

Package ‘CellMixS’

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Type Package

Title Evaluate Cellspecific Mixing

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Description

Evaluate Cellspecific Mixing Scores (CMS) for different batches/groups in scRNA-seq data.

License GPL (>=2)

Imports BiocNeighbors, ggplot2, scater, viridis, cowplot,
SummarizedExperiment, SingleCellExperiment, tidyr, magrittr,
dplyr, ggridges, stats, purrr, listarrays, methods,
BiocParallel

Depends kSamples, R (>= 3.6)

biocViews SingleCell, Transcriptomics, GeneExpression, BatchEffect

BugReports <https://github.com/almutlue/CellMixS/issues>

URL <https://github.com/almutlue/CellMixS>

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R topics documented:

CellMixS-package	2
.cmsCell	2
.defineSubspace	3
.filterLocMin	4

.ldfKnn	5
.smoothCms	6
cms	7
ldfDiff	8
ldfSce	9
visCluster	11
visGroup	12
visHist	13
visIntegration	14
visMetric	15
visOverview	16

Index	17
--------------	-----------

CellMixS-package	<i>Toolbox to explore batch effects and data integration in scRNA data.</i>
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Description

CellMixS provides metrics and functions to evaluate batch effects, data integration and batch effect correction in single cell transcriptome data with single cell resolution. Results can be visualized and summarised on different levels, e.g. on cell, celltype or dataset level.

Details

In particular, **CellMixS** includes two main metrics: Cellspecific mixing scores to determine the probability of random mixing in each cell's neighbourhood. It can be assessed via the `cms` function. Local Density Factor Differences to evaluate the effect of data integration methods on batch internal structures. It can be assessed via the `ldfDiff` function.

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.cmsCell	<i>.cmsCell</i>
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Description

Function to calculate a cellspecific mixing score (cms) of groups/batches.

Usage

```
.cmsCell(cell, group, knn, k_min = NA, cell_min = 4)
```

Arguments

cell	Character. Name of the cell to calculate cms for. Needs to be one of rownames(knn).
group	Character. Name of group/batch variable. Needs to be one of names(knn).
knn	List with three elements. First "index" with indices of KNN cells. Second "distance" with distances to KNN cells. Third a slot named by group variable with group level of KNN cells.
k_min	Numeric. Minimum number of Knn to include. Default is NA (see Details).
cell_min	Numeric. Minimum number of cells from each group to be included into the AD test. Should be > 4 to make 'ad.test' working.

Details

The cms function tests the hypothesis, that group-specific distance distributions of knn cells have the same underlying unspecified distribution. It performs Anderson-Darling tests as implemented in the kSamples package. In default the function uses all distances and group label defined in knn. If k_min is specified, the first local minimum of the overall distance distribution with at least kmin cells is used. This can be used to adapt to the local structure of the dataset e.g. prevent cells from a distinct different cluster to be included.

Value

A p.value as resulting from the ad.test.

See Also

[ad.test](#), [cms](#), [.smoothCms](#)

Other helper functions: [.defineSubspace](#), [.filterLocMin](#), [.ldfKnn](#), [.smoothCms](#)

`.defineSubspace` *.defineSubspace*

Description

Helper function for ldfSce and cms to define or recalculate the subspace for analysis.

Usage

```
.defineSubspace(sce, assay_name, dim_red, n_dim)
```

Arguments

sce	A SingleCellExperiment object with the data to define the subspace.
assay_name	Character. Name of the assay to use for PCA. Only relevant if no existing 'dim_red' is provided. Must be one of names(assays(sce)).
dim_red	Character. Name of embeddings to use as subspace.
n_dim	Numeric. Number of subspace elements to include to define subspace.

Details

Function to determine the subspace for `ldfDiff` and `cms`. Checks whether the defined 'dim_red' is present. Only if no subspace is defined or present it will perform a PCA using `runPCA`. To calculate PCA counts defined in 'assay_name' are used.

Value

A matrix of cell embeddings with reduced dimensions as columns.

See Also

[ldfSce](#), [cms](#).

Other helper functions: [.cmsCell](#), [.filterLocMin](#), [.ldfKnn](#), [.smoothCms](#)

<code>.filterLocMin</code>	<i>.filterLocMin</i>
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Description

Function to filter knn by overall distance density distribution.

