Using *R* and *Bioconductor* for Proteomics Data Analysis

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Abstract

This vignette shows and executes the code presented in the manuscript *Using R for proteomics data analysis*. It also aims at being a general overview for users who wish to explore the *R* environment and programming language for the analysis of proteomics data.

Keywords: proteomics, mass spectrometry, tutorial.

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Contents

1 Introduction

This document illustrates some existing *R* infrastructure for the analysis of proteomics data. It presents the code for the use cases taken from [1]. A pre-print of the manuscript is available on arXiv[1].

There are however numerous additional *R* resources distributed by the *Bioconductor*[2] and CRAN[3] repositories, as well as packages hosted on personal websites. Section 8 on page 45 tries to provide a wider picture of available packages, without going into details.

1.1 General R resources

The reader is expected to have basic *R* knowledge to find the document helpful. There are numerous *R* introductions freely available, some of which are listed below.

From the *R* project web-page:

- **An Introduction to R** is based on the former *Notes on R*, gives an introduction to the language and how to use R for doing statistical analysis and graphics. [browse HTML — download PDF]
- Several introductory tutorials in the contributed documentation section.
- The `TeachingMaterial` repository[4] contains several sets of slides and vignettes about *R* programming.

Relevant background on the *R* software and its application to computational biology in general and proteomics in particular can also be found in [1]. For details about the *Bioconductor* project, the reader is referred to [2].

1.2 Bioconductor resources

The Bioconductor offers many educational resources on its help page `http://bioconductor.org/help/`, in addition the package's vignettes (vignettes are a requirement for Bioconductor packages). We want to draw the attention to the Bioconductor work flows that offer a cross-package overview about a specific topic. In particular, there is now a *Mass spectrometry and proteomics data analysis*[5] work flow.

1.3 Getting help

All *R* packages come with ample documentation. Every command (function, class or method) a user is susceptible to use is documented. The documentation can be accessed by preceding the command by a `?` in the *R* console. For example, to obtain help about the `library` function, that will be used in the next section, one would type `?library`. In addition, all *Bioconductor* packages come with at least one vignette (this document is the vignette that comes with the *RforProteomics* package), a document that combines text and *R* code that is executed before the pdf is assembled. To look up all vignettes that come with a package, say *RforProteomics* and then open the vignette of interest, one uses the `vignette` function as illustrated below. More details can be found in `?vignette`.

```
## list all the vignettes in the RforProteomics package
vignette(package = "RforProteomics")
## Open the vignette called RforProteomics
vignette("RforProteomics", package = "RforProteomics")
## or just
vignette("RforProteomics")
```

[1] http://arxiv.org/abs/1305.6559
[2] http://www.bioconductor.org
[3] http://cran.r-project.org/web/packages/
[4] https://github.com/lgatto/TeachingMaterial
[5] http://bioconductor.org/help/workflows/proteomics/

R has several mailing lists[6]. The most relevant here being the main `R-help` list, *for discussion about problem and solutions using R*, ideal for general *R* content and is not suitable for bioinformatics or proteomics questions. *Bioconductor* also offers several resources dedicated to bioinformatics matters and *Bioconductor* packages, in particular the main *Bioconductor* support forum[7] for Bioconductor-related queries. A dedicated *RforProteomics* Google group[8] also welcomes questions/comments/annoucements related to *R* and mass-spectrometry/proteomics, although the Bioconductor forum is the preferred channel.

It is advised to read and comply to the posting guides (here and here) to maximise the chances to obtain good responses. It is important to specify the software versions using the `sessionInfo()` functions (see an example output at the end of this document, on page 48). It the question involves some code, make sure to isolate the relevant portion and report it with your question, trying to make your code/example reproducible[9].

1.4 Installation

The package should be installed using as described below:

```
## only first time you install Bioconductor packages
source("http://www.bioconductor.org/biocLite.R")
## else
library("BiocInstaller")
biocLite("RforProteomics")
```

To install all dependencies and reproduce the code in the vignette, replace the last line in the code chunk above with:)

```
biocLite("RforProteomics", dependencies = TRUE)
```

Finally, the package can be loaded with

```
library("RforProteomics")
```

```
##
## This is the 'RforProteomics' version 1.5.8.
##
##  To get started, visit
##    http://lgatto.github.com/RforProteomics/
##
##  or, in R, open package vignettes by typing
##    RforProteomics() # R/Bioc for proteomics overview
##    RProtVis()       # R/Bioc for proteomics visualisation
##
##  For a full list of available documents:
##    vignette(package='RforProteomics')
```

See also the *RforProteomics* web page[10] for more information on installation.

1.5 External dependencies

Some packages used in the document depend on external libraries that need to be installed prior to the *R* packages:

mzR depends on the Common Data Format[11] (CDF) to CDF-based raw mass-spectrometry data. On Linux, the `libcdf` library is required. On Debian-based systems, for instance, one needs to install the `libnetcdf-dev` package.

[6] http://www.r-project.org/mail.html
[7] https://support.bioconductor.org/
[8] https://groups.google.com/forum/#!forum/rbioc-sig-proteomics
[9] https://github.com/hadley/devtools/wiki/Reproducibility
[10] http://lgatto.github.io/RforProteomics/
[11] http://cdf.gsfc.nasa.gov/

IPPD (and others) depend on the *XML* package which requires the `libxml2` infrastructure on Linux. On Debian-based systems, one needs to install `libxml2-dev`.

biomaRt performs on-line requests using the `curl`[12] infrastructure. On Debian-based systems, you one needs to install `libcurl-dev` or `libcurl4-openssl-dev`.

MSGFplus Is based on the MS-GF+ java program and thus requires Java 1.7[13] in order to work.

1.6 Obtaining the code

The code in this document describes all the examples presented in [1] and can be copy, pasted and executed. It is however more convenient to have it in a separate text file for better interaction with *R* (using ESS[14] for Emacs or RStudio[15] for instance) to easily modify and explore it. This can be achieved with the `Stangle` function. One needs the Sweave source of this document (a document combining the narration and the *R* code) and the `Stangle` then specifically extracts the code chunks and produces a clean *R* source file. If the package is installed, the following code chunk will create a `RforProteomics.R` file in your working directory containing all the annotated source code contained in this document.

```
## gets the vignette source
rnwfile <- system.file("doc/vigsrc/RforProteomics.Rnw",
                       package = "RforProteomics")
## produces the R file in the working directory
library("knitr")
purl(rnwfile, quiet = TRUE)
```

```
## [1] ".R"
```

Alternatively, you can obtain the `Rnw` file on the github page https://github.com/lgatto/RforProteomics/blob/master/inst/doc/vigsrc/RforProteomics.Rnw.

1.7 Prepare the working environment

The packages that we will depend on to execute the examples will be loaded in the respective sections. Here, we pre-load packages that provide general functionality used throughout the document.

```
library("RColorBrewer") ## Color palettes
library("ggplot2")  ## Convenient and nice plotting
library("reshape2") ## Flexibly reshape data
```

2 Data standards and input/output

2.1 The mzR package

2.1.1 Raw MS data

The *mzR* package [3] provides a unified interface to various mass spectrometry open formats. This code chunk, taken from the `openMSfile` documentation, illustrated how to open a connection to an raw data file. The example `mzML` data is taken from the *msdata* data package. The code below would also be applicable to an `mzXML`, `mzData` or `netCDF` file.

```
## load the required packages
library("mzR") ## the software package
```

[12] http://curl.haxx.se/
[13] https://java.com
[14] http://ess.r-project.org/
[15] http://rstudio.org/

```
library("msdata") ## the data package
## below, we extract the releavant example file
## from the local 'msdata' installation
filepath <- system.file("microtofq", package = "msdata")
file <- list.files(filepath, pattern="MM14.mzML",
                   full.names=TRUE, recursive = TRUE)
## creates a commection to the mzML file
mz <- openMSfile(file)
## demonstraction of data access
basename(fileName(mz))
```

```
## [1] "MM14.mzML"
```

```
isInitialized(mz)
```

```
## [1] TRUE
```

```
runInfo(mz)
```

```
## $scanCount
## [1] 112
##
## $lowMz
## [1] 0
##
## $highMz
## [1] 0
##
## $dStartTime
## [1] 270.334
##
## $dEndTime
## [1] 307.678
##
## $msLevels
## [1] 1
```

```
instrumentInfo(mz)
```

```
## $manufacturer
## [1] "Unknown"
##
## $model
## [1] "instrument model"
##
## $ionisation
## [1] "electrospray ionization"
##
## $analyzer
## [1] "mass analyzer type"
##
## $detector
## [1] "detector type"
```

```
## once finished, it is good to explicitely
## close the connection
close(mz)
```

mzR is used by other packages, like *MSnbase* [4], *TargetSearch* [5] and *xcms* [6, 7, 8], that provide a higher level abstraction to the data.

2.1.2 Identification data

The *mzR* package also provides very fast access to `mzIdentML` data by leveraging proteowizard's `C++` parser.

```
file <- system.file("mzid", "Tandem.mzid.gz", package="msdata")
mzid <- openIDfile(file)
mzid
```

```
## Identification file handle.
## Filename:  Tandem.mzid.gz
## Number of psms:  171
```

Once and `mzRident` identification file handle has been established, various data and metadata can be extracted, as illustrated below.

```
softwareInfo(mzid)
```

```
## [1] "xtandem x! tandem CYCLONE (2010.06.01.5) "
## [2] "ProteoWizard MzIdentML 3.0.6239 ProteoWizard"
```

```
enzymes(mzid)
```

```
##      name nTermGain cTermGain minDistance missedCleavages
## 1 Trypsin         H        OH           0               1
```

```
names(psms(mzid))
```

```
##  [1] "spectrumID"               "chargeState"
##  [3] "rank"                     "passThreshold"
##  [5] "experimentalMassToCharge" "calculatedMassToCharge"
##  [7] "sequence"                 "modNum"
##  [9] "isDecoy"                  "post"
## [11] "pre"                      "start"
## [13] "end"                      "DatabaseAccess"
## [15] "DBseqLength"              "DatabaseSeq"
## [17] "DatabaseDescription"      "acquisitionNum"
```

```
head(psms(mzid))[, 1:13]
```

```
##   spectrumID chargeState rank passThreshold
## 1   index=12           3    1         FALSE
## 2  index=285           3    1         FALSE
## 3   index=83           3    1         FALSE
## 4   index=21           3    1         FALSE
## 5  index=198           3    1         FALSE
## 6   index=13           2    1         FALSE
##   experimentalMassToCharge calculatedMassToCharge
## 1                903.7209               903.4032
## 2                792.3792               792.3899
## 3                792.5295               792.3899
## 4                850.0782               849.7635
## 5                527.2592               527.2849
## 6                724.8816               724.3771
##                    sequence modNum isDecoy post pre start
## 1  LCYIALDFDEEMKAAEDSSDIEK      2   FALSE    S   K   217
```

```
## 2   KDLYGNVVLSGGTTMYEGIGER      1   FALSE    L   R   292
## 3   KDLYGNVVLSGGTTMYEGIGER      1   FALSE    L   R   292
## 4 VIDENFGLVEGLMTTVHAATGTQK      1   FALSE    V   K   842
## 5          GVGGAIVLVLYDEMK      1   FALSE    R   R   297
## 6            HAVGGRYSSLLCK      1    TRUE    D   K   392
##   end
## 1 239
## 2 313
## 3 313
## 4 865
## 5 311
## 6 404
```

2.2 Handling MS^2 identification data with mzID

The *mzID* package allows to load and manipulate MS^2 data in the `mzIdentML` format. The main `mzID` function reads such a file and constructs an instance of class `mzID`.

```
library("mzID")
id <- mzID("http://psi-pi.googlecode.com/svn/trunk/examples/1_1examples/55merge_tandem.mzid")
```

```
## reading 55merge_tandem.mzid... DONE!
```

```
id
```

```
## An mzID object
##
## Software used:   X\!Tandem (version: x! tandem CYCLONE (2010.06.01.5))
##
## Rawfile:         D:/TestSpace/NeoTestMarch2011/55merge.mgf
##
## Database:        D:/Software/Databases/Neospora_3rndTryp/Neo_rndTryp_3times.fasta.pro
##
## Number of scans: 169
## Number of PSM's: 170
```

Multiple files can be parsed in one go, possibly in parallel if the environment supports it. When this is done an mzIDCollection object is returned:

```
ids <- mzID(c(
    "http://psi-pi.googlecode.com/svn/trunk/examples/1_1examples/55merge_tandem.mzid",
    "http://psi-pi.googlecode.com/svn/trunk/examples/1_1examples/55merge_omssa.mzid"))
ids
```

```
## An mzIDCollection object containing 2 samples
```

Peptides, scans, parameters, ... can be extracted with the respective `pepetides`, `scans`, `parameters`, ... functions. The `mzID` object can also be converted into a `data.frame` using the `flatten` function.

```
fid <- flatten(id)
names(fid)
```

```
##  [1] "spectrumid"               "spectrum title"
##  [3] "acquisitionnum"           "passthreshold"
##  [5] "rank"                     "calculatedmasstocharge"
##  [7] "experimentalmasstocharge" "chargestate"
##  [9] "x\\!tandem:expect"        "x\\!tandem:hyperscore"
```

```
## [11] "isdecoy"                  "post"
## [13] "pre"                      "end"
## [15] "start"                    "accession"
## [17] "length"                   "sequence"
## [19] "pepseq"                   "modified"
## [21] "modification"             "idFile"
## [23] "databaseFile"
```

```
dim(fid)
```

```
## [1] 171  23
```

3 Raw data abstraction with `MSnExp` objects

MSnbase [4] provides base functions and classes for MS-based proteomics that allow facile data and meta-data processing, manipulation and plotting (see for instance figure 1 on page 11).

```
library("MSnbase")
## uses a simple dummy test included in the package
mzXML <- dir(system.file(package="MSnbase",dir="extdata"),
             full.name=TRUE,
             pattern="mzXML$")
basename(mzXML)
```

```
## [1] "dummyiTRAQ.mzXML"
```

```
## reads the raw data into and MSnExp instance
raw <- readMSData(mzXML, verbose = FALSE)
raw
```

