

RSVSim

an R/Bioconductor package for the simulation of structural variations

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1 Introduction

The simulation of structural variations (SV) is an important measure to assess the performance of algorithms dealing with SVs and their detection and can help with the design of sequencing experiments. A simulation generates a base exact ground truth, which can be used to test the sensitivity and precision of SV callers.

A FASTA-file with the simulated, rearranged genome can be used by common, published read simulators (like [Huang *et al.*, 2011]), [Hu *et al.*, 2012]) to generate NGS datasets from various platforms that can then be used to asses an SV algorithm . A typical workflow consists of

SV simulation \Rightarrow (Paired-End) Read simulation \Rightarrow SV algorithm \Rightarrow Evaluation

Varying parameters of the SV simulation like SV type, size or location and of the read simulator like number of reads (coverage), insert-size (for paired-end) or read length can give helpful information for future sequencing experiment designs.

This package addresses the very first step of SV simulation and provides the following features:

- Simulation of deletions, insertions, inversions, tandem duplications and translocations (balanced and unbalanced) of various sizes
- Rearrangement of the human genome (hg19) by default or any other kind of genome available as FASTA file or *BSgenome* package
- Non-overlapping positioning of SV breakpoints within the whole genome or only a subset (e.g. coding, non-coding or low-complexity regions)
- Implementation of, e.g. previously detected or known, SVs at user-supplied coordinates
- Uniform distribution of SV breakpoints or simulation of biases towards repeat regions and regions of high homology according to different SV formation mechanisms (for hg19 only)
- Simulation of smaller mutations (SNPs and indels) close to the SV breakpoint
- Estimation of SV size distribution from real datasets
- Comparison of SV simulation with results from SV detection algorithms

1.1 Loading the package

After installation, the package can be loaded into R by typing

```
> library(RSVSim)
```

into the R console.

RSVSim requires the R-packages *Biostrings*, *IRanges*, *GenomicRanges* and *ShortRead*. Mainly for efficient and convenient storing and access of sequences and genomic coordinates. The packages *BSgenome.Hsapiens.UCSC.hg19*, *GenomicFeatures*, *rtracklayer* and *MASS* are suggested for certain functionalities.

2 Structural variation simulation

The main function for simulation is called *simulateSV*. The simulation works pretty similar for every different SV type by specifying number and size of the variation(s) and (optionally) the regions, where to place the variation (randomly or not). The size can be either one value for every SV type or a vector of values for every single SV.

The following sections give a short example for every SV type using a simple toy example with two chromosomes of 40bp each:

```
> genome = DNASTringSet(  
  c("AAAAAAAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTTTTTT",  
    "GGGGGGGGGGGGGGGGGGGGCCCCCCCCCCCCCCCCCCCC")  
> names(genome) = c("chr1", "chr2")  
> genome
```

DNAStrngSet object of length 2:

	width	seq	names
[1]	40	AAAAAAAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTTTTT	chr1
[2]	40	GGGGGGGGGGGGGGGGGGGGCCCCCCCCCCCCCCCCCCCC	chr2

The genome has to be a named DNAStrngSet or a filename that points to a FASTA-file saved somewhere on disk. By default, when omitting the genome parameter, simulateSV will load the human genome (hg19) automatically. This requires an installation of the R-package *BSgenome.Hsapiens.UCSC.hg19*. When using other *BSgenome* packages, it is recommended to extract the desired sequences first and combine them into a named DNAStrngSet. For example, the preparation of the genome of an Ecoli strain (str. 536) would look like:

```
> library(BSgenome.Ecoli.NCBI.20080805)
> genome = DNAStrngSet(Ecoli[["NC_008253"]])
> names(genome) = "NC_008253"
```

Each implemented SV will be reported with its position in the "normal" reference genome, and the break-point sequences. The rearranged genome is returned as DNAStrngSet and the SV information is stored in its metadata slot as a named list of data.frames. All this can also be written to disk by specifying an output directory (which is the current directory by default). The SV tables are saved as CSV files (called deletions.csv, insertions.csv etc.) and the genome in FASTA format (genome_rearranged.fasta).

Note, that the seeds for the randomizations in the following examples were set for demonstration purposes only. The seed parameter can be omitted or used to reproduce the same simulation several times.

The parameter output is set to NA in all examples to avoid wrting the output to disc. The parameter verbose is set to FALSE to suppress progress information about the simulation (which is enabled by default).