Usage

```
.filterLocMin(knn_cell, k_min)
```

Arguments

<code>knn_cell</code>	Data frame with one column "distance" and one column named by the group variable. Rows correspond to the knn cells and do not need rownames.
<code>k_min</code>	Numeric. Minimum number of Knn to include.

Details

Internal function to filter cells used for `cms` testing to come from a continuous overall density distribution function (similar to cluster definitions). 'filterLocMin' is only applied, if k-min is specified as parameter in [.cmsCell](#) or [cms](#).

Value

data.frame with two columns (index, distance) for filtered knn cells.

See Also

[.cmsCell](#)

Other helper functions: [.cmsCell](#), [.defineSubspace](#), [.ldfKnn](#), [.smoothCms](#)

.ldfKnn *.ldfKnn*

Description

Calculates the Local Density Factor as implemented in the `DDoutlier` package with a predefined KNN neighbourhood.

Usage

```
.ldfKnn(dataset, knn_object, k = k, h = 1, c = 1)
```

Arguments

<code>dataset</code>	Matrix with cell embeddings with cells as rows and reduced dimensions as cloumns. Subspace to determine LDF in.
<code>knn_object</code>	List with k-nearest neighbours (KNN) as provided by <code>get.knn</code> from the <code>FNN</code> package. First element named "indices" contains indices of KNN in <code>dataset</code> . Second element named "distance" contains distances of KNN in <code>dataset</code> . Third element named "cell_name" contains rownames of KNN in <code>dataset</code> .
<code>k</code>	Numeric. Number of KNN used. Should correspond to <code>knn_object</code> .
<code>h</code>	Numeric. Bandwidth for kernel functions. The greater the bandwidth, the smoother kernels and lesser weight are put on outliers. Default is 1
<code>c</code>	Scaling constant for comparison of LDE to neighboring observations. Default is 1.

Details

LDF fuction modified from the `DDoutlier` package. Calculates a Local Density Estimate (LDE) and Local Density Factor (LDF) with a gaussian kernel. Modified to use a predefined KNN neighbourhood. For [ldfSce](#) this is essential to determine LDF after data integration on the same set of cells.

Value

List with two elements "LDE" and "LDF".

See Also

[ldfSce](#)

Other helper functions: `.cmsCell`, `.defineSubspace`, `.filterLocMin`, `.smoothCms`

`.smoothCms`*.smoothCms*

Description

Performs weighted smoothening of cms scores

Usage

```
.smoothCms(knn, cms_raw, cell_names, k_min, k)
```

Arguments

<code>knn</code>	List with three elements. First "index" with indices of KNN cells. Second "distance" with distances to KNN cells. Third a slot named by group variable with group level of KNN cells.
<code>cms_raw</code>	Matrix with raw cms scores for all cells specified in <code>cell_names</code> and <code>knn</code> . Column names need to be "cms".
<code>cell_names</code>	Character vector with cell names corresponding to the rownames of the list elements in <code>knn</code> and <code>rownames(cms_raw)</code> .
<code>k_min</code>	Numeric. Minimum number of Knn to include. Default is NA (see Details).
<code>k</code>	Numeric. Number of k-nearest neighbours (Knn) to use.

Details

Internal function to smooth cms scores. In a complete random setting cms scores are uniform distributed. To reduce the resulting random variance and enable visualization of local pattern cms scores can be smoothened assuming that within one region mixing is uniform. Generates smoothened cms scores using weighed means of cms scores within the k-nearest neighbourhood. Reciprocal distances are used as weights.

Value

matrix with two columns ("cms_smooth", "cms").

See Also

[.cmsCell](#), [cms](#)

Other helper functions: [.cmsCell](#), [.defineSubspace](#), [.filterLocMin](#), [.ldfKnn](#)

cms

cms

Description

Calculates cell-specific mixing scores based on euclidean distances within a subspace of integrated data.