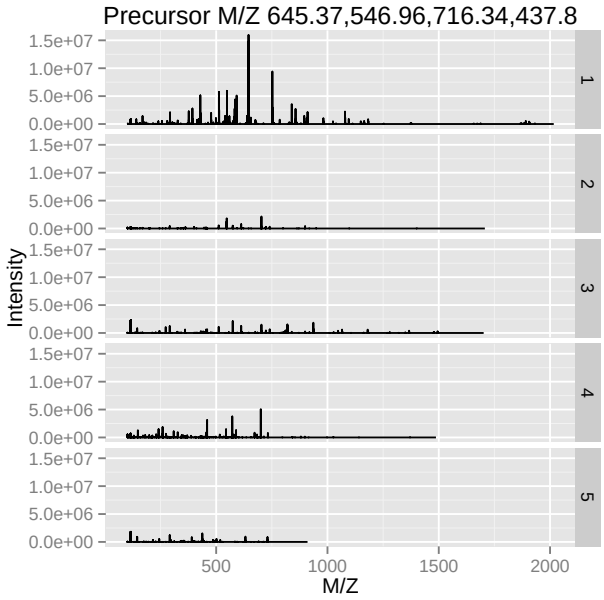
```
## Object of class "MSnExp"
##  Object size in memory: 0.2 Mb
## - - - Spectra data - - -
##  MS level(s): 2
##  Number of MS1 acquisitions: 1
##  Number of MSn scans: 5
##  Number of precursor ions: 5
##  4 unique MZs
##  Precursor MZ's: 437.8 - 716.34
##  MSn M/Z range: 100 2016.66
##  MSn retention times: 25:1 - 25:2 minutes
## - - - Processing information - - -
## Data loaded: Mon Mar 23 16:12:15 2015
##  MSnbase version: 1.15.13
## - - - Meta data  - - -
## phenoData
##   rowNames: 1
##   varLabels: sampleNames
##   varMetadata: labelDescription
## Loaded from:
##   dummyiTRAQ.mzXML
## protocolData: none
## featureData
##   featureNames: X1.1 X2.1 ... X5.1 (5 total)
```

```
##   fvarLabels: spectrum
##   fvarMetadata: labelDescription
## experimentData: use 'experimentData(object)'
```

```
## Extract a single spectrum
raw[[3]]
```

```
## Object of class "Spectrum2"
##  Precursor: 645.3741
##  Retention time: 25:2
##  Charge: 2
##  MSn level: 2
##  Peaks count: 2125
##  Total ion count: 150838188
```

```
plot(raw, full=TRUE)
```




```
plot(raw[[3]], full=TRUE, reporters=iTRAQ4)
```

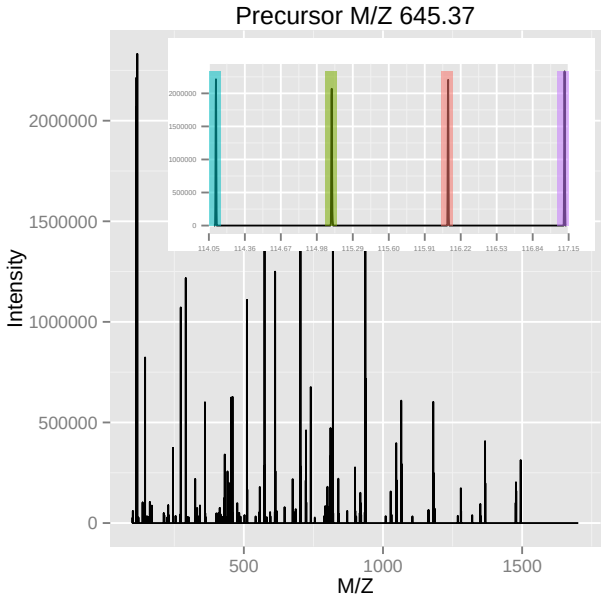


Figure 1: The `plot` method can be used on experiments, i.e. spectrum collections (top), or individual spectra (bottom).

3.1 mgf read/write support

Read and write support for data in the mgf[16] and mzTab[17] formats are available via the readMgfData/writeMgfData and readMzTabData/writeMzTabData functions, respectively. An example for the latter is shown in the next section.

4 Quantitative proteomics

As an running example throughout this document, we will use a TMT 6-plex data set, PXD000001 to illustrate quantitative data processing. The code chunk below first downloads this data file from the ProteomeXchange server using the *rpx* package.

4.1 The mzTab format

The first code chunk downloads the mzTab data from the ProteomeXchange repository [9].

```
## Experiment information
library("rpx")
px1 <- PXDataset("PXD000001")
px1
```

```
## Object of class "PXDataset"
##  Id: PXD000001 with 10 files
##  [1] 'F063721.dat' ... [10] 'erwinia_carotovora.fasta'
##  Use 'pxfiles(.)' to see all files.
```

```
pxfiles(px1)
```

```
##  [1] "F063721.dat"
##  [2] "F063721.dat-mztab.txt"
##  [3] "PRIDE_Exp_Complete_Ac_22134.xml.gz"
##  [4] "PRIDE_Exp_mzData_Ac_22134.xml.gz"
##  [5] "PXD000001_mztab.txt"
##  [6] "TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01-20141210.mzML"
##  [7] "TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01-20141210.mzXML"
##  [8] "TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01.mzXML"
##  [9] "TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01.raw"
## [10] "erwinia_carotovora.fasta"
```

```
## Downloading the mzTab data
mztab <- pxget(px1, "PXD000001_mztab.txt")
```

```
## Downloading 1 file
## PXD000001_mztab.txt already present.
```

```
mztab
```

```
## [1] "PXD000001_mztab.txt"
```

The code below loads the mzTab file into *R* and generates an MSnSet instance, removes missing values and calculates protein intensities by summing the peptide quantitation data. Figure 2 illustrates the intensities for 5 proteins.

```
## Load mzTab peptide data
qnt <- readMzTabData(mztab, what = "PEP")
```

[16] http://www.matrixscience.com/help/data_file_help.html#GEN
[17] https://code.google.com/p/mztab/

```
## Warning in readMzTabData(mztab, what = "PEP"): Support for mzTab version 0.9 only. Support will
be added soon.

## Detected a metadata section
## Detected a peptide section

## Warning: NAs introduced by coercion

sampleNames(qnt) <- reporterNames(TMT6)
head(exprs(qnt))

##   TMT6.126 TMT6.127 TMT6.128 TMT6.129 TMT6.130 TMT6.131
## 1       NA       NA       NA       NA       NA       NA
## 2 10630132 11238708 12424917 10997763  9928972 10398534
## 3       NA       NA       NA       NA       NA       NA
## 4       NA       NA       NA       NA       NA       NA
## 5 11105690 12403253 13160903 12229367 11061660 10131218
## 6  1183431  1322371  1599088  1243715  1306602  1159064

## remove missing values
qnt <- filterNA(qnt)
processingData(qnt)

## - - - Processing information - - -
## mzTab read: Mon Mar 23 16:12:22 2015
## Subset [2351,6][1504,6] Mon Mar 23 16:12:22 2015
## Removed features with more than 0 NAs: Mon Mar 23 16:12:22 2015
## Dropped featureData's levels Mon Mar 23 16:12:22 2015
##  MSnbase version: 1.15.13

## combine into proteins
## - using the 'accession' feature meta data
## - sum the peptide intensities
protqnt <- combineFeatures(qnt,
                           groupBy = fData(qnt)$accession,
                           fun = sum)

## Combined 1504 features into 399 using user-defined function
```

```
qntS <- normalise(qnt, "sum")
qntV <- normalise(qntS, "vsn")
qntV2 <- normalise(qnt, "vsn")

acc <- c("P00489", "P00924",
         "P02769", "P62894",
         "ECA")

idx <- sapply(acc, grep, fData(qnt)$accession)
idx2 <- sapply(idx, head, 3)
small <- qntS[unlist(idx2), ]

idx3 <- sapply(idx, head, 10)
medium <- qntV[unlist(idx3), ]

m <- exprs(medium)
colnames(m) <- c("126", "127", "128",
                 "129", "130", "131")
rownames(m) <- fData(medium)$accession
```

```
cls <- brewer.pal(5, "Set1")
matplot(t(tail(exprs(protqnt), n = 5)), type = "b",
        lty = 1, col = cls,
        ylab = "Protein intensity (summed peptides)",
        xlab = "TMT reporters")
legend("topright", tail(featureNames(protqnt), n=5),
       lty = 1, bty = "n", cex = .8, col = cls)
```

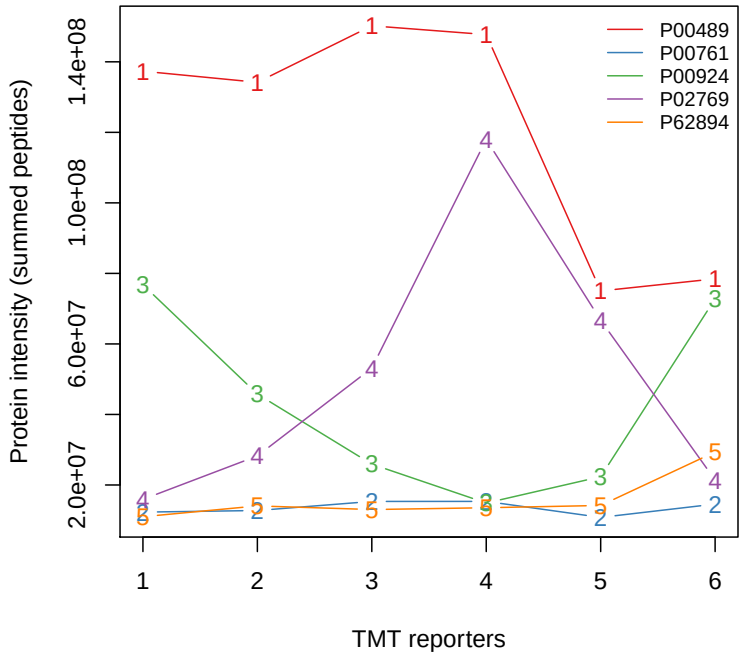


Figure 2: Protein quantitation data.

```
rownames(m)[grep("CYC", rownames(m))] <- "CYT"
rownames(m)[grep("ENO", rownames(m))] <- "ENO"
rownames(m)[grep("ALB", rownames(m))] <- "BSA"
rownames(m)[grep("PYGM", rownames(m))] <- "PHO"
rownames(m)[grep("ECA", rownames(m))] <- "Background"

cls <- c(brewer.pal(length(unique(rownames(m)))-1, "Set1"),
         "grey")
names(cls) <- unique(rownames(m))
wbcol <- colorRampPalette(c("white", "darkblue"))(256)
```

```
heatmap(m, col = wbcol, RowSideColors=cls[rownames(m)])
```

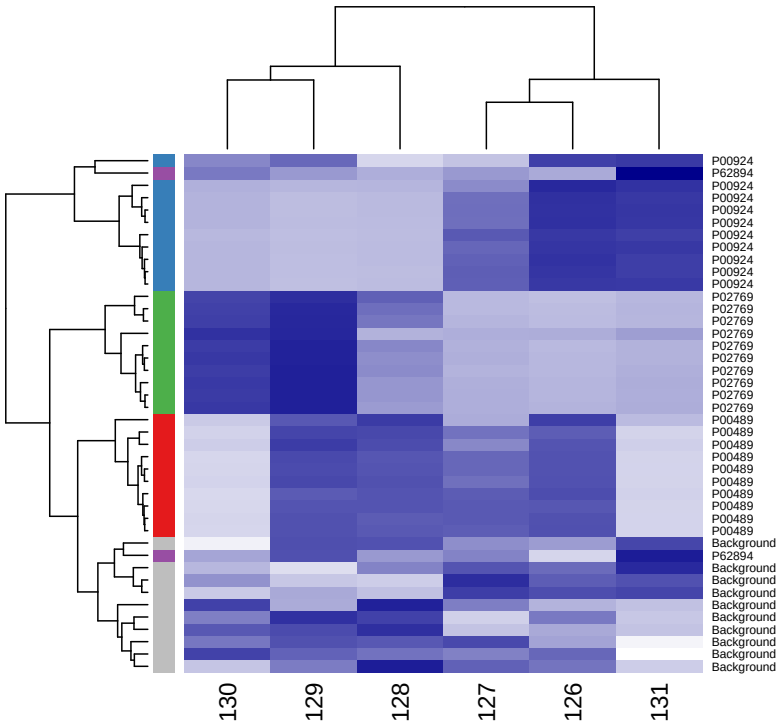


Figure 3: A heatmap.

```
dfr <- data.frame(exprs(small),
                  Protein = as.character(fData(small)$accession),
                  Feature = featureNames(small),
                  stringsAsFactors = FALSE)
colnames(dfr) <- c("126", "127", "128", "129", "130", "131",
                   "Protein", "Feature")
dfr$Protein[dfr$Protein == "sp|P00924|ENO1_YEAST"] <- "ENO"
dfr$Protein[dfr$Protein == "sp|P62894|CYC_BOVIN"]  <- "CYT"
dfr$Protein[dfr$Protein == "sp|P02769|ALBU_BOVIN"] <- "BSA"
dfr$Protein[dfr$Protein == "sp|P00489|PYGM_RABIT"] <- "PHO"
dfr$Protein[grep("ECA", dfr$Protein)] <- "Background"
dfr2 <- melt(dfr)
## Using Protein, Feature as id variables
ggplot(aes(x = variable, y = value, colour = Protein),
       data = dfr2) +
  geom_point() +
  geom_line(aes(group=as.factor(Feature)), alpha = 0.5) +
  facet_grid(. ~ Protein) + theme(legend.position="none") +
  labs(x = "Reporters", y = "Normalised intensity")
```

