPLocations" object: lengths of slots 'snpid', 'chrom'
  and 'pos' differ
```

Defining a coercion method

```
> setAs("SNPLocations", "data.frame",
+     function(from)
+         data.frame(snpid=snpid(from), chrom=chrom(from), pos=pos(from))
+ )

> as(snplocs, "data.frame")  # testing

  snpid chrom     pos
1 rs0001     A  224033
2 rs0002     B 1266886
```

Outline

Slot inheritance

- Most of the time (but not always), the child class will have additional slots:

```
> setClass("AnnotatedSNPs",
+     contains="SNPLocations",
+     representation(
+         geneid="character"  # a character vector of length N
+     )
+ )
[1] "AnnotatedSNPs"
```

- The slots from the parent class are inherited:

```
> showClass("AnnotatedSNPs")

Class "AnnotatedSNPs" [in ".GlobalEnv"]

Slots:

Name:    geneid    genome    snpid    chrom    pos
Class: character character character character integer

Extends: "SNPLocations"
```

- Constructor:

```
> AnnotatedSNPs <- function(genome, snpid, chrom, pos, geneid)
+ {
+     new("AnnotatedSNPs",
+         SNPLocations(genome, snpid, chrom, pos),
+         geneid=geneid)
+ }
```

Method inheritance

- Let's create an AnnotatedSNPs object:

```
> snps <- AnnotatedSNPs("hg19",
+                c("rs0001", "rs0002"),
+                c("chr1", "chrX"),
+                c(224033L, 1266886L),
+                c("AAU1", "SXW-23"))
```

- All the methods defined for SNPLocations objects work out-of-the-box:

```
> snps
```

```
AnnotatedSNPs instance with 2 SNPs on genome hg19
```

- But sometimes they don't do the right thing:

```
> as(snps, "data.frame")  # the 'geneid' slot is ignored
```

```
  snpid chrom     pos
1 rs0001  chr1  224033
2 rs0002  chrX 1266886
```

- ▶ Being a SNPLocations *object* vs being a SNPLocations *instance*:

```
> is(snps, "AnnotatedSNPs")      # 'snps' is an AnnotatedSNPs object
[1] TRUE
> is(snps, "SNPLocations")       # and is also a SNPLocations object
[1] TRUE
> class(snps)                    # but is *not* a SNPLocations *instance*
[1] "AnnotatedSNPs"
attr(,"package")
[1] ".GlobalEnv"
```

- ▶ Method overriding: for example we could define a `show` method for AnnotatedSNPs objects. `callNextMethod` can be used in that context to call the method defined for the parent class from within the method for the child class.

- ▶ Automatic coercion method:

```
> as(snps, "SNPLocations")
SNPLocations instance with 2 SNPs on genome hg19
```

Incremental validity method

- The *validity method* for AnnotatedSNPs objects only needs to validate what's not already validated by the *validity method* for SNPLocations objects:

```
> setValidity("AnnotatedSNPs",
+     function(object) {
+         if (length(object@geneid) != length(object))
+             return("'geneid' slot must have the length of the object")
+         TRUE
+     }
+ )
```

- In other words: before an AnnotatedSNPs object can be considered valid, it must first be a valid SNPLocations object.

Outline

Other important S4 features

- ▶ *Virtual* classes: equivalent to *abstract* classes in Java
- ▶ Class unions (see `?setClassUnion`)
- ▶ Multiple inheritance: a powerful feature that should be used with caution. If used inappropriately, can lead to a class hierarchy that is hard or impossible to maintain

Resources

- ▶ Man pages in the *methods* package: `?setClass`, `?showMethods`, `?selectMethod`, `?getMethod`, `?is`, `?setValidity`, `?as`
- ▶ Note: S4 is *not* covered in the *An Introduction to R* or *The R language definition* manuals[2]
- ▶ The *Writing R Extensions* manual for details about integrating S4 classes to a package
- ▶ The *R Programming for Bioinformatics* book by Robert Gentleman[3]

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[2] http://cran.fhcrc.org/manuals.html

[3] http://bioconductor.org/help/publications/books/r-programming-for-bioinformatics/