Usage

```
cms(sce, k, group, dim_red = "PCA", assay_name = "logcounts",
    res_name = NULL, k_min = NA, smooth = TRUE, n_dim = 20,
    cell_min = 10, BPPARAM = SerialParam())
```

Arguments

sce	A SingleCellExperiment object with the combined data.
k	Numeric. Number of k-nearest neighbours (Knn) to use.
group	Character. Name of group/batch variable. Needs to be one of names(colData(sce))
dim_red	Character. Name of embeddings to use as subspace for distance distributions. Default is "PCA".
assay_name	Character. Name of the assay to use for PCA. Only relevant if no existing 'dim_red' is provided. Must be one of names(assays(sce)). Default is "logcounts".
res_name	Character. Appendix of the result score's name (e.g. method used to combine batches).
k_min	Numeric. Minimum number of Knn to include. Default is NA (see Details).
smooth	Logical. Indicating if cms results should be smoothed within each neighbourhood using the weighted mean.
n_dim	Numeric. Number of dimensions to include to define the subspace.
cell_min	Numeric. Minimum number of cells from each group to be included into the AD test. Should be > 4 to make the ad.test function working.
BPPARAM	A BiocParallelParam object specifying whether cms scores shall be calculated in parallel.

Details

The cms function tests the hypothesis, that group-specific distance distributions of knn cells have the same underlying unspecified distribution. It performs Anderson-Darling tests as implemented in the kSamples package. In default the function uses all distances and group label defined in knn. If k_min is specified, the first local minimum of the overall distance distribution with at least k_min cells is used. This can be used to adapt to the local structure of the dataset e.g. prevent cells from a different cluster to be included. If 'dim_red' is not defined or default cms will calculate a PCA using runPCA. Results will be appended to colData(sce). Names can be specified using res_name. If multiple cores are available cms scores can be calculated in parallel (does not work on Windows). Parallelization can be specified using BPPARAM.

Value

A `SingleCellExperiment` with `cms` (and `cms_smooth`) within `colData`.

References

Scholz, F. W. and Stephens, M. A. (1987). K-Sample Anderson-Darling Tests. *J. Am. Stat. Assoc.*

See Also

[.cmsCell](#), [.smoothCms](#).

Examples

```
library(SingleCellExperiment)
sim_list <- readRDS(system.file("extdata/sim50.rds", package = "CellMixS"))
sce <- sim_list[[1]][, c(1:50)]

sce_cms <- cms(sce, k = 20, group = "batch", n_dim = 2)
```

 ldfDiff

ldfDiff

Description

Determines cell-specific changes in the Local Density Factor before and after data integration.

Usage

```
ldfDiff(sce_pre_list, sce_combined, group, k = 75, dim_red = "PCA",
        dim_combined = dim_red, assay_pre = "logcounts",
        assay_combined = "logcounts", n_dim = 20, res_name = NULL)
```

Arguments

<code>sce_pre_list</code>	A list of <code>SingleCellExperiment</code> objects with single datasets before integration. Names should correspond to levels in <code>colData(sce_combined)\$group</code>
<code>sce_combined</code>	A <code>SingleCellExperiment</code> object with the combined data.
<code>group</code>	Character. Name of group/batch variable that separates elements of <code>sce_pre_list</code> . Needs to be one of <code>names(colData(sce_combined))</code> .
<code>k</code>	Numeric. Number of k-nearest neighbours (Knn) to use.
<code>dim_red</code>	Character. Name of embeddings to use as subspace to calculate LDF before integration. Default is "PCA".
<code>dim_combined</code>	Character. Name of embeddings to use as subspace to calculate LDF after integration. Default is <code>dim_red</code> .
<code>assay_pre</code>	Character. Name of the assay to use for PCA. Only relevant if no existing 'dim_red' is provided. Must be one of <code>names(assays(sce_pre))</code> . Default is "logcounts".

assay_combined	Character. Name of the assay to use for PCA. Only relevant if no existing 'dim_red' is provided. Must be one of names(assays(sce_combined)). Default is "logcounts".
n_dim	Numeric. Number of PCs to include to define subspaces.
res_name	Character. Appendix of the result score's name (e.g. method used to combine batches). Used to specify result name for more than one run on the same input.

Details

The `ldfDiff` function calculates differences in LDF for each element in `sce_pre_list` and their corresponding cells in `sce_combined` using [ldfSce](#). If 'dim_red' is not defined a PCA will be calculated using `runPCA`. In this case 'assay_pre' need to refer to the data slot that shall define the subspace. Similar refer 'dim-combined' and 'assay_combined' to the integrated subspace or to the resp. "corrected" count data slot. 'k' can be used to define the level of local structure that is tested. The smaller 'k' the more focus is on detailed structures, while a large k will tests overall changes.

Value

A `SingleCellExperiment` object.