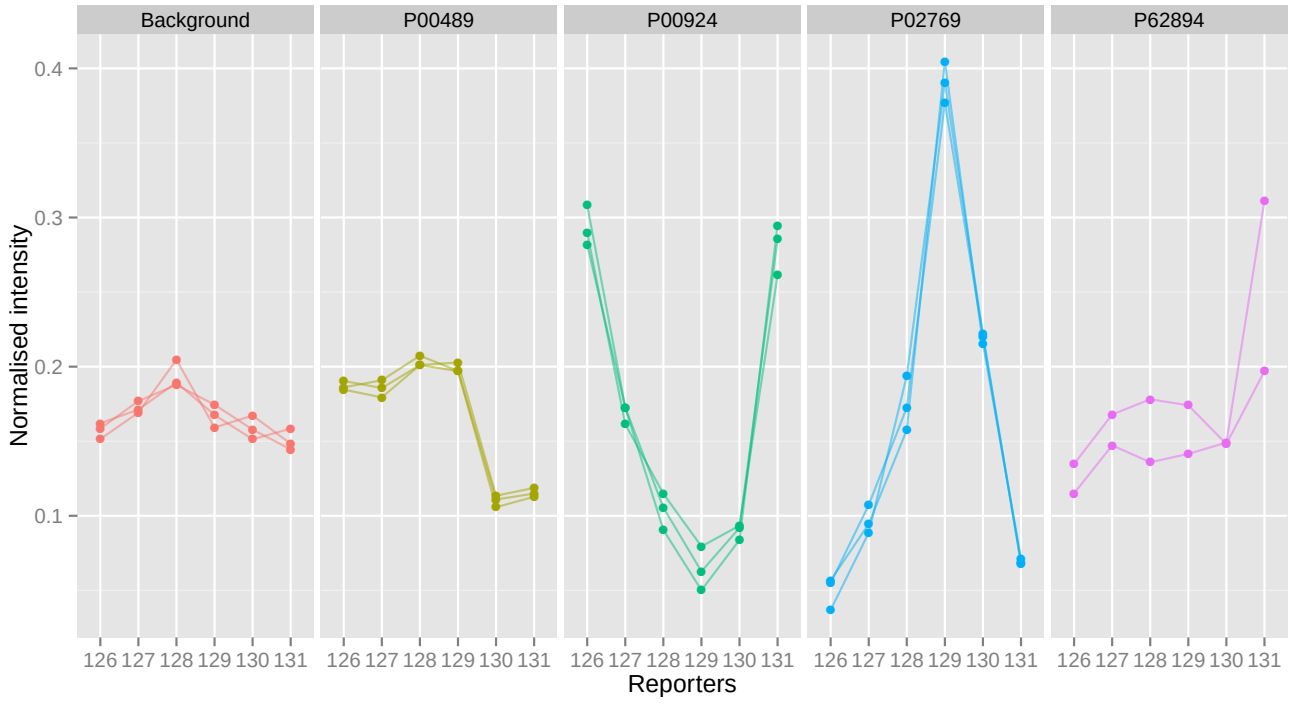


Figure 4: Spikes plot using *ggplot2*.

4.2 Working with raw data

We reuse our dedicated `px1` ProteomeXchange data object to download the raw data (in `mzXML` format) and load it with the `readMSData` from the *MSnbase* package that produces a raw data experiment object of class `MSnExp`. The raw data is then quantified using the `quantify` method specifying the TMT 6-plex isobaric tags and a 7^{th} peak of interest corresponding to the un-dissociated reporter tag peaks (see the `MSnbase-demo` vignette in *MSnbase* for details).

```
mzxml <- pxget(px1, "TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01.mzXML")
```

```
## Downloading 1 file
```

```
rawms <- readMSData(mzxml, centroided = TRUE, verbose = FALSE)
qntms <- quantify(rawms, reporters = TMT7, method = "max")
```

```
## Using default parallel backend:  MulticoreParam
## Original MSnExp and new MSnSet have different number of samples in phenoData.  Dropping original.
## Creating 'MSnSet' object
```

```
qntms
```

```
## MSnSet (storageMode: lockedEnvironment)
## assayData: 6103 features, 7 samples
##   element names: exprs
## protocolData: none
## phenoData
##   sampleNames: TMT7.126 TMT7.127 ... TMT7.230 (7
##     total)
##   varLabels: mz reporters
##   varMetadata: labelDescription
## featureData
##   featureNames: X1000.1 X100.1 ... X999.1 (6103
##     total)
##   fvarLabels: spectrum file ... collision.energy (12
##     total)
##   fvarMetadata: labelDescription
## experimentData: use 'experimentData(object)'
## Annotation: No annotation
## - - - Processing information - - -
## Data loaded: Mon Mar 23 14:05:03 2015
## TMT7 quantification by max: Mon Mar 23 14:08:21 2015
##  MSnbase version: 1.15.13
```

Identification data in the `mzIdentML` format can be added to `MSnExp` or `MSnSet` instances with the `addIdentificationData` function. See the function documentation for examples.

```
d <- data.frame(Signal = rowSums(exprs(qntms)[, 1:6]),
                Incomplete = exprs(qntms)[, 7])
d <- log(d)
cls <- rep("#00000050", nrow(qnt))
pch <- rep(1, nrow(qnt))
cls[grep("P02769", fData(qnt)$accession)] <- "gold4" ## BSA
cls[grep("P00924", fData(qnt)$accession)] <- "dodgerblue" ## ENO
cls[grep("P62894", fData(qnt)$accession)] <- "springgreen4" ## CYT
cls[grep("P00489", fData(qnt)$accession)] <- "darkorchid2" ## PHO
pch[grep("P02769", fData(qnt)$accession)] <- 19
pch[grep("P00924", fData(qnt)$accession)] <- 19
pch[grep("P62894", fData(qnt)$accession)] <- 19
pch[grep("P00489", fData(qnt)$accession)] <- 19
```

```
mzp <- plotMzDelta(rawms, reporters = TMT6, verbose = FALSE) + ggtitle("")
```

```
mzp
```

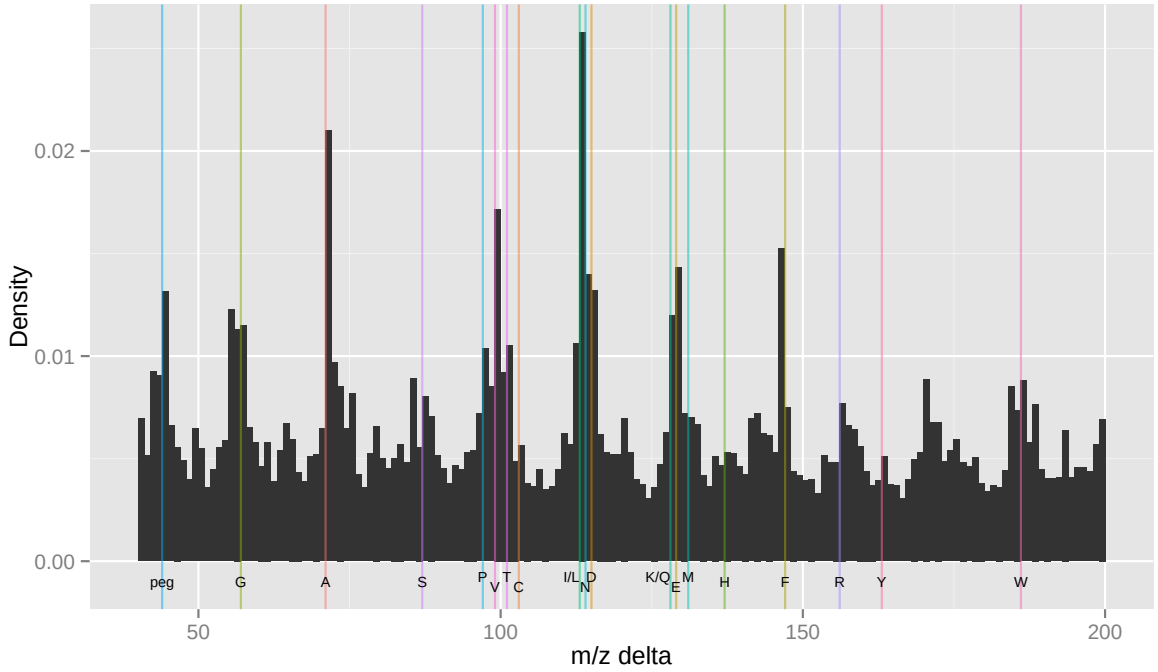


Figure 5: A m/z delta plot.

```
plot(Signal ~ Incomplete, data = d,
     xlab = expression(Incomplete~dissociation),
     ylab = expression(Sum~of~reporters~intensities),
     pch = 19,
     col = "#4582B380")
grid()
abline(0, 1, lty = "dotted")
abline(lm(Signal ~ Incomplete, data = d), col = "darkblue")
```

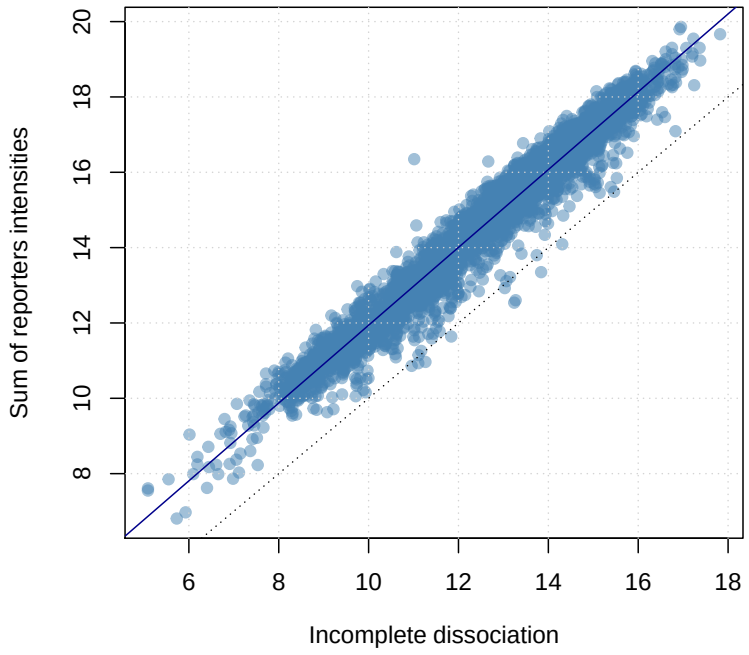


Figure 6: Incomplete dissociation.

```
MAplot(qnt[, c(4, 2)], cex = .9, col = cls, pch = pch, show.statistics = FALSE)
```

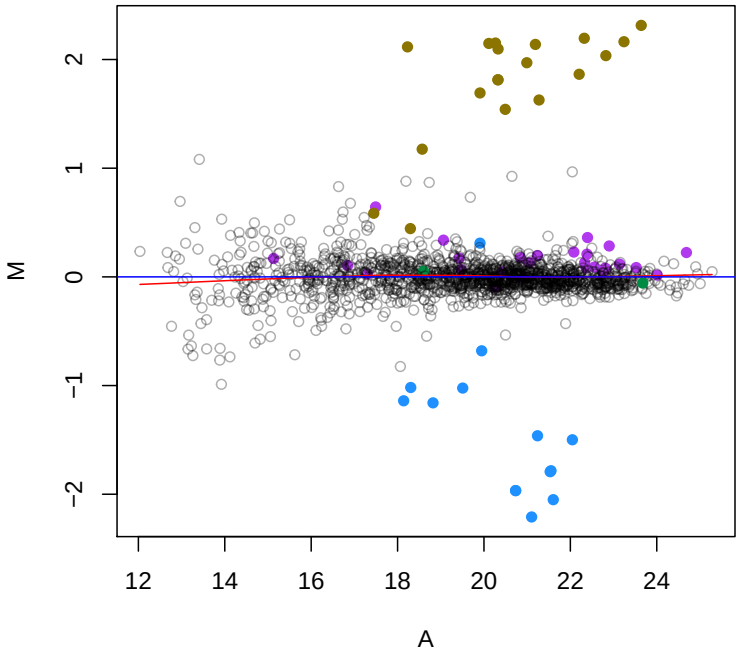


Figure 7: MAplot on an `MSnSet` instance.

4.3 The MALDIquant package

This section illustrates some of *MALDIquant*'s data processing capabilities [10]. The code is taken from the `processing-peaks.R` script downloaded from the package homepage[18].