2.1 Deletions

A segment is cut out from the genome. The following example generates three deletions of 10bp each:

```
> sim = simulateSV(output=NA, genome=genome, dels=3, sizeDels=10,
  bpSeqSize=6, seed=456, verbose=FALSE)
> sim
```

DNAStrngSet object of length 2:

	width	seq	names
[1]	20	AAAAAAAAAAAAAAAAAAAAAATT	chr1
[2]	30	GGGGGGGGGGGGGGGGGGGGCCCCCCCC	chr2

```
> metadata(sim)
```

```
$deletions
      Name Chr Start End Size BpSeq
1 deletion1 chr1  21  30  10  ATT
2 deletion2 chr2  31  40  10  CCC
3 deletion3 chr1  19  28  10 AAATT
```

Chromosome 2, which harbours two deletions, is now 10bp shorter than chromosome 1. The breakpoint sequence of 6bp shows the 3bp up- and downstream of the deletion breakpoint in the rearranged genome.

2.5 Translocations

A segment from the 5' or 3' end of one chromosome A is exchanged with the 5' or 3' end of another chromosome B. If it is not balanced, the segment from chromosome B will be lost, what results in a duplicated sequence from chromosome A. The parameter `percBalancedTrans` sets the amount of balanced translocation (0-1, i.e. 0%-100%); by default, all translocations will be balanced. Segments which are translocated between two different ends (5'↔3' of 3'↔5') are always inverted. After random generation of the breakpoint, the translocation spans the chromosome until the closest of both ends (which may include the centromere in the human genome).

```
> sim = simulateSV(output=NA, genome=genome,trans=1, bpSeqSize=6, seed=123, verbose=FALSE)
> sim
```

DNAStrngSet object of length 2:

	width	seq	names
[1]	39	AAAAAAAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTCCC	chr1
[2]	41	AAAAGGGGGGGGGGGGGGGGGGGGGCCCCCCCCCCCCCCCCCCC	chr2

```
> metadata(sim)
```

```
$translocations
```

	Name	ChrA	StartA	EndA	SizeA	ChrB	StartB	EndB	SizeB	Balanced	BpSeqA	BpSeqB
1	translocation_1	chr2	1	3	3	chr1	37	40	4	TRUE	AAAGGG	TTCCC

This example exchanges the last 3bp of chromosome 2 with the first 19bp of chromosome 1. Both sequences were inverted.

The same example in an unbalanced fashion:

```
> sim = simulateSV(output=NA, genome=genome,trans=1, percBalancedTrans=0,
bpSeqSize=6, seed=123, verbose=FALSE)
> sim
```

DNAStrngSet object of length 2:

	width	seq	names
[1]	39	AAAAAAAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTCCC	chr1
[2]	40	GGGGGGGGGGGGGGGGGGGGGGCCCCCCCCCCCCCCCCCCC	chr2

```
> metadata(sim)
```

```
$translocations
```

	Name	ChrA	StartA	EndA	SizeA	ChrB	StartB	EndB	SizeB	Balanced	BpSeqA	BpSeqB
1	translocation_1	chr2	1	3	3	chr1	37	40	4	FALSE		TTCCC

The sequence from chr1:1-19 is lost, while there are now two copies of the translocated (inverted) segment from chr2:38-40. The breakpoint sequence for chr2 (denoted by chrA in general) is not reported, since this chromosome is not altered.

3 Simulation of biases towards SV formation mechanisms and repeat regions

By default, the SV breakpoints are placed uniformly across the genome. Several studies have shown, that structural variation formation in the human genome is not a random process but rather the

result of mechanisms such as nonallelic homologous recombination (NAHR), nonhomologous recombination (NHR), variable number of tandem repeats (VNTRs) and transposable element insertions (TEIs) ([Mills *et al.*, 2011], [Pang *et al.*, 2013]). These mechanisms can be further associated with repeat elements and regions of high homology such as LINEs, SINEs, Micro-/Minisatellites and segmental duplications ([Lam *et al.*, 2010]).

Using the default genome hg19 and setting `repeatBias` to `TRUE`, *RSVSim* simulates a bias of breakpoint positioning towards repeat regions. This is done in two steps:

1. Weighting SV formation mechanisms (here: NAHR, NHR, VNTR, TEI, Other) for each SV type. The type "Other" can be used for any individual weighting of repeats; by default, "Other" is associated with a random breakpoint.
2. Weighting each SV formation mechanism for each kind of repeat. The following types of repeat regions are supported: LINE/L1, LINE/L2, SINE/Alu, SINE/MIR, segmental duplications (SD), tandem repeats (TR; mainly micro-/minisatellites) and Random. The latter, "Random", means any region on the genome.

For the mechanism NAHR, both breakpoints will lie within a repeat region (with at least 50bp distance to the repeat margins), while for NHR, VNTR, TEI and Other, the repeat must make up for at least 75% of the SV region.

This feature is turned off automatically, when the user specifies his own genome (i.e. any genome other than hg19).