References

Latecki, Longin Jan and Lazarevic, Aleksandar and Pokrajac, Dragoljub (2007). Outlier Detection with Kernel Density Functions. Mach. Learn. Data Min. Pattern Recognit.. Springer Berlin Heidelberg.

See Also

[ldfSce](#), [.ldfKnn](#).

Other ldf functions: [ldfSce](#)

Examples

```
library(SingleCellExperiment)
sim_list <- readRDS(system.file("extdata/sim50.rds", package = "CellMixS"))
sce <- sim_list[["batch20"]][, c(1:50, 300:350)]
sce_batch1 <- sce[,colData(sce)$batch == "1"]
sce_batch2 <- sce[,colData(sce)$batch == "2"]
sce_pre_list <- list("1" = sce_batch1, "2" = sce_batch2)

sce_ldf <- ldfDiff(sce_pre_list, sce, k = 10, group = "batch",
dim_combined = "MNN", n_dim = 2)
```

ldfSce

ldfSce

Description

Determines cell-specific changes in the Local Density Factor before and after data integration for one specific group.

Usage

```
ldfSce(sce_name, sce_pre_list, sce_combined, group, k = 75,
       dim_red = "PCA", dim_combined = dim_red, assay_pre = "logcounts",
       assay_combined = "logcounts", n_dim = 20)
```

Arguments

sce_name	Character. Name of the element in sce_pre_list to calculate LDF differences in.
sce_pre_list	A list of SingleCellExperiment objects with single datasets before integration. Names need to correspond to levels in colData(sce_combined)\$group and sce_name!!
sce_combined	A SingleCellExperiment object with combined data.
group	Character. Name of group/batch variable that separates elements of sce_pre_list. Needs to be one of names(colData(sce_combined)).
k	Numeric. Number of k-nearest neighbours (Knn) to use.
dim_red	Character. Name of embeddings to use as subspace to calculate LDF before integration. Default is "PCA".
dim_combined	Character. Name of embeddings to use as subspace to calculate LDF after integration. Default is dim_red.
assay_pre	Character. Name of the assay to use for PCA. Only relevant if no existing 'dim_red' is provided. Must be one of names(assays(sce_pre)). Default is "logcounts".
assay_combined	Character. Name of the assay to use for PCA. Only relevant if no existing 'dim_red' is provided. Must be one of names(assays(sce_combined)). Default is "logcounts".
n_dim	Numeric. Number of PCs to include to define subspaces.

Details

The ldfSce function calculates differences in LDF for one specified element in sce_pre_list and their corresponding cells in sce_combined. If 'dim_red' is not defined a PCA will be calculated using runPCA. In this case 'assay_pre' need to refer to the data slot that shall define the subspace. Similar refer 'dim-combined' and 'assay_combined' to the integrated subspace or to the resp. "corrected" count data slot. 'k' can be used to define the level of local structure that is tested. The smaller 'k' the more focus is on detailed structures, while a large k will test overall changes. K-nearest neighbours (KNN) are determined in the subspaces before integration defined by 'dim_red'. The same set of KNN are used to determine LDF before and after integration.

Value

A data.frame with difference in LDF as column named "diff_ldf".

References

Latecki, Longin Jan and Lazarevic, Aleksandar and Pokrajac, Dragoljub (2007). Outlier Detection with Kernel Density Functions. Mach. Learn. Data Min. Pattern Recognit.. Springer Berlin Heidelberg.

See Also

[ldfDiff](#), [.ldfKnn](#).

Other ldf functions: [ldfDiff](#)

Examples

```
library(SingleCellExperiment)
sim_list <- readRDS(system.file("extdata/sim50.rds", package = "CellMixS"))
sce <- sim_list[["batch20"]][, c(1:50, 300:350)]
sce_batch1 <- sce[,colData(sce)$batch == "1"]
sce_pre_list <- list("1" = sce_batch1)

ldf_1 <- ldfSce("1", sce_pre_list, sce, k = 10, group = "batch",
dim_combined = "MNN", n_dim = 5)
```

visCluster

visCluster

Description

Creates summary plots of metric scores for different groups/cluster.

Usage

```
visCluster(sce_cms, cluster_var, metric_var = "cms", violin = FALSE)
```

Arguments

sce_cms	A SingleCellExperiment object with the result scores (e.g. cms) to plot within colData(res_object).
cluster_var	Character. Name of the factor level variable to summarize metric scores on.
metric_var	Character Name of the metric scores to use. Default is "cms".
violin	A logical. If true violin plots are plotted, while the default (FALSE) will plot ridge plots.