Loading the data

```
## load packages
library("MALDIquant")
library("MALDIquantForeign")
## getting test data
datapath <-
  file.path(system.file("Examples",
                        package = "readBrukerFlexData"),
            "2010_05_19_Gibb_C8_A1")
dir(datapath)
```

```
## [1] "0_A1" "0_A2"
```

```
sA1 <- importBrukerFlex(datapath, verbose=FALSE)
# in the following we use only the first spectrum
s <- sA1[[1]]

summary(mass(s))
```

```
##    Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##   999.9  2373.0  4331.0  4721.0  6874.0 10000.0
```

```
summary(intensity(s))
```

```
##    Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##       4     180    1562    2841    4656   32590
```

```
head(as.matrix(s))
```

```
##           mass intensity
## [1,]  999.9388     11278
## [2,] 1000.1316     11350
## [3,] 1000.3244     10879
## [4,] 1000.5173     10684
## [5,] 1000.7101     10740
## [6,] 1000.9030     10947
```

Preprocessing

```
## sqrt transform (for variance stabilization)
s2 <- transformIntensity(s, method="sqrt")
s2
```

```
## S4 class type            : MassSpectrum
## Number of m/z values     : 22431
## Range of m/z values      : 999.939 - 10001.925
## Range of intensity values: 2e+00 - 1.805e+02
## Memory usage             : 359.875 KiB
## Name                     : 2010_05_19_Gibb_C8_A1.A1
```

[18] http://strimmerlab.org/software/maldiquant/

```
plot(s)
```

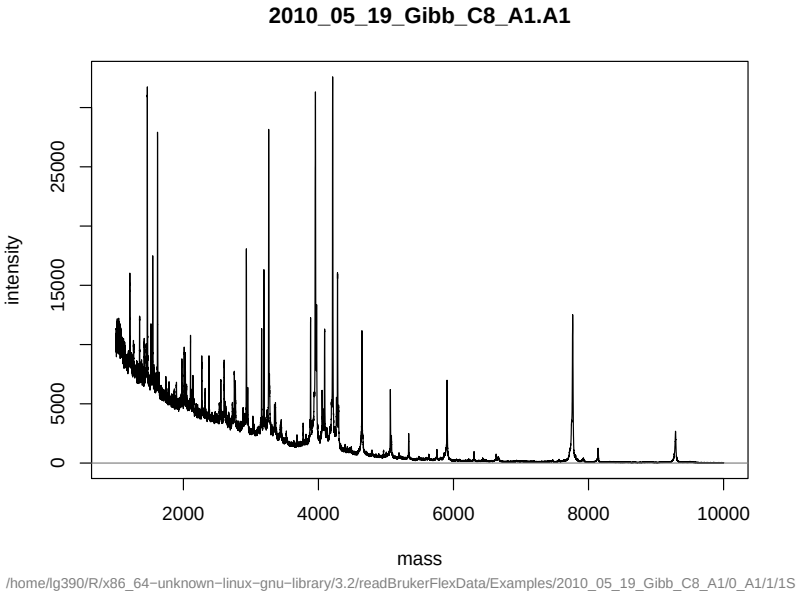


Figure 8: Spectrum plotting in *MALDIquant*.

```
## File                    : /home/lg390/R/x86_64-unknown-linux-gnu-library/3.2/readBrukerFlexData/Exampl
```

```
## smoothing - 5 point moving average
s3 <- smoothIntensity(s2, method="MovingAverage", halfWindowSize=2)
s3
```

```
## S4 class type           : MassSpectrum
## Number of m/z values    : 22431
## Range of m/z values     : 999.939 - 10001.925
## Range of intensity values: 3.606e+00 - 1.792e+02
## Memory usage            : 359.875 KiB
## Name                    : 2010_05_19_Gibb_C8_A1.A1
## File                    : /home/lg390/R/x86_64-unknown-linux-gnu-library/3.2/readBrukerFlexData/Exampl
```

```
## baseline subtraction
s4 <- removeBaseline(s3, method="SNIP")
s4
```

```
## S4 class type           : MassSpectrum
## Number of m/z values    : 22431
## Range of m/z values     : 999.939 - 10001.925
## Range of intensity values: 0e+00 - 1.404e+02
## Memory usage            : 359.875 KiB
## Name                    : 2010_05_19_Gibb_C8_A1.A1
## File                    : /home/lg390/R/x86_64-unknown-linux-gnu-library/3.2/readBrukerFlexData/Exampl
```

Peak picking

```
## peak picking
p <- detectPeaks(s4)
```

```
length(p) # 181
```

```
## [1] 186
```

```
peak.data <- as.matrix(p) # extract peak information
```

```
par(mfrow=c(2,3))
xl <- range(mass(s))
# use same xlim on all plots for better comparison
plot(s, sub="", main="1: raw", xlim=xl)
plot(s2, sub="", main="2: variance stabilisation", xlim=xl)
plot(s3, sub="", main="3: smoothing", xlim=xl)
plot(s4, sub="", main="4: base line correction", xlim=xl)
plot(s4, sub="", main="5: peak detection", xlim=xl)
points(p)
top20 <- intensity(p) %in% sort(intensity(p), decreasing=TRUE)[1:20]
labelPeaks(p, index=top20, underline=TRUE)
plot(p, sub="", main="6: peak plot", xlim=xl)
labelPeaks(p, index=top20, underline=TRUE)
```

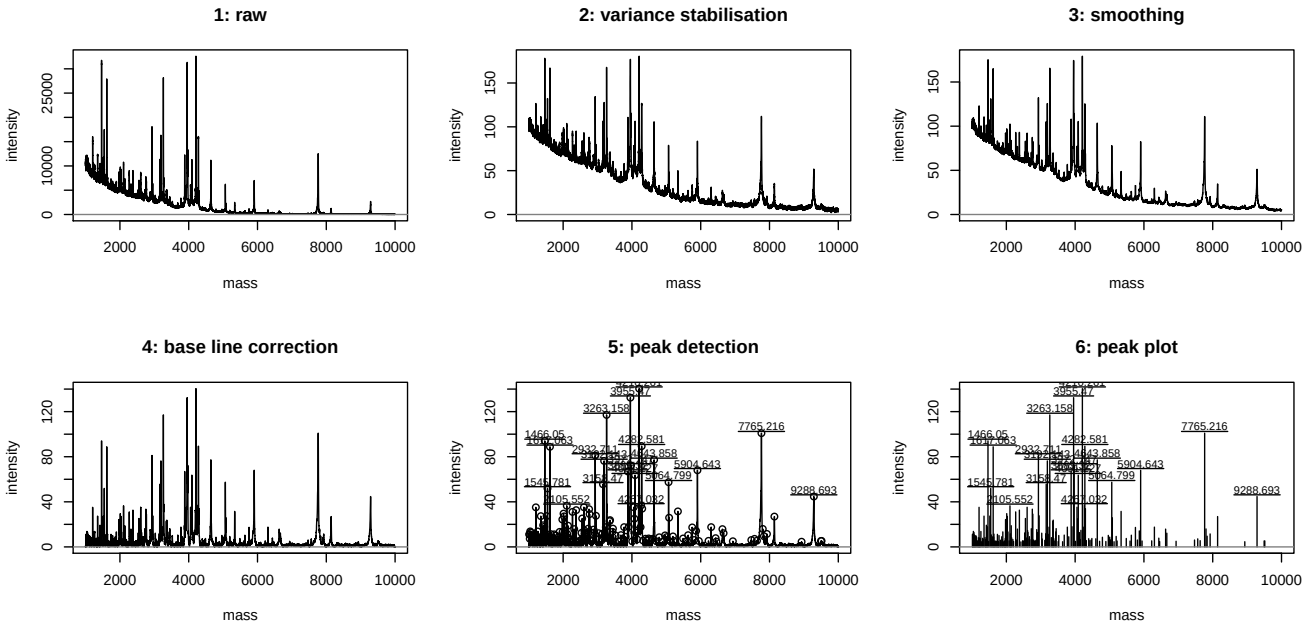


Figure 9: Spectrum plotting in *MALDIquant*.

4.4 Working with peptide sequences

```
library(IPPD)
library(BRAIN)
atoms <- getAtomsFromSeq("SIVPSGASTGVHEALEMR")
unlist(atoms)
```

```
##   C   H   N   O   S
##  77 129  23  27   1
```

```
library(Rdisop)
pepmol <- getMolecule(paste0(names(atoms),
                             unlist(atoms),
                             collapse = ""))
pepmol
```

```
## $formula
## [1] "C77H129N23O27S"
##
## $score
## [1] 1
##
## $exactmass
## [1] 1839.915
##
## $charge
## [1] 0
##
## $parity
## [1] "e"
##
## $valid
## [1] "Valid"
##
## $DBE
## [1] 25
##
## $isotopes
## $isotopes[[1]]
##              [,1]         [,2]         [,3]         [,4]
## [1,] 1839.9148973 1840.9177412 1841.9196777 1.842921e+03
## [2,]    0.3427348    0.3353456    0.1960976 8.474135e-02
##              [,5]         [,6]         [,7]         [,8]
## [1,] 1.843923e+03 1.844925e+03 1.845927e+03 1.846928e+03
## [2,] 2.952833e-02 8.691735e-03 2.226358e-03 5.066488e-04
##              [,9]        [,10]
## [1,] 1.847930e+03 1.848932e+03
## [2,] 1.040196e-04 1.949686e-05
```

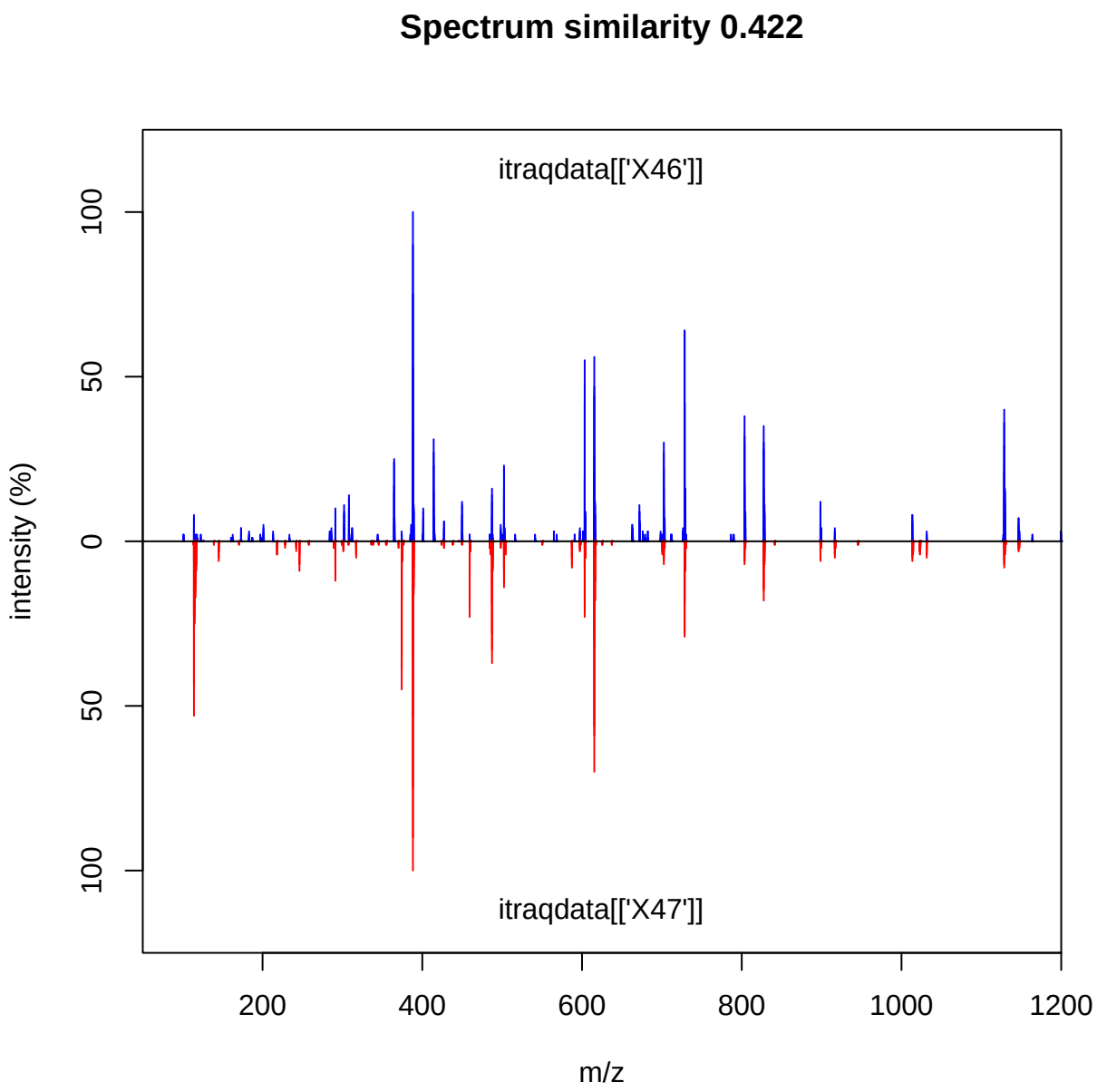
```
##
library(OrgMassSpecR)
data(itraqdata)

simplottest <-
  itraqdata[featureNames(itraqdata) %in% paste0("X", 46:47)]
sim <- SpectrumSimilarity(as(simplottest[[1]], "data.frame"),
```

```
                        as(simplottest[[2]], "data.frame"),
                        top.lab = "itraqdata[['X46']]",
                        bottom.lab = "itraqdata[['X47']]",
                        b = 25)
```

```
##           mz intensity.top intensity.bottom
## 1   114.1091             0               44
## 2   114.1109             0               53
## 3   114.1127             0               43
## 4   115.1085             0               25
## 5   364.7215            25                0
## 6   374.2082             0               39
## 7   374.2191             0               45
## 8   374.2301             0               35
## 9   388.2442             0               35
## 10  388.2558             0               75
## 11  388.2673             0              100
## 12  388.2789             0               90
## 13  388.2904            35               53
## 14  388.2904           100               53
## 15  388.2904            90               53
## 16  388.2904            53               53
## 17  388.2904            75               53
## 18  414.2582            31                0
## 19  414.2709            27                0
## 20  487.2887             0               33
## 21  487.3050             0               37
## 22  487.3213             0               28
## 23  603.3339            42                0
## 24  603.3563            55                0
## 25  603.3787            48                0
## 26  603.4011            27                0
## 27  615.3124             0               28
## 28  615.3354             0               56
## 29  615.3585             0               70
## 30  615.3816             0               59
## 31  615.4047            26               32
## 32  615.4047            44               32
## 33  615.4047            56               32
##  [ reached getOption("max.print") -- omitted 17 rows ]
```

```
title(main = paste("Spectrum similarity", round(sim, 3)))
```




```
MonoisotopicMass(formula = list(C = 2, O = 1, H=6))
```

```
## [1] 46.04186
```

```
molecule <- getMolecule("C2H5OH")
molecule$exactmass
```

```
## [1] 46.04186
```

```
## x11()
## plot(t(.pepmolfisotopes[[1]]), type = "h")

## x <- IsotopicDistribution(formula = list(C = 2, O = 1, H=6))
## t(moleculefisotopes[[1]])
```

```
## par(mfrow = c(2,1))
## plot(t(moleculeIsotopes[[1]]), type = "h")
## plot(x[, c(1,3)], type = "h")

## data(myo500)
## masses <- c(147.053, 148.056)
## intensities <- c(93, 5.8)
## molecules <- decomposeIsotopes(masses, intensities)

## experimental eno peptides
exppep <-
  as.character(fData(qnt[grep("ENO", fData(qnt)[, 2]), ])[, 1]) ## 13
minlength <- min(nchar(exppep))


if (!file.exists("P00924.fasta"))
    eno <- download.file("http://www.uniprot.org/uniprot/P00924.fasta",
                         destfile = "P00924.fasta")
eno <- paste(readLines("P00924.fasta")[-1], collapse = "")
enopep <- Digest(eno, missed = 1)
nrow(enopep) ## 103
```

```
## [1] 103
```

```
sum(nchar(enopep$peptide) >= minlength) ## 68
```

```
## [1] 0
```

```
pepcnt <- enopep[enopep[, 1] %in% exppep, ]
nrow(pepcnt) ## 13
```

```
## [1] 0
```