The default weights for SV mechanisms for deletions, insertions and duplications are based on figure 4b in [Mills *et al.*, 2011]. The weights for inversions refer to figure 3c in [Pang *et al.*, 2013]. The mechanisms and breakpoint sequences of translocations have not been studied as extensively as for other kinds of SVs. The default weights for translocations were chosen according to some exemplary publications ([Ou *et al.*, 2011], [Chen *et al.*, 2008]), so that NAHR, NHR and random breakpoint positioning contribute equally. In all cases, the results for SVs >1.000bp were used. The exact weights are:

	dels	ins	invs	dups	trans
NAHR	0.23	0.10	0.65	0.10	0.33
NHR	0.69	0.03	0.35	0.03	0.33
TEI	0.04	0.82	0.00	0.82	0.00
VNTR	0.04	0.05	0.00	0.05	0.00
Other	0.00	0.00	0.00	0.00	0.34

The default weights for repeat regions for every SV mechanism were based on the enrichment analysis in [Lam *et al.*, 2010] (see their supplemental table 5). The exact values are:

```
> data(weightsRepeats, package="RSVSim")
> show(weightsRepeats)
```

	NAHR	NHR	TEI	VNTR	Other
L1	0.59	1.04	1.66	0	0
L2	0.13	0.62	0.25	0	0
Alu	2.72	1.16	0.47	0	0
MIR	0.14	0.74	0.14	0	0
SD	5.95	2.06	0.57	0	0
TR	0.00	0.00	0.00	1	0
Random	1.00	1.00	1.00	0	1

The user may provide other weights by passing his own `data.frames`, using the function arguments `weightsMechanisms` and `weightsRepeats`. The structure of the `data.frames` has to be identical to the default ones shown above (i.e. same dimensions, column and row names). The effect of the weights

is comparable to the `prob` argument in the R function `sample`.

For example would exclude tandem repeats, segmental duplications and random regions from the simulation (except for VNTRs) by setting their weights to zero for all mechanisms. NAHRs, would be related to SINEs only. For `weightsMechanisms`, the default values will be used, because the argument is missing here. Note, that `repeatBias=TRUE` has to be set to use this feature.

This feature requires the coordinates of repeat regions for hg19. This can be handled in two ways:

- *RSVSim* downloads the coordinates once automatically from the UCSC Browser's RepeatMasker track (which may take up to 45 Minutes!).
- The user may specify the filename of a RepeatMasker output file downloaded from their homepage ([Smit *et al.*, 1996-2010]): <http://www.repeatmasker.org/species/homSap.html> (e.g. hg19.fa.out.gz). Loading this file takes only a few minutes.

In both cases, RSVSim saves the coordinates as RData object `repeats_hg19.RData` to the RSVSim installation directory for a faster access in the future (if write privileges allow to do so). After that, one of the two steps mentioned above is not necessary anymore and next time, *RSVSim* is going to load the coordinates automatically from the RData file.

When loading the repeats, neighboured ones with a distance up to 50bp will be merged, to obtain larger repeat regions and to allow SVs to span more than one repeat. But, breakpoints will only be placed within repeats of the same type (e.g. LINE/L1-LINE/L1, or SINE/MIR-SINE/MIR etc.).

4 Simulation of additional breakpoint mutations

SV breakpoints are usually not clean but tend to co-occur with other, usually smaller mutations, such as indels or SNPs. *RSVSim* allows to randomly generate additional SNPs and indels within the flanking regions of each breakpoint. Their generation can be configured by the four arguments `bpFlankSize`, `percSNPs`, `indelProb` and `maxIndelSize`, which specify the size of the flanking regions in bp (i.e. proximity of the mutations to the breakpoint), the fraction on SNPs (in %), the probability of an indel (insertions and deletions are equally likely) and the maximum size of an indel (size is selected randomly between 1 and `maxIndelSize`). Each flanking region may only contain one indel. SNPs can affect 0-100% of the region. By default, this feature is turned off.

The following example creates one deletion with 10% SNPs and 100% indel probability within 10bp up-/downstream of the breakpoint:

```
> sim = simulateSV(output=NA, genome=genome, dels=1, sizeDels=5, bpFlankSize=10,
  percSNPs=0.25, indelProb=1, maxIndelSize=3, bpSeqSize=10, seed=123, verbose=FALSE)
> sim
```

DNASTringSet object of length 2:

	width	seq	names
[1]	40	AAAAAAAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTTTT	chr1
[2]	34	GGGGGTGGGGGGCCCCCCCCCCCCCCCCCCCC	chr2

```
> metadata(sim)
```

\$deletions

	Name	Chr	Start	End	Size	BpSeq
1	deletion1	chr2	3	7	5	GGGGGT

In addition to the 5bp deletion of the sequence CCCC, two SNPs C- >T and C- >A, and a deletion of two more Cs were added up- and downstream of the breakpoint.