Details

Plots summarized metric scores. This function is intended to visualize and compare metric scores among clusters or other dataset variables specified in 'cluster_var'.

Value

a ggplot object.

See Also

[visIntegration](#)

Other visualize functions: [visGroup](#)

Examples

```
library(SingleCellExperiment)

sim_list <- readRDS(system.file("extdata/sim50.rds", package = "CellMixS"))
sce <- sim_list[[1]][, c(1:30,300:320)]
sce_cms <- cms(sce, "batch", k = 20, n_dim = 2)

visCluster(sce_cms, "batch")
```

visGroup

visGroup

Description

Plot group label in a reduced dimensional plot.

Usage

```
visGroup(sce, group, dim_red = "TSNE")
```

Arguments

sce	A SingleCellExperiment object.
group	Character. Name of group/batch variable. Needs to be one of names(colData(sce)).
dim_red	Character. Name of embeddings to use as subspace for plotting. Default is "TSNE".

Details

Plots a reduced dimension plot colored by group parameter. The dimension reduction embedding can be specified, but only tsne embeddings will automatically be computed by runTSNE. Embeddings from data integration methods (e.g. mnn.correct) can be used as long as they are specified in reducedDimNames(sce).

Value

a ggplot object.

See Also

[visOverview](#), [visMetric](#)

Other visualize functions: [visCluster](#)

Examples

```
library(SingleCellExperiment)
sim_list <- readRDS(system.file("extdata/sim50.rds", package = "CellMixS"))
sce <- sim_list[[1]][, c(1:50, 300:350)]

visGroup(sce, "batch")
```

visHist	<i>visHist</i>
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Description

Plot pvalue histograms of metric score distributions

Usage

```
visHist(res_object, metric_prefix = "cms", n_col = 1)
```

Arguments

res_object	SingleCellExperiment object, matrix or data.frame. The SingleCellExperiment object should contain the result scores (e.g. cms) to plot in colData(res_object). Matrix or data frame should have result scores in columns and cells in rows.
metric_prefix	Character. Prefix to specify names of colData(sce) to be plotted. Applies only if 'res_object' is a SingleCellExperiment object. Default is 'cms'.
n_col	Numeric. Number of columns of the pval histogram.

Details

Plots metric score distribution similar to a pvalue histogram distribution. Without dataset-specific bias, cms scores should be approx. flat distributed. If 'res_object' is a matrix or data.frame, it will create a histogram for each column. If 'res_object' is a SingleCellExperiment object, it will create a histogram of all colData(res_object) that start with 'metric_prefix'.

Value

a ggplot object.

See Also

Other visualize metric functions: [visMetric](#), [visOverview](#)

Examples

```
library(SingleCellExperiment)
sim_list <- readRDS(system.file("extdata/sim50.rds", package = "CellMixS"))
sce <- sim_list[[1]][, c(1:50)]
sce_cms <- cms(sce, "batch", k = 20, n_dim = 2)
visHist(sce_cms)
```

visIntegration	<i>visIntegration</i>
----------------	-----------------------

Description

Creates a summary plot of metric scores (for different integration methods).

Usage

```
visIntegration(res_object, metric_prefix = "cms", violin = FALSE)
```

Arguments

<code>res_object</code>	SingleCellExperiment object, list, matrix or data.frame. The SingleCellExperiment object should contain the result scores (cms) to compare within <code>colData(res_object)</code> . List, matrix or data frame should have result scores in list elements resp. columns.
<code>metric_prefix</code>	Character. Prefix to specify names of <code>colData(sce)</code> to be compared. Applies only if 'res_object' is a SingleCellExperiment object. Default is 'cms'.
<code>violin</code>	A logical. If true violin plots are plotted, while the default (FALSE) will plot ridge plots.

Details

Plots summarized cms scores from an SingleCellExperiment object, list or dataframe. This function is intended to visualize and compare different methods and views of the same dataset, not to compare different datasets.

Value

a ggplot object.

See Also

[visCluster](#), [ggridges](#)

Examples

```
library(SingleCellExperiment)

sim_list <- readRDS(system.file("extdata/sim50.rds", package = "CellMixS"))

sce <- sim_list[["batch20"]][, c(1:30,300:320)]
sce_mnn <- cms(sce,"batch", k = 20, dim_red = "MNN", res_name = "MNN",
n_dim = 2)

visIntegration(sce_mnn, metric_prefix = "cms.", violin = TRUE)
```

`visMetric`*visMetric*

Description

Plot metric scores in a reduced dimensional plot.