The following code chunks demonstrate how to use the *cleaver* package for in-silico cleavage of polypeptides, e.g. cleaving of *Gastric juice peptide 1 (P01358)* using *Trypsin*:

```
library(cleaver)
cleave("LAAGKVEDSD", enzym = "trypsin")
```

```
## $LAAGKVEDSD
## [1] "LAAGK" "VEDSD"
```

Sometimes cleavage is not perfect and the enzym miss some cleavage positions:

```
## miss one cleavage position
cleave("LAAGKVEDSD", enzym = "trypsin", missedCleavages = 1)
```

```
## $LAAGKVEDSD
## [1] "LAAGKVEDSD"
```

```
## miss zero or one cleavage positions
cleave("LAAGKVEDSD", enzym = "trypsin", missedCleavages = 0:1)
```

```
## $LAAGKVEDSD
## [1] "LAAGK"      "VEDSD"      "LAAGKVEDSD"
```

Example code to generate an Texshade image to be included directly in a Latex document or *R* vignette is presented below. The *R* code generates a Texshade environment and the annotated sequence display code that is written to a TEX file that can itself be included into a LATEX of Sweave document.

```
seq1file <- "seq1.tex"
```

```
cat("\\begin{texshade}{Figures/P00924.fasta}
     \\setsize{numbering}{footnotesize}
     \\setsize{residues}{footnotesize}
     \\residuesperline*{70}
     \\shadingmode{functional}
     \\hideconsensus
     \\vsepspace{1mm}
     \\hidenames
     \\noblockskip\n", file = seq1file)
tmp <- sapply(1:nrow(pepcnt), function(i) {
  col <- ifelse((i %% 2) == 0, "Blue", "RoyalBlue")
  cat("\\shaderegion{1}{", pepcnt$start[i], "..", pepcnt$stop[i], "}{White}{", col, "}\n",
      file = seq1file, append = TRUE)
})
cat("\\end{texshade}
    \\caption{Visualising observed peptides for the Yeast enolase protein. Peptides are shaded in blue and black.
              The last peptide is a mis-cleavage and overlaps with \\texttt{IEEELGDNAVFAGENFHHGDK}.}
    \\label{fig:seq}
  \\end{center}
\\end{figure}\n\n",
    file = seq1file, append = TRUE)
```

^{15}N incorporation

```
## 15N incorporation rates from 0, 0.1, ..., 0.9, 0.95, 1
increate <- c(seq(0, 0.9, 0.1), 0.95, 1)
inc <- lapply(increate, function(inc)
             IsotopicDistributionN("YEVQGEVFTKPQLWP", inc))
par(mfrow = c(4,3))
for (i in 1:length(inc))
  plot(inc[[i]][, c(1, 3)], xlim = c(1823, 1848), type = "h",
       main = paste0("15N incorporation at ", increate[i]*100, "%"))
```

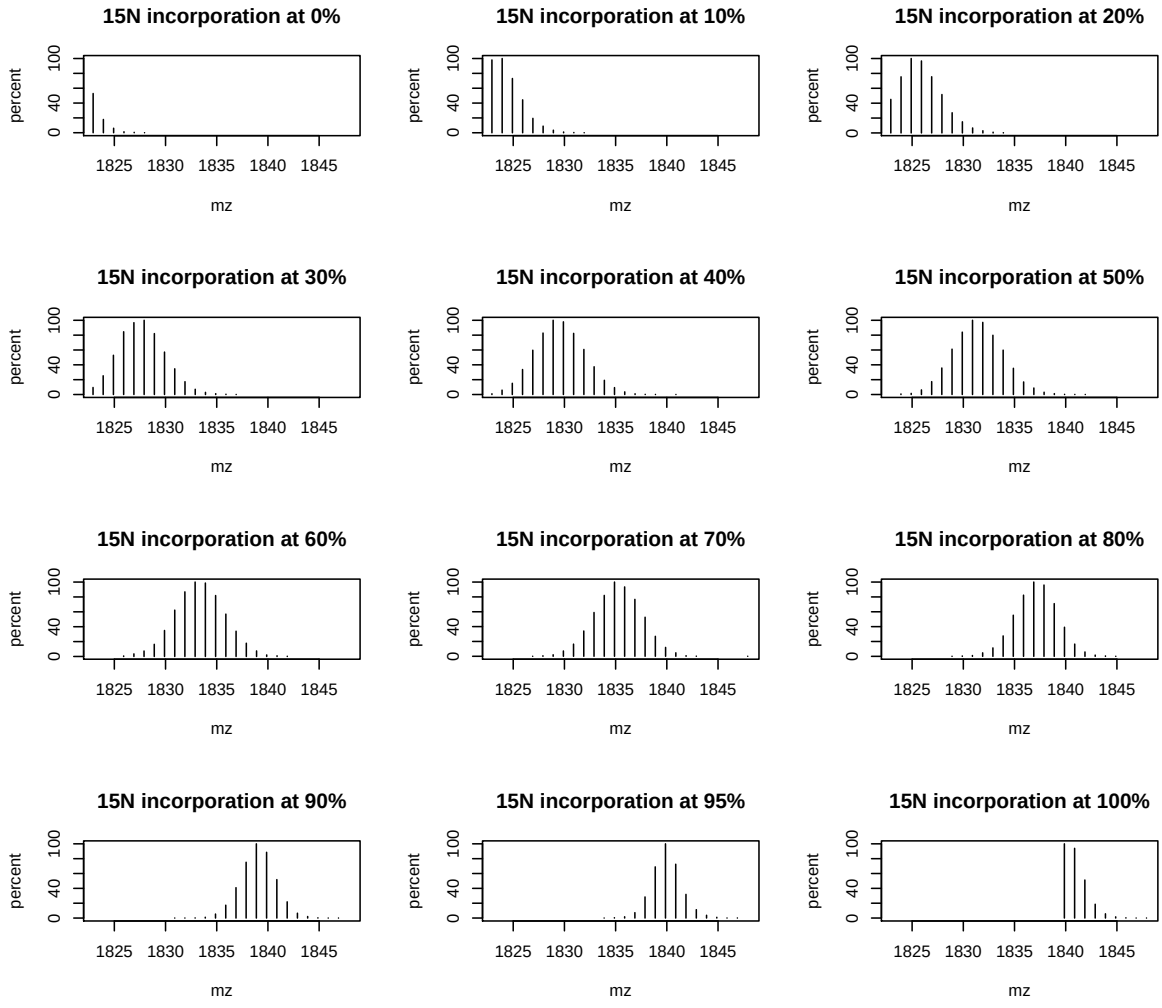


Figure 10: Isotopic envelope for the YEVQGEVFTKPQLWP peptide at different ^{15}N incorporation rates.

4.5 The isobar package

The *isobar* package [11] provides methods for the statistical analysis of isobarically tagged MS^2 experiments.

```
library(isobar)

## Prepare the PXD000001 data for isobar analysis
.ions <- exprs(qnt)
.mass <- matrix(mz(TMT6), nrow(qnt), byrow=TRUE, ncol = 6)
colnames(.ions) <- colnames(.mass) <-
  reporterTagNames(new("TMT6plexSpectra"))
rownames(.ions) <- rownames(.mass) <-
  paste(fData(qnt)$accession, fData(qnt)$sequence, sep = ".")
pgtbl <- data.frame(spectrum = rownames(.ions),
                    peptide = fData(qnt)$sequence,
                    modif = ":",
                    start.pos = 1,
                    protein = fData(qnt)$accession,
                    accession = fData(qnt)$accession)
x <- new("TMT6plexSpectra", pgtbl, .ions, .mass)
```

```
##  data.frame columns OK
## Creating ProteinGroup ...  done
```

```
featureData(x)$proteins <- as.character(fData(qnt)$accession)

x <- correctIsotopeImpurities(x) ## using identity matrix here
```

```
## LOG: isotopeImpurities.corrected:  TRUE
```

```
x <- normalize(x, per.file = FALSE)
```

```
## LOG: is.normalized:  TRUE
## LOG: normalization.multiplicative.factor channel 126:  0.8846
## LOG: normalization.multiplicative.factor channel 127:  0.9244
## LOG: normalization.multiplicative.factor channel 128:  1
## LOG: normalization.multiplicative.factor channel 129:  0.9421
## LOG: normalization.multiplicative.factor channel 130:  0.8593
## LOG: normalization.multiplicative.factor channel 131:  0.889
```

```
## spikes
spks <- c(protein.g(proteinGroup(x), "P00489"),
          protein.g(proteinGroup(x), "P00924"),
          protein.g(proteinGroup(x), "P02769"),
          protein.g(proteinGroup(x), "P62894"))

cls2 <- rep("#00000040", nrow(x))
pch2 <- rep(1, nrow(x))
cls2[grep("P02769", featureNames(x))] <- "gold4" ## BSA
cls2[grep("P00924", featureNames(x))] <- "dodgerblue" ## ENO
cls2[grep("P62894", featureNames(x))] <- "springgreen4" ## CYT
cls2[grep("P00489", featureNames(x))] <- "darkorchid2" ## PHO
pch2[grep("P02769", featureNames(x))] <- 19
pch2[grep("P00924", featureNames(x))] <- 19
pch2[grep("P62894", featureNames(x))] <- 19
pch2[grep("P00489", featureNames(x))] <- 19
```

```
nm <- NoiseModel(x)

## [1] 7.306091e-02 1.140614e+04 3.489853e+00

ib.background <- subsetIBSpectra(x, protein=spks,
                                 direction = "exclude")

## Creating ProteinGroup ...  done

nm.background <- NoiseModel(ib.background)

## [1] 0.01425222 3.49812516 0.89685036

ib.spks <- subsetIBSpectra(x, protein = spks,
                           direction="include",
                           specificity="reporter-specific")

## Creating ProteinGroup ...  done

nm.spks <- NoiseModel(ib.spks, one.to.one=FALSE, pool=TRUE)

## 4 proteins with more than 10 spectra, taking top 50.
## [1] 0.0000000001 6.1927071539 0.6721054619

ratios <- 10^estimateRatio(x, nm,
                           channel1="127", channel2="129",
                           protein = spks,
                           combine = FALSE)[, "lratio"]


res <- estimateRatio(x, nm,
                     channel1="127", channel2="129",
                     protein = unique(fData(x)$proteins),
                     combine = FALSE,
                     sign.level = 0.01)[, c(1, 2, 6, 8)]
res <- as.data.frame(res)
res$lratio <- -(res$lratio)

cls3 <- rep("#00000050", nrow(res))
pch3 <- rep(1, nrow(res))
cls3[grep("P02769", rownames(res))] <- "gold4" ## BSA
cls3[grep("P00924", rownames(res))] <- "dodgerblue" ## ENO
cls3[grep("P62894", rownames(res))] <- "springgreen4" ## CYT
cls3[grep("P00489", rownames(res))] <- "darkorchid2" ## PHO
pch3[grep("P02769", rownames(res))] <- 19
pch3[grep("P00924", rownames(res))] <- 19
pch3[grep("P62894", rownames(res))] <- 19
pch3[grep("P00489", rownames(res))] <- 19

rat.exp <- c(PHO = 2/2,
             ENO = 5/1,
             BSA = 2.5/10,
             CYT = 1/1)
```

```
maplot(x,
       noise.model = c(nm.background, nm.spks, nm),
       channel1="127", channel2="129",
       pch = 19, col = cls2,
       main = "Spectra MA plot")
abline(h = 1, lty = "dashed", col = "grey")
legend("topright",
       c("BSA", "ENO", "CYT", "PHO"),
       pch = 19, col = c("gold4", "dodgerblue",
                  "springgreen4", "darkorchid2"),
       bty = "n", cex = .7)
```

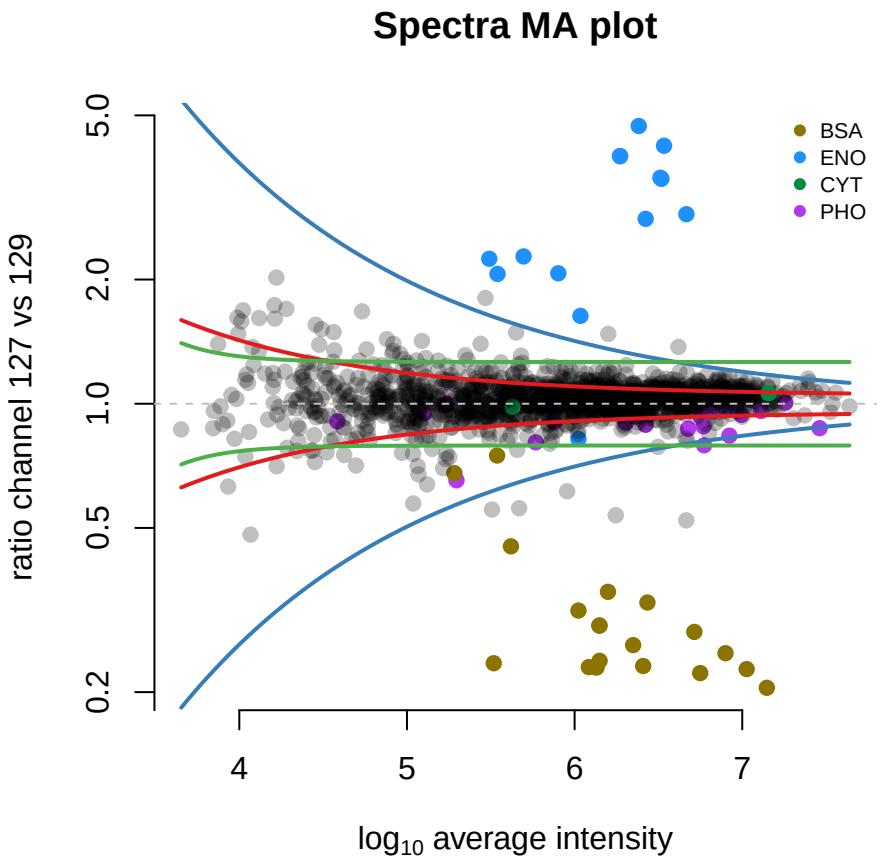


Figure 11: Result from the *isobar* pipeline.