5 Simulation within a genome subset

It is possible to run the simulation to certain chromosomes only by specifying the chromosome names in the parameter `chrs`. It has to be taken care that these chromosome names match the names in the `DNAStrngSet` containing the genome sequences (e.g. "chr1", "chr2", ..., "chrY" for the default genome hg19 from the package *BSgenome.Hsapiens.UCSC.hg19*).

Furthermore, every SV has it's own parameter to restrict the simulation to a desired set of genomic regions: `regionsDels`, `regionsIns`, `regionsInvs`, `regionsDups` and `regionsTrans`. Each one being a `GRanges` object with a chromosome name, start- and end-position.

The following example places randomly four inversions into the second half of chr1 and the first half of chr2:

```
> regions = GRanges(IRanges(c(21,1),c(40,20)), seqnames=c("chr1","chr2"))
> regions
```

GRanges object with 2 ranges and 0 metadata columns:

```
      seqnames      ranges strand
     <Rle> <IRanges> <Rle>
[1]   chr1      21-40      *
[2]   chr2       1-20      *
```

seqinfo: 2 sequences from an unspecified genome; no seqlengths

```
> sim = simulateSV(output=NA, genome=genome, invs=4, sizeInvs=5,
  regionsInvs=regions, bpSeqSize=6, seed=2345, verbose=FALSE)
> sim
```

DNAStrngSet object of length 2:

```
      width seq                                     names
[1]   40 AAAAAAAAAAAAAAAAAAAAAAAAAATAAAAATAAAATTTTTTTT      chr1
[2]   40 CCCCCCCCCCGGGGGGGGGCCCCCCCCCCCCCCCCCCCC      chr2
```

```
> metadata(sim)
```

\$inversions

```
      Name Chr Start End Size BpSeq_3prime BpSeq_5prime
1 inversion1 chr1  22  26   5      AAATAA      AATAAA
2 inversion2 chr1  28  32   5      AAATTT      AATAAA
3 inversion3 chr2   6  10   5      CCCGGG      CCCCC
4 inversion4 chr2   1   5   5      CCCCC
```

For translocations, the regions only say where to place the breakpoint, since the translocated region spans the chromosome until the closest of both ends.

Some applications may focus on certain parts of the hg19 only, like exons, introns or transcripts. The package *GenomicFeatures* provides functionalities to export such coordinates from the UCSC Genome Browser to R (see for example `makeTxDbFromUCSC`, `exonsBy`, `intronsBy`, `transcriptsBy`). In the following example, 100 deletions would be placed somewhere in the exonic regions on hg19:

```
> transcriptDB = makeTxDbFromUCSC(genome = "hg19",tablename = "knownGene")
> exons = exonsBy(transcriptDB)
> exons = unlist(exons)
> exons = GRanges(IRanges(start=start(exons), end=end(exons)), seqnames=seqnames(exons))
> simulateSV(output=NA, dels=100, regionsDels=exons, sizeDels=1000, bpSeqSize=50)
```

SVs will not be placed within unknown regions or assembly gaps denoted by the letter N. Such regions are detected and filtered automatically.

5.1 Inserting a set of SVs

The simulation allows to turn off the random generation of breakpoints and to insert a set of (for example previously detected or known) SVs. It works by using the same regions parameters and setting the parameter `random` to `FALSE`. This may also be a vector of five `TRUE/FALSE` values (in the order: deletions, insertions, inversions, tandem duplications, translocations) if some SVs shall be generated randomly and others not.

The example below inserts a deletion at `chr2:16-25`:

```
> knownDeletion = GRanges(IRanges(16,25), seqnames="chr2")
> names(knownDeletion) = "myDeletion"
> knownDeletion
```

GRanges object with 1 range and 0 metadata columns:

```
      seqnames      ranges strand
      <Rle> <IRanges> <Rle>
myDeletion   chr2     16-25     *
```

seqinfo: 1 sequence from an unspecified genome; no seqlengths

```
> sim = simulateSV(output=NA, genome=genome, regionsDels=knownDeletion,
  bpSeqSize=10, random=FALSE, verbose=FALSE)
> sim
```

DNASTringSet object of length 2:

```
      width seq
[1]    40 AAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTTTT
[2]    30 GGGGGGGGGGGGGGCCCCCCCCCCCCCCCC
```

names

```
chr1
chr2
```

```
> metadata(sim)
```

\$deletions

```
      Name Chr Start End Size BpSeq
myDeletion myDeletion chr2  16 25  10 GGGGGCCCCC
```

Note, that the output adopts the names, that were given the `GRanges` object of the inserted SV(s).

It's a little different for insertions and translocations, since they involve two genomic regions. Thus, the `GRanges` object for `regionsIns` has to be extended by columns `chrB` and `startB`, saying, that the sequence within ranges of the `GRanges` object will be inserted at `chrB:startB`.