Usage

```
visMetric(sce_cms, metric_var = "cms", dim_red = "TSNE",  
          log10_val = FALSE)
```

Arguments

<code>sce_cms</code>	A <code>SingleCellExperiment</code> object with the result scores (e.g. <code>cms</code>) to plot within <code>colData(res_object)</code> .
<code>metric_var</code>	Character Name of the metric scores to use. Default is "cms".
<code>dim_red</code>	Character. Name of embeddings to use as subspace for plotting. Default is "TSNE".
<code>log10_val</code>	Logical. Indicating if $-\log_{10}(\text{metric})$ should be plotted.

Details

Plots a reduced dimension plot colored by metric scores. The dimension reduction embedding can be specified, but only tsne embeddings will automatically be computed using `runTSNE`. Embeddings from data integration methods (e.g. `mnn.correct`) can be used as long as they are present in `reducedDimNames(sce)`.

Value

a `ggplot` object.

See Also

[visOverview](#), [visGroup](#)

Other visualize metric functions: [visHist](#), [visOverview](#)

Examples

```
library(SingleCellExperiment)  
sim_list <- readRDS(system.file("extdata/sim50.rds", package = "CellMixS"))  
sce <- sim_list[[1]][, c(1:30, 300:320)]  
sce_cms <- cms(sce, "batch", k = 20, n_dim = 2)  
  
visMetric(sce_cms)
```

visOverview

*visOverview***Description**

Plot an overview of metric results, group label and any colData variable in a reduced dimensional representation.

Usage

```
visOverview(sce_cms, group, metric_prefix = "cms", dim_red = "TSNE",
            log10_val = FALSE, other_var = NULL)
```

Arguments

sce_cms	A SingleCellExperiment object with the result scores (e.g. cms) to plot in colData(sce_cms).
group	Character. Name of group/batch variable. Needs to be one of names(colData(sce)).
metric_prefix	Character. Prefix to specify names of colData(sce) to be plotted. Default is 'cms'.
dim_red	Character. Name of embeddings to use as subspace for plotting. Default is "TSNE".
log10_val	Logical. Indicating if -log10(metric) should be plotted.
other_var	Character string. Name(s) of other variables to be plotted asided. Need correspond to one of colData(sce).

Details

Plots reduced dimensions of cells colored by group variable and metric score. If 'red_dim' is not defined in reducedDimNames(sce) a tsne is calculated using runTSNE. Other color label as celltype label or smoothed scores can be plotted asided. Embeddings from data integration methods (e.g. mnn.correct) can be used if they are specified in reducedDimNames(sce).

Value

a ggplot object.

See Also

[visMetric](#), [visGroup](#)

Other visualize metric functions: [visHist](#), [visMetric](#)

Examples

```
library(SingleCellExperiment)
sim_list <- readRDS(system.file("extdata/sim50.rds", package = "CellMixS"))
sce <- sim_list[[1]][, c(1:30, 300:330)]
sce_cms <- cms(sce, "batch", k = 20, n_dim = 2)

visOverview(sce_cms, "batch", other_var = "batch")
```


Index

.cmsCell, [2](#), [4–6](#), [8](#)
.defineSubspace, [3](#), [3](#), [4–6](#)
.filterLocMin, [3](#), [4](#), [4](#), [5](#), [6](#)
.ldfKnn, [3](#), [4](#), [5](#), [6](#), [9](#), [11](#)
.smoothCms, [3–5](#), [6](#), [8](#)

ad.test, [3](#)

BiocParallelParam, [7](#)

CellMixS-package, [2](#)
cms, [2–4](#), [6](#), [7](#)

ldfDiff, [2](#), [8](#), [11](#)
ldfSce, [4](#), [5](#), [9](#), [9](#)

visCluster, [11](#), [12](#), [14](#)
visGroup, [11](#), [12](#), [15](#), [16](#)
visHist, [13](#), [15](#), [16](#)
visIntegration, [11](#), [14](#)
visMetric, [12](#), [13](#), [15](#), [16](#)
visOverview, [12](#), [13](#), [15](#), [16](#)