4.6 The synapter package

The *synapter* [12] package comes with a detailed vignette that describes how to prepare the MS^E data and then process it in *R*. Several interfaces are available provided the user with maximum control, easy batch processing capabilities or a graphical user interface. The conversion into `MSnSet` instances and filter and combination thereof as well as statistical analysis are also described.

```
## open the synapter vignette
library("synapter")
synapterGuide()
```

5 MS^2 spectra identification

At the moment two packages allow the user to run peptide identifications from within R. Each of the packages interface to an external peptide database search tool and have more or less the same workflow, though their syntax differs:

1. Prepare the input data.
2. Run the search.
3. Import the search results and extract the peptides and proteins

5.1 X! Tandem

Following Bioconductor 2.12 the *rTANDEM* package provides the means to run the popular X! Tandem software [13].

Using example code/data from the *rTANDEM* vignette/package, the following is an example of a typical workflow

5.1.1 Preparation of the input data

```
library(rTANDEM)
taxonomy <- rTTaxo(taxon = "yeast",
                   format = "peptide",
                   URL = system.file(
                     "extdata/fasta/scd.fasta.pro",
                     package="rTANDEM"))
param <- rTParam()
param <- setParamValue(param,
                       'protein', 'taxon',
                       value="yeast")
param <- setParamValue(param, 'list path',
                       'taxonomy information', taxonomy)
param <- setParamValue(param,
                       'list path', 'default parameters',
                       value = system.file(
                         "extdata/default_input.xml",
                         package="rTANDEM"))
param <- setParamValue(param, 'spectrum', 'path',
                       value = system.file(
                         "extdata/test_spectra.mgf",
                         package="rTANDEM"))
param <- setParamValue(param, 'output', 'xsl path',
                       value = system.file(
                         "extdata/tandem-input-style.xsl",
```

```
                          package="rTANDEM"))
param <- setParamValue(param, 'output', 'path',
                       value = paste(getwd(),
                         "output.xml", sep="/"))
```

5.1.2 Performing the search

The analysis is run using the `tandem` function (see also the `rtandem` function), which returns the results data file path (only the file name is displayed below).

```
resultPath <- tandem(param)
```

```
## Loading spectra
##  (mgf). loaded.
## Spectra matching criteria = 242
## Starting threads . started.
## Computing models:
##  testin
##  sequences modelled = 5 ks
## Model refinement:
##  partial cleavage ..... done.
##  unanticipated cleavage ..... done.
##  modified N-terminus ..... done.
##  finishing refinement ... done.
## Creating report:
##  initial calculations  ..... done.
##  sorting  ..... done.
##  finding repeats ..... done.
##  evaluating results ..... done.
##  calculating expectations ..... done.
##  writing results ..... done.
##
## Valid models = 40
## Unique models = 41
## Estimated false positives = 1 +/- 1
```

```
basename(resultPath)
```

```
## [1] "output.2015_03_23_16_12_30.t.xml"
```

5.1.3 Import and analyse results

```
res <- GetResultsFromXML(resultPath)
## the inferred proteins
proteins <- GetProteins(res,
                        log.expect = -1.3,
                        min.peptides = 2)
proteins[, -(4:5), with = FALSE]
```

```
##      uid expect.value   label description num.peptides
##  1:  576        -27.2 YCR012W     YCR012W            5
##  2: 1811        -14.5 YFR053C     YFR053C            3
##  3: 2301        -12.8 YGR254W     YGR254W            3
##  4:    4        -12.0 YAL005C     YAL005C            3
```

```
##  5: 3517        -12.0 YLL024C     YLL024C            3
##  6: 3328        -10.3 YKL152C     YKL152C            2
##  7: 3386        -10.1 YKL216W     YKL216W            2
##  8: 2281         -7.9 YGR234W     YGR234W            2
##  9: 2568         -7.5 YHR174W     YHR174W            2
## 10: 2044         -7.1 YGL253W     YGL253W            2
```

```
## the identified peptides for YFR053C
peptides <- GetPeptides(protein.uid = 1811,
                        results = res,
                        expect = 0.05)
peptides[, c(1:4, 9, 10:16), with = FALSE]
```

```
##     pep.id prot.uid spectrum.id spectrum.mh expect.value
## 1: 102.1.1     1811         102    942.5147      0.00660
## 2: 250.1.1     1811         250   1212.5610      0.00043
## 3:  60.1.1     1811          60    863.4933      0.00870
##    tandem.score        mh   delta peak.count
## 1:         31.9  942.5370 -0.0220         NA
## 2:         35.0 1212.5531  0.0079         NA
## 3:         21.7  863.4985 -0.0052         NA
##    missed.cleavages start.position end.position
## 1:                0            166          173
## 2:                0            437          447
## 3:                0            309          315
```

More details are provided in the vignette available with (vignette("rTANDEM")), for instance the extraction of degenerated peptides, i.e. peptides found in multiple proteins.

The *shinyTANDEM* package offers a web-based graphical interface to *rTANDEM*.

5.2 MS-GF+

With the release of Bioconductor 3.0 the *MSGFplus* package has provided an interface to MS-GF+ [14, 15]. The package vignette describe in detail the different ways an MS-GF+ analysis can be initiated and only a simple example will be given here:

5.2.1 Preparation of the input data

```
library(MSGFplus)
## Create a parameter object with a set of parameters
param <- msgfPar(database = system.file('extdata',
                                        'milk-proteins.fasta',
                                        package='MSGFplus'),
                 tolerance = '10 ppm',
                 enzyme = 'Trypsin')

## Add parameters after creation
instrument(param) <- 'QExactive'
tda(param) <- TRUE

## Add expected modifications
mods(param)[[1]] <- msgfParModification('Carbamidomethyl',
```

```
                                        composition = 'C2H3N1O1',
                                        residues = 'C',
                                        type = 'fix',
                                        position = 'any')
nMod(param) <- 2    # Number of allowed modifications per peptide

## Get a summary of your parameters
show(param)
```

```
## An msgfPar object
##
## Database:                    /home/lg390/R/x86_64-unknown-linux-gnu-library/3.2/MSGFplus/extdata/milk-pr
## Tolerance:                   10 ppm
## TDA:                         TRUE
## Instrument:                  3: QExactive
## Enzyme:                      1: Trypsin
##
## Modifications:
##
## Number of modifications per peptide: 2
##
## Carbamidomethyl: C2H3N1O1, C, fix, any
```

5.2.2 Performing the search

Initiating the search is done using the `runMSGF` method. As a minimum it takes a parameter object and a list of raw data files and performs the search for each data file in sequence. More specialised operations are also possible such as running it asynchronously, but interested readers should refer to the *MSGFplus* vignette for additional information.

The first time a search is initialised the MS-GF+ code is downloaded, so be sure to have an active internet connection (only applies to the first time a search is run).

```
result <- runMSGF(param, 'path/to/a/rawfile.mzML')
```

5.2.3 Import and analyse results

By default MSGFplus imports the results automatically using *mzID*. If only one file was analysed, the return value is an mzID object; if multiple files are analysed at once the return value is an mzIDCollection object.

If `import=FALSE` the results are not imported and can be accessed at a later time using the *mzID* package (see section 2.2 on page 8).

5.2.4 Running MS-GF+ through a GUI

MSGFplus comes with a sister package, *MSGFgui*, which provide a graphic interface to setting up and running MS-GF+ through *R*. Besides facilitating MS-GF+ analyses, which is arguably just as easy from the command line, it provides an intuitive way to investigate and evaluate the resulting identification data.

Figure 12 shows an example of using *MSGFgui*. It is possible to gradually drill down in the results starting from the protein level and ending at the raw spectrum level. mzIdentML files already created with MS-GF+ (using *MSGFplus* or in other ways) can easily be imported into the gui to take advantage of the visualisation features, and results can be exported as either rds (for R), xlsx (for excel) or txt (for everything else) files.

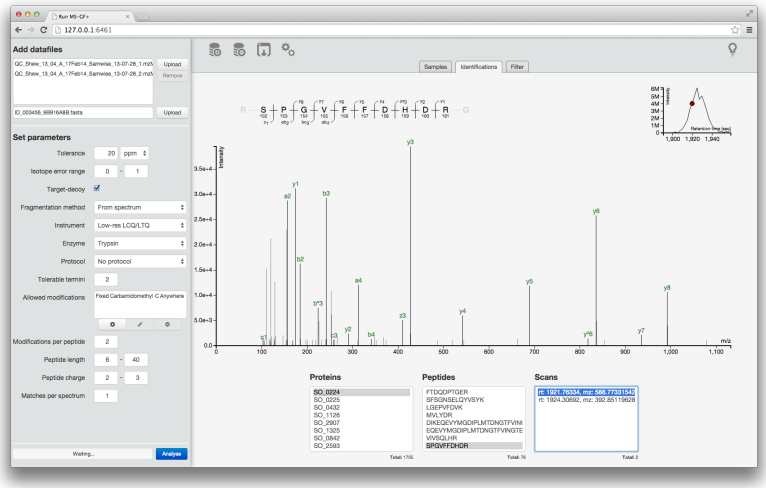


Figure 12: A screenshot of MSGFgui

5.3 Post-search Filtering of MS/MS IDs Using MSnID

The main purpose of MSnID package is to make sure that the peptide and protein identifications resulting from MS/MS searches are sufficiently confident for a given application. MS/MS peptide and protein identification is a process that prone to uncertainties. A typical and currently most reliable way to quantify uncertainty in the list of identify spectra, peptides or proteins relies on so-called decoy database. For bottom-up (i.e. involving protein digestion) approaches a common way to construct a decoy database is simple inversion of protein amino-acid sequences. If the spectrum matches to normal protein sequence it can be true or false match. Matches to decoy part of the database are false only (excluding the palindromes). Therefore the false discovery rate (FDR) of identifications can be estimated as ratio of hits to decoy over normal parts of the protein sequence database. There are multiple levels of identification that FDR can be estimated for. First, is at the level of peptide/protein- to-spectrum matches. Second is at the level of unique peptide sequences. Note, true peptides tend to be identified by more then one spectrum. False peptide tend to be sporadic. Therefore, after collapsing the redundant peptide identifications from multiple spectra to the level of unique peptide sequence, the FDR typically increases. The extend of FDR increase depends on the type and complexity of the sample. The same trend is true for estimating the identification FDR at the protein level. True proteins tend to be identified with multiple peptides, while false protein identifications are commonly covered only by one peptide. Therefore FDR estimate tend to be even higher for protein level compare to peptide level. The estimation of the FDR is also affected by the number of LC-MS (runs) datasets in the experiment. Again, true identifications tend to be more consistent from run to run, while false are sporadic. After collapsing the redundancy across the runs, the number of true identification reduces much stronger compare to false identifications. Therefore, the peptide and protein FDR estimates need to be re-evaluated. The main objective of the MSnID package is to provide convenience tools for handling tasks on estimation of FDR, defining and optimizing the filtering criteria and ensuring confidence in MS/MS identification data. The user can specify the criteria for filtering the data (e.g. goodness or p-value of matching of experimental and theoretical fragmentation mass spectrum, deviation of theoretical from experimentally measured mass, presence of missed cleavages in the peptide sequence, etc), evaluate the performance of the filter judging by FDRs at spectrum, peptide and protein levels, and finally optimize the filter to achieve the maximum number of identifications while not exceeding maximally allowed FDR upper threshold.

5.3.1 Starting Project & Importing Data

To start a project one have to specify a directory. Currently the only use of the directory is for storing cached results.

```
library("MSnID")

##
## Attaching package:  'MSnID'
```

```
##
## The following object is masked from 'package:isobar':
##
##     peptides
##
## The following object is masked from 'package:mzID':
##
##     peptides
##
## The following objects are masked from 'package:ProtGenerics':
##
##     accessions, peptides, proteins
```

```
msnid <- MSnID(".")
```

```
## Note, the anticipated/suggested columns in the
## peptide-to-spectrum matching results are:
## ------------------------------------------------
## accession
## calculatedMassToCharge
## chargeState
## experimentalMassToCharge
## isDecoy
## peptide
## spectrumFile
## spectrumID
```

Data can imported as `data.frame` or read from mzIdentML file.

```
PSMresults <- read.delim(system.file("extdata", "human_brain.txt",
                                     package="MSnID"),
                         stringsAsFactors=FALSE)
psms(msnid) <- PSMresults
show(msnid)
```

```
## MSnID object
## Working directory: "."
## #Spectrum Files:  1
## #PSMs: 997 at 37 % FDR
## #peptides: 687 at 57 % FDR
## #accessions: 665 at 65 % FDR
```

```
mzids <- system.file("extdata", "c_elegans.mzid.gz", package="MSnID")
msnid <- read_mzIDs(msnid, mzids)
```

```
## Reading from mzIdentMLs ...
```

```
## reading c_elegans.mzid.gz... DONE!
```

```
show(msnid)
```

```
## MSnID object
## Working directory: "."
## #Spectrum Files:  1
## #PSMs: 19055 at 29 % FDR
## #peptides: 9489 at 44 % FDR
## #accessions: 7414 at 76 % FDR
```

5.3.2 Analysis of Peptide Sequences

A particular properties of peptide sequences we are interested in are

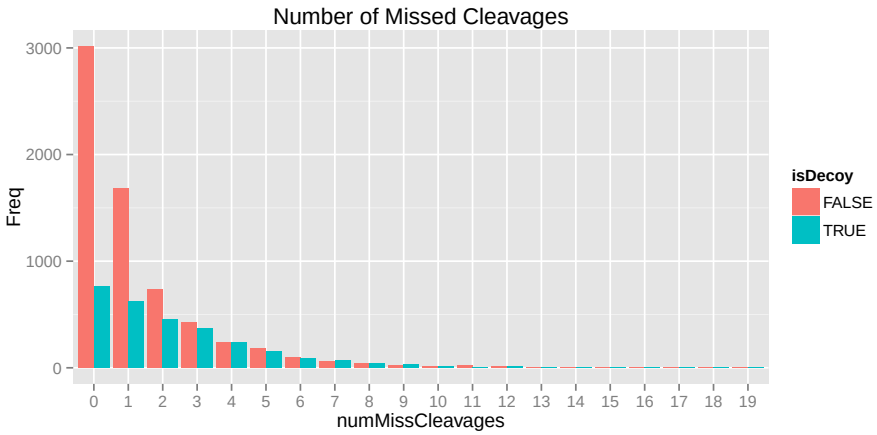
1. irregular cleavages at the termini of the peptides and
2. missing cleavage site within the peptide sequences.