The next example inserts the sequence from `chr1:16:25` at `chr2:26`:

```
> knownInsertion = GRanges(IRanges(16,25),seqnames="chr1", chrB="chr2", startB=26)
> names(knownInsertion) = "myInsertion"
> knownInsertion
```

GRanges object with 1 range and 2 metadata columns:

```
      seqnames      ranges strand |      chrB      startB
      <Rle> <IRanges> <Rle> | <character> <numeric>
myInsertion   chr1     16-25     * |      chr2         26
```

seqinfo: 1 sequence from an unspecified genome; no seqlengths

```
> sim = simulateSV(output=NA, genome=genome, regionsIns=knownInsertion,
  bpSeqSize=10, random=FALSE, verbose=FALSE)
> sim
```


BEDPE format (<http://code.google.com/p/bedtools>). Deletions, inversions and tandem duplications, which affect one region on the genome, can be either given in both formats. Translocations and insertions, which affect to regions on the genome, require the BEDPE-format. Eventually, the output of the SV detection format has to be converted accordingly (for example in R).

The function only compares one SV type at a time, so `querySVs` and `simSVs` may not contain a mixture of different kinds of SVs.

If the BED-tables for `querySVs` or the simulation output are saved on disk, `compareSV` also accepts their filenames and loads the tables automatically as `data.frame` in R.

The following example simulates first five 5bp deletions in the small toy genome defined above:

```
> sim = simulateSV(output=NA, genome=genome, dels=5, sizeDels=5,
  bpSeqSize=10, seed=2345, verbose=FALSE)
> simSVs = metadata(sim)$deletions
> simSVs
```

	Name	Chr	Start	End	Size	BpSeq
1	deletion1	chr1	34	38	5	TTTTTTT
2	deletion2	chr1	2	6	5	AAAAAA
3	deletion3	chr2	22	26	5	GGGGCCCCC
4	deletion4	chr2	31	35	5	CCCCCCCCC
5	deletion5	chr1	12	16	5	AAAAAAAAT

An SV detection in BED format (the `querySVs`) may look like this: Four of five deletions were detected, two with exact and two with an approximate breakpoint. Two additional deletions were detected, which were not part of the simulation.

```
> querySVs = data.frame(
  chr=c("chr1", "chr1", "chr1", "chr2", "chr2"),
  start=c(12, 17, 32, 2, 16),
  end=c(15, 24, 36, 6, 20),
  bpSeq=c("AAAAAAAAT", "AAAAAATTT", "TTTTTTTTT",
    "GGGGGGGGG", "GGGGGCCCC")
)
> querySVs
```

	chr	start	end	bpSeq
1	chr1	12	15	AAAAAAAAT
2	chr1	17	24	AAAAAATTT
3	chr1	32	36	TTTTTTTTT
4	chr2	2	6	GGGGGGGGG
5	chr2	16	20	GGGGGCCCC

The column with the breakpoint sequence is optional, the column names not important (BED-files have no header).

A comparison with 0bp tolerance yields only two overlaps:

```
> compareSV(querySVs, simSVs, tol=0)
```

	Name	Chr	Start	End	Size	BpSeq	Overlap	OverlapBpSeq
1	deletion1	chr1	34	38	5	TTTTTTT		NA
2	deletion2	chr1	2	6	5	AAAAAA		NA

```

3 deletion3 chr2    22  26    5 GGGGCCCCCC      NA
4 deletion4 chr2    31  35    5 CCCCCCCCCC      NA
5 deletion5 chr1    12  16    5 AAAAAAAAAAT      NA

```

A higher breakpoint tolerance of +/- 3bp also includes more imprecise detections:

```

> compareSV(querySVs, simSVs, tol=3)

      Name Chr Start End Size      BpSeq  Overlap OverlapBpSeq
1 deletion1 chr1   34  38    5  TTTTTTT chr1:32-36      100
2 deletion2 chr1    2   6    5   AAAAAA      NA
3 deletion3 chr2   22  26    5 GGGGCCCCCC      NA
4 deletion4 chr2   31  35    5 CCCCCCCCCC      NA
5 deletion5 chr1   12  16    5 AAAAAAAAAAT chr1:12-15      90

```

Note that for deletion1, the breakpoint sequence matched only by 80%.