The default regular expressions of valid and missed cleavage patterns correspond to trypsin. Counting the number of irregular cleavage termimi (0,1 or 2) in peptides sequence creates a new column `numIrregCleavages`. Counting the number of missed cleavages in peptides sequence creates a new column `numMissCleavages`.

```
msnid <- assess_termini(msnid, validCleavagePattern="[KR]\\.[^P]")
msnid <- assess_missed_cleavages(msnid, missedCleavagePattern="[KR](?=[^P$])")
```

Now the object has two more columns, `numIrregCleavages` and `numMissCleavages`, evidently corresponding to the number of termini with irregular cleavages and number of missed cleavages within the peptide sequence. The figure below shows that peptides with 2 or more missed cleavages are likely to be false identifications.

```
pepCleav <- unique(psms(msnid)[,c("numMissCleavages", "isDecoy", "peptide")])
pepCleav <- as.data.frame(table(pepCleav[,c("numMissCleavages", "isDecoy")]))
library("ggplot2")
ggplot(pepCleav, aes(x=numMissCleavages, y=Freq, fill=isDecoy)) +
    geom_bar(stat='identity', position='dodge') +
    ggtitle("Number of Missed Cleavages")
```



5.3.3 Defining the Filter

The criteria that will be used for filtering the MS/MS data has to be present in the `MSnID` object. We will use -log10 transformed MS-GF+ Spectrum E-value, reflecting the goodness of match experimental and theoretical fragmentation patterns as one the filtering criteria. Let's store it under the "msmsScore" name. The score density distribution shows that it is a good discriminant between non-decoy (red) and decoy hits (green).
For alternative MS/MS search engines refer to the engine-specific manual for the names of parameters reflecting the quality of MS/MS spectra matching. Examples of such parameters are `E-Value` for X!Tandem and `XCorr` and Δ`Cn2` for SEQUEST.
As a second criterion we will be using the absolute mass measurement error (in ppm units) of the parent ion. The mass measurement errors tend to be small for non-decoy (enriched with real identificaiton) hits (red line) and is effectively uniformly distributed for decoy hits.

```
msnid$msmsScore <- -log10(msnid$`MS-GF:SpecEValue`)
msnid$absParentMassErrorPPM <- abs(mass_measurement_error(msnid))
```

MS/MS fiters are handled by a special *MSnIDFilter* class objects. Individual filtering criteria can be set by name (that is present in `names(msnid)`), comparison operator (¿, ¡, =, ...) defining if we should retain hits with higher or lower given the threshold and finally the threshold value itself. The filter below set in such a way that retains only those matches that has less then 5 ppm of parent ion mass measurement error and more the 10^7 MS-GF:SpecEValue.

```
filtObj <- MSnIDFilter(msnid)
filtObj$absParentMassErrorPPM <- list(comparison="<", threshold=5.0)
filtObj$msmsScore <- list(comparison=">", threshold=8.0)
show(filtObj)
```

```
## MSnIDFilter object
## (absParentMassErrorPPM < 5) & (msmsScore > 8)
```

The stringency of the filter can be evaluated at different levels.

```
evaluate_filter(msnid, filtObj, level="PSM")
```

```
##             fdr    n
## PSM 0.002307439 9122
```

```
evaluate_filter(msnid, filtObj, level="peptide")
```

```
##                fdr    n
## peptide 0.00424371 3313
```

```
evaluate_filter(msnid, filtObj, level="accession")
```

```
##                  fdr    n
## accession 0.01770658 1207
```

5.3.4 Optimizing the Filter

The threshold values in the example above are not necessarily optimal and set just be in the range of probable values. Filters can be optimized to ensure maximum number of identifications (peptide-to-spectrum matches, unique peptide sequences or proteins) within a given FDR upper limit.
First, the filter can be optimized simply by stepping through individual parameters and their combinations. The idea has been described in [16]. The resulting `MSnIDFilter` object can be used for final data filtering or can be used as a good starting parameters for follow-up refining optimizations with more advanced algorithms.

```
filtObj.grid <- optimize_filter(filtObj, msnid, fdr.max=0.01,
                                method="Grid", level="peptide",
                                n.iter=500)
show(filtObj.grid)
```

```
## MSnIDFilter object
## (absParentMassErrorPPM < 10) & (msmsScore > 7.8)
```

The resulting `filtObj.grid` can be further fine tuned with such optimization routines as simulated annealing or Nelder-Mead optimization.

```
filtObj.nm <- optimize_filter(filtObj.grid, msnid, fdr.max=0.01,
                                method="Nelder-Mead", level="peptide",
                                n.iter=500)
show(filtObj.nm)
```

```
## MSnIDFilter object
## (absParentMassErrorPPM < 10) & (msmsScore > 7.8)
```

Evaluate non-optimized and optimized filters.

```
evaluate_filter(msnid, filtObj, level="peptide")
```

```
##                fdr    n
## peptide 0.00424371 3313
```

```
evaluate_filter(msnid, filtObj.grid, level="peptide")
```

```
##                 fdr    n
## peptide 0.009220702 3393
```

```
evaluate_filter(msnid, filtObj.nm, level="peptide")
```

```
##                 fdr    n
## peptide 0.009777778 3408
```

Finally applying filter to remove predominantly false identifications.

```
msnid <- apply_filter(msnid, filtObj.nm)
show(msnid)
```

```
## MSnID object
## Working directory: "."
## #Spectrum Files:  1
## #PSMs: 9480 at 0.49 % FDR
## #peptides: 3408 at 0.98 % FDR
## #accessions: 1253 at 3.8 % FDR
```

Removing hits to decoy and contaminant sequences using the same `apply_filter` method.

```
msnid <- apply_filter(msnid, "isDecoy == FALSE")
show(msnid)
```

```
## MSnID object
## Working directory: "."
## #Spectrum Files:  1
## #PSMs: 9434 at 0 % FDR
## #peptides: 3375 at 0 % FDR
## #accessions: 1207 at 0 % FDR
```

```
msnid <- apply_filter(msnid, "!grepl('Contaminant',accession)")
show(msnid)
```

```
## MSnID object
## Working directory: "."
## #Spectrum Files:  1
## #PSMs: 9425 at 0 % FDR
## #peptides: 3368 at 0 % FDR
## #accessions: 1205 at 0 % FDR
```

5.3.5 Interface with Other Bioconductor Packages

One can extract the entire PSMs tables as `data.frame` or `data.table`

```
psm.df <- psms(msnid)
psm.dt <- as(msnid, "data.table")
```

If only interested in the non-redundant list of confidently identified peptides or proteins

```
peps <- peptides(msnid)
head(peps)
```

```
## [1] "K.AISQIQEYVDYYGGSGVQHIALNTSDIITAIEALR.A"
## [2] "K.SAGSGYLVGDSLTFVDLLVAQHTADLLAANAALLDEFPQFK.A"
## [3] "K.NSIFTNVAETANGEYFWEGLEDEIADKNVDITTWLGEK.W"
## [4] "R.VFCLLGDGESAEGSVWEAAAFASIYKLDNLVAIVDVNR.L"
## [5] "R.TTDSDGNNTGLDLYTVDQVEHSNYVEQNFLDFIFVFR.K"
## [6] "R.KFDADGSGKLEFDEFCALVYTVANTVDKETLEKELR.E"

prots <- accessions(msnid)
head(prots)

## [1] "CE02347" "CE07055" "CE12728" "CE36358" "CE36359"
## [6] "CE36360"

prots <- proteins(msnid) # may be more intuitive then accessions
head(prots)

## [1] "CE02347" "CE07055" "CE12728" "CE36358" "CE36359"
## [6] "CE36360"
```

The *MSnID* package is aimed at providing convenience functionality to handle MS/MS identifications. Quantification *per se* is outside of the scope of the package. The only type of quantitation that can be seamlessly tied with MS/MS identification analysis is so-called *spectral counting* approach. In such an approach a peptide abundance is considered to be directly proportional to the number of matched MS/MS spectra. In its turn protein abunance is proportional to the sum of the number of spectra of the matching peptides. The *MSnID* object can be converted to an *MSnSet* object defined in *MSnbase* that extends generic Bioconductor *eSet* class to quantitative proteomics data. The spectral count data can be analyzed with *msmsEDA*, *msmsTests* or *DESeq* packages.

```
msnset <- as(msnid, "MSnSet")
library("MSnbase")
head(fData(msnset))

##                                                 peptide
## A.AGLKPTQAMVTK.A                          A.AGLKPTQAMVTK.A
## A.AVLEYLAAEVLELAGNAAR.D              A.AVLEYLAAEVLELAGNAAR.D
## A.DCLHCICMR.E                              A.DCLHCICMR.E
## A.DLFTSIADMQNLLETER.N                  A.DLFTSIADMQNLLETER.N
## A.EKKRKAAETSLMEK.D                        A.EKKRKAAETSLMEK.D
## A.EQLPEKFYGTFDLDHSENFDEYLTAK.G A.EQLPEKFYGTFDLDHSENFDEYLTAK.G
##                                                   accession
## A.AGLKPTQAMVTK.A                                CE01236, CE30652
## A.AVLEYLAAEVLELAGNAAR.D                         CE04501, CE05477
## A.DCLHCICMR.E                  CE04442, CE17549, CE24850, CE34002
## A.DLFTSIADMQNLLETER.N                             CE20261
## A.EKKRKAAETSLMEK.D                                CE27133
## A.EQLPEKFYGTFDLDHSENFDEYLTAK.G                    CE04532

head(exprs(msnset))

##                               c_elegans_A_3_1_21Apr10_Draco_10-03-04_dta.txt
## A.AGLKPTQAMVTK.A                                                           1
## A.AVLEYLAAEVLELAGNAAR.D                                                    1
## A.DCLHCICMR.E                                                              1
## A.DLFTSIADMQNLLETER.N                                                      1
## A.EKKRKAAETSLMEK.D                                                         1
## A.EQLPEKFYGTFDLDHSENFDEYLTAK.G                                             1
```

Note, the convertion from `MSnID` to `MSnSet` uses peptides as features. The number of redundant peptide observations represent so-called spectral count that can be used for rough quantitative analysis. Summing of all of the peptide counts

to a proteins level can be done with `combineFeatures` function from *MSnbase* package.

```
msnset <- combineFeatures(msnset,
                          fData(msnset)$accession,
                          redundancy.handler="unique",
                          fun="sum",
                          cv=FALSE)
```

```
## Combined 2082 features into 670 using sum
```

```
head(fData(msnset))
```

```
##                                     peptide accession
## CE00078                          K.RLPVAPR.G   CE00078
## CE00103 K.LPNDDIGVQVSYLGEPHTFTPEQVLAALLTK.L   CE00103
## CE00134                         I.PAEVAEHLK.A   CE00134
## CE00209            K.ALEGPGPGEDAAHSENNPPR.N   CE00209
## CE00302                  K.LTYFDIHGLAEPIR.L   CE00302
## CE00318        K.ALNALCAQLMTELADALEVLDTDK.S   CE00318
```

```
head(exprs(msnset))
```

```
##         c_elegans_A_3_1_21Apr10_Draco_10-03-04_dta.txt
## CE00078                                              4
## CE00103                                              3
## CE00134                                              4
## CE00209                                              8
## CE00302                                              2
## CE00318                                             11
```

6 Quality control

Quality control (QC) is an essential part of any high throughput data driven approach. Bioconductor has a rich history of QC for various genomics data and currently two packages support proteomics QC.

proteoQC provides a dedicated a dedicated pipeline that will produce a dynamic and extensive html report. It uses the *rTANDEM* package to automate the generation of identification data and uses information about the experimental/replication design.

The *qcmetrics* package is a general framework to define QC metrics and bundle them together to generate html or pdf reports. It provides some ready made metrics for MS data and ^{15}N labelled data.

7 Annotation

In this section, we briefly present some *Bioconductor* annotation infrastructure.

We start with the *hpar* package, an interface to the *Human Protein Atlas* [17, 18], to retrieve subcellular localisation information for the `ENSG00000002746` ensemble gene.

```
id <- "ENSG00000002746"
library("hpar")
getHpa(id, "SubcellularLoc")
```

```
## [1] Gene            Main.location   Other.location
## [4] Expression.type Reliability
## <0 rows> (or 0-length row.names)
```

Below, we make use of the human annotation package *org.Hs.eg.db* and the Gene Ontology annotation package *GO.db* to retrieve the same information as above.

```
library(org.Hs.eg.db)
library(GO.db)
ans <- select(org.Hs.eg.db,
              keys = id, columns = c("ENSEMBL", "GO", "ONTOLOGY"),
              keytype = "ENSEMBL")
ans <- ans[ans$ONTOLOGY == "CC", ]
ans
```

```
##           ENSEMBL         GO EVIDENCE ONTOLOGY
## 3 ENSG00000002746 GO:0005829      TAS       CC
```

```
sapply(as.list(GOTERM[ans$GO]), slot, "Term")
```

```
## GO:0005829
##  "cytosol"
```

Finally, this information can also be retrieved from on-line databases using the *biomaRt* package [19].

```
library("biomaRt")
ensembl <- useMart("ensembl",dataset="hsapiens_gene_ensembl")
efilter <- "ensembl_gene_id"
eattr <- c("go_id", "name_1006", "namespace_1003")
bmres <- getBM(attributes=eattr, filters = efilter, values = id, mart = ensembl)
bmres[bmres$namespace_1003 == "cellular_component", "name_1006"]
```

```
## [1] "cellular_component" "cytoplasm"
```

8 Other packages

8.1 Bioconductor packages

This section provides a complete list of packages available in the relevant *Bioconductor* version 3.1 (as of March 23, 2015) *biocView*[19] categories. Tables 1, 2 and 3 represent the packages for the `Proteomics` (68 packages), `MassSpectrometry` (49 packages) and `MassSpectrometryData` (11 experiment packages) categories.