The second example compares translocations:

```

> sim = simulateSV(output=NA, genome=genome, trans=2, percBalancedTrans=0.5,
bpSeqSize=10, seed=246, verbose=FALSE)
> simSVs = metadata(sim)$translocations
> simSVs

      Name ChrA StartA EndA SizeA ChrB StartB EndB SizeB Balanced      BpSeqA
1 translocation_1 chr1     1  13   13 chr2     1   2    2   FALSE
2 translocation_2 chr1   37  40    4 chr2    29  40   12   TRUE TTTTCCCCC
      BpSeqB
1 AAAAAGGGGG
2 CCCCTTTT

```

Detected translocations have to be given in BEDPE-format (i.e. at least six columns chr1, start1, end1, chr2, start2, end2 for the breakpoints on both chromosomes). In this example, the breakpoints were approximated up to 1bp or 2bp, optional breakpoint sequences are missing:

```

> querySVs = data.frame(
  chr=c("chr1", "chr1", "chr2"),
  start1=c(15,32,32),
  end1=c(18,36,33),
  chr2=c("chr2","chr2","chr1"),
  start2=c(10,31,32),
  end2=c(12,33,36)
)
> querySVs

      chr start1 end1 chr2 start2 end2
1 chr1     15  18 chr2     10  12
2 chr1     32  36 chr2     31  33
3 chr2     32  33 chr1     32  36

```

Here, all detected SVs span the simulated breakpoints:

```

> compareSV(querySVs, simSVs, tol=0)

```


The `minSize` and `maxSize` can be omitted; they are then calculated from the given set of `svSizes`. It is recommended to use a `minSize` and `maxSize` that is consistent with the minimum/maximum values in `svSizes`.

Setting the parameter `hist=TRUE` also plots a histogram of the SV sizes to give an impression of their distribution (see Fig.1).

For deletions, insertions, inversions and tandem duplications, `estimateSVSizes` can use default parameters for the beta distribution. They were estimated from the Database of Genomic Variants (DGV) release 2012-03-29 ([Iafrate *et al.*, 2004]). Hence, no set of SV sizes is needed for fitting the distribution. In total, 1.129 deletions, 490 insertions, 202 inversions and 145 tandem duplications between 500bp and 10kb were used to estimate the shape. The parameter `default` can be set to either "deletions", "insertions", "inversions" or "tandemDuplications" to use the according set of shape parameters:

```
> delSizes = estimateSVSizes(n=10000, minSize=500, maxSize=10000,
  default="deletions", hist=TRUE)
> head(delSizes, n=15)

[1] 1838 503 2240 1071 853 751 1147 671 629 776 1814 1773 517 502 581

> delSizes = estimateSVSizes(n=10000, minSize=500, maxSize=10000,
  default="insertions", hist=TRUE)
> head(delSizes, n=15)

[1] 833 1033 3816 1334 1723 2048 2521 2874 1877 509 6090 743 1387 1796 5053

> invSizes = estimateSVSizes(n=10000, minSize=500, maxSize=10000,
  default="inversions", hist=TRUE)
> head(invSizes, n=15)

[1] 2976 1769 6532 1991 1083 1678 2841 1886 4044 1037 649 2563 771 3487 1213

> delSizes = estimateSVSizes(n=10000, minSize=500, maxSize=10000,
  default="tandemDuplications", hist=TRUE)
> head(delSizes, n=15)

[1] 6742 1260 762 5371 2623 1483 1593 1801 1388 559 1107 3039 649 2442 755
```

See Fig.2, Fig.3, Fig.4 and Fig.5 to see the estimated distribution based on the SVs in the DGV. When using these default values, it is recommended to simulate SVs that do not differ too much in size (around 500bp-10kb).

8 Runtime

The runtime of *RSVSim* mainly depends on the number of simulated breakpoints and the size of the genome.

The following test case simulates 50 SVs (10 per SV type) on the complete hg19:

```
> simulateSV(output=NA, dels=10, ins=10, inv=10, dups=10, trans=10,
  sizeDels=10000, sizeIns=10000, sizeInvs=10000, sizeDups=10000,
  repeatBias=FALSE, bpFlankSize=50, percSNPs=0.25, indelProb=0.5, maxIndelSize=10)
```

Ten repetitions of the simulation yield an average time of 6 minutes on a single Intel Xeon CPU with 2.40GHz an R version 2.15.2 (2012-10-26) including loading of the hg19 and writing of the output to

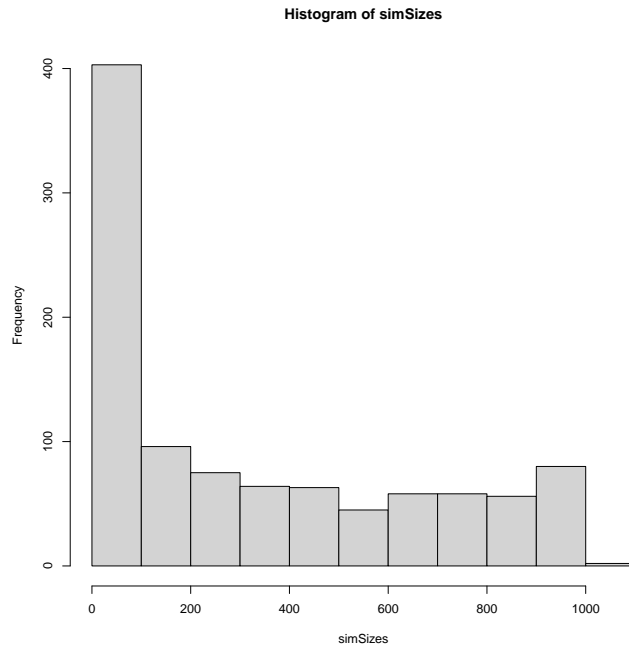


Figure 1: Distribution of 1.000 SV sizes drawn from a beta distribution using function `estimateSVSizes`.

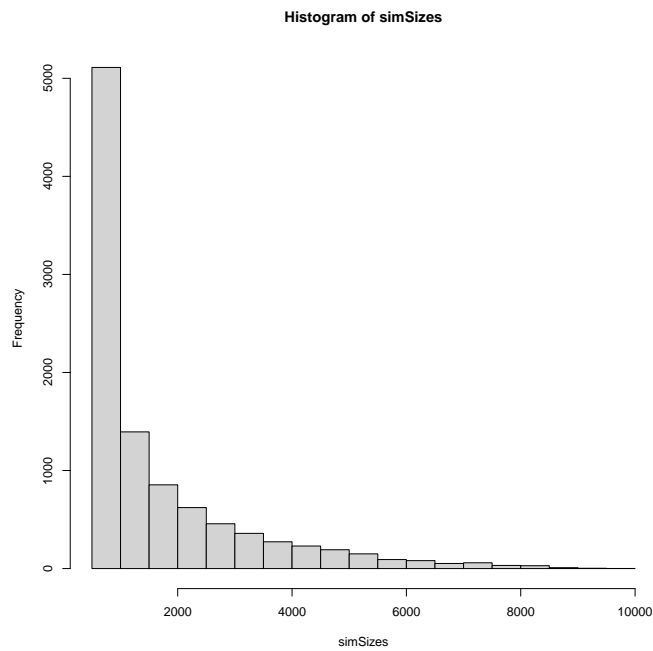


Figure 2: Distribution of 10.000 deletion sizes based on deletions from the Database of Genomic Variants.

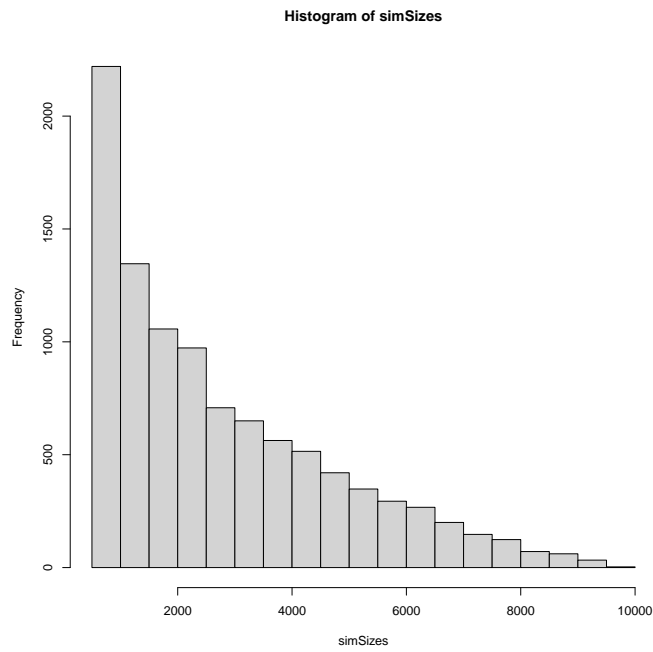


Figure 3: Distribution of 10.000 insertion sizes based on insertions from the Database of Genomic Variants.

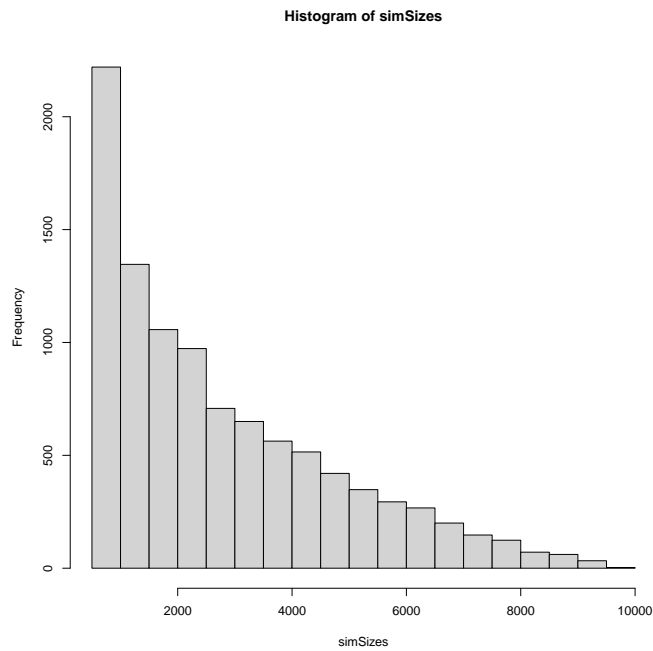


Figure 4: Distribution of 10.000 inversion sizes based on inversions from the Database of Genomic Variants.

disc.

Enabling biases for repeat regions for the same test case (i.e. `repeatBias=TRUE`), yield an average 7 minutes.

For other simulations on hg19, the runtime will scale linearly with the number of SV breakpoints.

Note, that the one-time, initial download of the repeat coordinates for hg19 from the UCSC browser may take up to 45 minutes. Alternatively, providing a RepeatMasker output file is much quicker (see section 3 for more details).

References

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- [Smit *et al.*, 1996-2010] Smit A. *et al* (1996-2010) RepeatMasker Open-3.0., <<http://www.repeatmasker.org>>.

9 Session Information

R version 4.0.3 (2020-10-10)

Platform: x86_64-pc-linux-gnu (64-bit)

Running under: Ubuntu 18.04.5 LTS

Matrix products: default

BLAS: /home/biocbuild/bbs-3.12-bioc/R/lib/libRblas.so

LAPACK: /home/biocbuild/bbs-3.12-bioc/R/lib/libRlapack.so

locale:

[1] LC_CTYPE=en_US.UTF-8	LC_NUMERIC=C	LC_TIME=en_US.UTF-8
[4] LC_COLLATE=C	LC_MONETARY=en_US.UTF-8	LC_MESSAGES=en_US.UTF-8
[7] LC_PAPER=en_US.UTF-8	LC_NAME=C	LC_ADDRESS=C
[10] LC_TELEPHONE=C	LC_MEASUREMENT=en_US.UTF-8	LC_IDENTIFICATION=C

attached base packages:

[1] stats4 parallel stats graphics grDevices utils datasets methods
[9] base

other attached packages:

[1] MASS_7.3-53 RSVSim_1.30.0 GenomicRanges_1.42.0 GenomeInfoDb_1.26.0
[5] Biostrings_2.58.0 XVector_0.30.0 IRanges_2.24.0 S4Vectors_0.28.0
[9] BiocGenerics_0.36.0

loaded via a namespace (and not attached):

[1] zlibbioc_1.36.0 GenomicAlignments_1.26.0 BiocParallel_1.24.0
[4] lattice_0.20-41 jpeg_0.1-8.1 hwriter_1.3.2
[7] tools_4.0.3 SummarizedExperiment_1.20.0 grid_4.0.3
[10] Biobase_2.50.0 png_0.1-7 latticeExtra_0.6-29
[13] matrixStats_0.57.0 crayon_1.3.4 Matrix_1.2-18
[16] GenomeInfoDbData_1.2.4 RColorBrewer_1.1-2 bitops_1.0-6
[19] RCurl_1.98-1.2 DelayedArray_0.16.0 compiler_4.0.3
[22] MatrixGenerics_1.2.0 Rsamtools_2.6.0 ShortRead_1.48.0

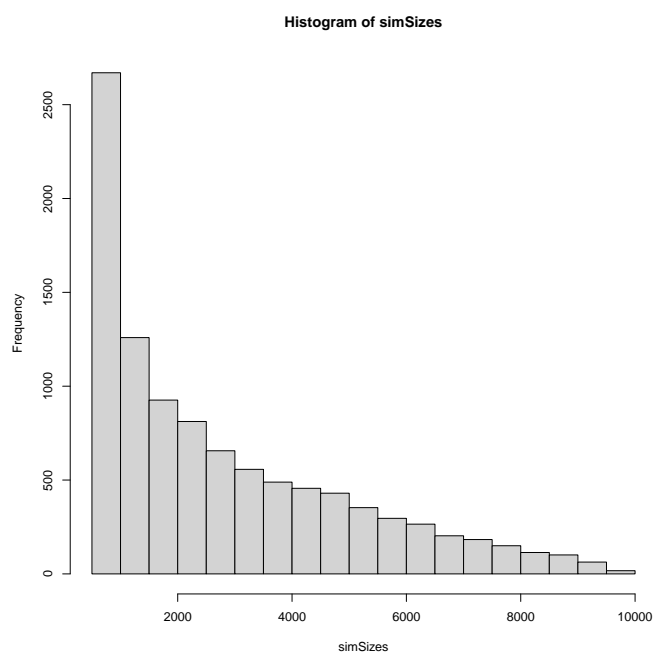


Figure 5: Distribution of 10,000 tandem duplication sizes based on tandem duplications from the Database of Genomic Variants.