Package	Title	Version
ASEB	Predict Acetylated Lysine Sites	1.11.0
bioassayR	R library for Bioactivity analysis	1.5.10
BRAIN	Baffling Recursive Algorithm for Isotope distributioN calculations	1.13.0
Cardinal	A mass spectrometry imaging toolbox for statistical analysis	0.99.5
CellNOptR	Training of boolean logic models of signalling networks using prior knowledge networks and perturbation data.	1.13.0
ChemmineR	Cheminformatics Toolkit for R	2.19.0
cisPath	Visualization and management of the protein-protein interaction networks.	1.7.4
cleaver	Cleavage of Polypeptide Sequences	1.5.3
clippda	A package for the clinical proteomic profiling data analysis	1.17.0
CNORdt	Add-on to CellNOptR: Discretized time treatments	1.9.0
CNORfeeder	Integration of CellNOptR to add missing links	1.7.0
CNORode	ODE add-on to CellNOptR	1.9.1
customProDB	Generate customized protein database from NGS data, with a focus on RNA-Seq data, for proteomics search.	1.7.0
deltaGseg	deltaGseg	1.7.0
eiR	Accelerated similarity searching of small molecules	1.7.2
fmcsR	Mismatch Tolerant Maximum Common Substructure Searching	1.9.0
GraphPAC	Identification of Mutational Clusters in Proteins via a Graph Theoretical Approach.	1.9.0
hpar	Human Protein Atlas in R	1.9.1
iPAC	Identification of Protein Amino acid Clustering	1.11.0
IPPD	Isotopic peak pattern deconvolution for Protein Mass Spectrometry by template matching	1.15.0
isobar	Analysis and quantitation of isobarically tagged MSMS proteomics data	1.13.2
LPEadj	A correction of the local pooled error (LPE) method to replace the asymptotic variance adjustment with an unbiased adjustment based on sample size.	1.27.0
MassSpecWavelet	Mass spectrum processing by wavelet-based algorithms	1.33.0
MSGFgui	A shiny GUI for MSGFplus	1.1.2
MSGFplus	An interface between R and MS-GF+	1.1.3
msmsEDA	Exploratory Data Analysis of LC-MS/MS data by spectral counts	1.5.0
msmsTests	LC-MS/MS Differential Expression Tests	1.5.0
MSnbase	Base Functions and Classes for MS-based Proteomics	1.15.11
MSnID	Utilities for Exploration and Assessment of Confidence of LC-MSn Proteomics Identifications.	1.1.3
MSstats	Protein Significance Analysis in DDA, SRM and DIA for Label-free or Label-based Proteomics Experiments	2.5.0
mzID	An mzIdentML parser for R	1.5.2
mzR	parser for netCDF, mzXML, mzData and mzML and mzIdentML files (mass spectrometry data)	2.1.15
PAA	PAA (Protein Array Analyzer)	1.1.1
PAnnBuilder	Protein annotation data package builder	1.31.1
pathview	a tool set for pathway based data integration and visualization	1.7.0
Pbase	Manipulating and exploring protein and proteomics data	0.6.12
PCpheno	Phenotypes and cellular organizational units	1.29.0
pepXMLTab	Parsing pepXML files and filter based on peptide FDR.	1.1.0
plgem	Detect differential expression in microarray and proteomics datasets with the Power Law Global Error Model (PLGEM)	1.39.1
PLPE	Local Pooled Error Test for Differential Expression with Paired High-throughput Data	1.27.0
ppiStats	Protein-Protein Interaction Statistical Package	1.33.0
proBAMr	Generating SAM file for PSMs in shotgun proteomics data.	1.1.2
PROcess	Ciphergen SELDI-TOF Processing	1.43.0
procoil	Prediction of Oligomerization of Coiled Coil Proteins	1.17.0
ProCoNA	Protein co-expression network analysis (ProCoNA).	1.5.2
pRoloc	A unifying bioinformatics framework for spatial proteomics	1.7.5
pRolocGUI	Interactive visualisation of spatial proteomics data	1.1.4
prot2D	Statistical Tools for volume data from 2D Gel Electrophoresis	1.5.0
proteoQC	An R package for proteomics data quality control	1.3.2
ProtGenerics	S4 generic functions for Bioconductor proteomics infrastructure	0.99.3
Pviz	Peptide Annotation and Data Visualization using Gviz	1.1.1
qcmetrics	A Framework for Quality Control	1.5.1
QuartPAC	Identification of mutational clusters in protein quaternary structures.	0.99.3
rain	Rhythmicity Analysis Incorporating Non-parametric Methods	1.1.1
RCASPAR	A package for survival time prediction based on a piecewise baseline hazard Cox regression model.	1.13.0
Rchemcpp	Similarity measures for chemical compounds	2.5.0
Rcpi	Toolkit for Compound-Protein Interaction in Drug Discovery	1.3.0
RpsiXML	R interface to PSI-MI 2.5 files	2.9.0

[19] http://www.bioconductor.org/packages/devel/BiocViews.html

rpx	R Interface to the ProteomeXchange Repository	1.3.0
rTANDEM	Interfaces the tandem protein identification algorithm in R	1.7.0
sapFinder	A package for variant peptides detection and visualization in shotgun proteomics.	1.5.10
ScISI	In Silico Interactome	1.39.0
shinyTANDEM	Provides a GUI for rTANDEM	1.5.0
SLGI	Synthetic Lethal Genetic Interaction	1.27.0
SpacePAC	Identification of Mutational Clusters in 3D Protein Space via Simulation.	1.5.0
specL	specL - Prepare Peptide Spectrum Matches for Use in Targeted Proteomics	1.1.14
spliceSites	Manages align gap positions from RNA-seq data	1.5.0
synapter	Label-free data analysis pipeline for optimal identification and quantitation	1.9.4

Table 1: Packages available under the `Proteomics` *biocViews* category.

Package	Title	Version
apComplex	Estimate protein complex membership using AP-MS protein data	2.33.0
BRAIN	Baffling Recursive Algorithm for Isotope distributioN calculations	1.13.0
CAMERA	Collection of annotation related methods for mass spectrometry data	1.23.2
Cardinal	A mass spectrometry imaging toolbox for statistical analysis	0.99.5
cosmiq	cosmiq - COmbining Single Masses Into Quantities	1.1.0
cytofkit	cytofkit: an integrated analysis pipeline for mass cytometry data	0.99.18
flagme	Analysis of Metabolomics GC/MS Data	1.23.1
gaga	GaGa hierarchical model for high-throughput data analysis	2.13.0
iontree	Data management and analysis of ion trees from ion-trap mass spectrometry	1.13.0
isobar	Analysis and quantitation of isobarically tagged MSMS proteomics data	1.13.2
MAIT	Statistical Analysis of Metabolomic Data	1.1.0
MassArray	Analytical Tools for MassArray Data	1.19.0
MassSpecWavelet	Mass spectrum processing by wavelet-based algorithms	1.33.0
Metab	Metab: An R Package for a High-Throughput Analysis of Metabolomics Data Generated by GC-MS.	1.1.0
metabomxtr	A package to run mixture models for truncated metabolomics data with normal or lognormal distributions.	1.1.0
metaMS	MS-based metabolomics annotation pipeline	1.3.5
MSGFgui	A shiny GUI for MSGFplus	1.1.2
MSGFplus	An interface between R and MS-GF+	1.1.3
msmsEDA	Exploratory Data Analysis of LC-MS/MS data by spectral counts	1.5.0
msmsTests	LC-MS/MS Differential Expression Tests	1.5.0
MSnbase	Base Functions and Classes for MS-based Proteomics	1.15.11
MSnID	Utilities for Exploration and Assessment of Confidence of LC-MSn Proteomics Identifications.	1.1.3
MSstats	Protein Significance Analysis in DDA, SRM and DIA for Label-free or Label-based Proteomics Experiments	2.5.0
mzID	An mzIdentML parser for R	1.5.2
mzR	parser for netCDF, mzXML, mzData and mzML and mzIdentML files (mass spectrometry data)	2.1.15
PAPi	Predict metabolic pathway activity based on metabolomics data	1.7.0
Pbase	Manipulating and exploring protein and proteomics data	0.6.12
pepXMLTab	Parsing pepXML files and filter based on peptide FDR.	1.1.0
plgem	Detect differential expression in microarray and proteomics datasets with the Power Law Global Error Model (PLGEM)	1.39.1
proBAMr	Generating SAM file for PSMs in shotgun proteomics data.	1.1.2
PROcess	Ciphergen SELDI-TOF Processing	1.43.0
pRoloc	A unifying bioinformatics framework for spatial proteomics	1.7.5
proteoQC	An R package for proteomics data quality control	1.3.2
ProtGenerics	S4 generic functions for Bioconductor proteomics infrastructure	0.99.3
qcmetrics	A Framework for Quality Control	1.5.1
Rdisop	Decomposition of Isotopic Patterns	1.27.0
Risa	Converting experimental metadata from ISA-tab into Bioconductor data structures	1.9.1
RMassBank	Workflow to process tandem MS files and build MassBank records	1.9.1
rols	An R interface to the Ontology Lookup Service	1.9.0
rpx	R Interface to the ProteomeXchange Repository	1.3.0
rTANDEM	Interfaces the tandem protein identification algorithm in R	1.7.0
sapFinder	A package for variant peptides detection and visualization in shotgun proteomics.	1.5.10
shinyTANDEM	Provides a GUI for rTANDEM	1.5.0
sidap	sidap: an integrated analysis pipeline for mass cytometry data	0.99.9
SIMAT	GC-SIM-MS data processing and alaysis tool	0.99.3
specL	specL - Prepare Peptide Spectrum Matches for Use in Targeted Proteomics	1.1.14
synapter	Label-free data analysis pipeline for optimal identification and quantitation	1.9.4
TargetSearch	A package for the analysis of GC-MS metabolite profiling data.	1.23.0
xcms	LC/MS and GC/MS Data Analysis	1.43.3

Table 2: Packages available under the `MassSpectrometry` *biocViews* category.

Package	Title	Version
CardinalWorkflows	Datasets and workflows for the Cardinal mass spectrometry imaging package	0.99.2
faahKO	Saghatelian et al. (2004) FAAH knockout LC/MS data	1.7.1
gcspikelite	Spike-in data for GC/MS data and methods within flagme	1.5.1
iontreeData	Data provided to show the usage of functions in iontree package	1.3.1
metaMSdata	Example CDF data for the metaMS package	1.3.0

msdata	Various Mass Spectrometry raw data example files	0.5.0
ProData	SELDI-TOF data of Breast cancer samples	1.5.1
pRolocdata	Data accompanying the pRoloc package	1.5.5
RforProteomics	Companion package to the 'Using R and Bioconductor for proteomics data analysis' publication	1.5.8
RMassBankData	Test dataset for RMassBank	1.5.0
synapterdata	Data accompanying the synapter package	1.5.1

Table 3: Experimental Packages available under the `MassSpectrometryData` *biocViews* category.

The tables can easily be generated with the `proteomicsPackages`, `massSpectrometryPackages` and `massSpectrometryDataPackag` functions. The respective package tables can then be interactively explored using the `display` function.

```
pp <- proteomicsPackages()
display(pp)
```

8.2 Other CRAN packages

The CRAN task view on Chemometrics and Computational Physics[20] is another useful ressource listing 81 packages, including a set of packages for mass spectrometry and proteomics, some of which are illustrated in this document.

MALDIquant provides tools for quantitative analysis of MALDI-TOF mass spectrometry data, with support for baseline correction, peak detection and plotting of mass spectra (http://cran.r-project.org/web/packages/MALDIquant/index.html).

OrgMassSpecR is for organic/biological mass spectrometry, with a focus on graphical display, quantification using stable isotope dilution, and protein hydrogen/deuterium exchange experiments (http://cran.r-project.org/web/packages/OrgMassSpecR/index.html).

FTICRMS provides functions for Analyzing Fourier Transform-Ion Cyclotron Resonance Mass Spectrometry Data (http://cran.r-project.org/web/packages/FTICRMS/index.html).

titan provides a GUI to analyze mass spectrometric data on the relative abundance of two substances from a titration series (http://cran.r-project.org/web/packages/titan/index.html).

digeR provides a GUI interface for analysing 2D DIGE data. It allows to perform correlation analysis, score plot, classification, feature selection and power analysis for 2D DIGE experiment data. (http://cran.r-project.org/web/packages/digeR/index.html)

protViz helps with quality checks, visualizations and analysis of mass spectrometry data, coming from proteomics experiments. The package is developed, tested and used at the Functional Genomics Center Zurich. (http://cran.r-project.org/web/packages/protViz/index.html)

Suggestions for additional *R* packages are welcome and will be added to the vignette. Please send suggestions and possibly a short description and/or a example utilisation with code to lg390@cam.ac.uk. The only requirement is that the package must be available on an official package channel (CRAN, *Bioconductor*, R-forge, Omegahat), i.e. not only available through a personal web page.

[20] http://cran.r-project.org/web/views/ChemPhys.html

9 Session information

All software and respective versions used in this document, as returned by `sessionInfo()` are detailed below.

- R Under development (unstable) (2015-01-22 r67580), `x86_64-unknown-linux-gnu`
- Base packages: base, datasets, graphics, grDevices, methods, parallel, stats, stats4, utils
- Other packages: AnnotationDbi 1.29.17, Biobase 2.27.2, BiocGenerics 0.13.7, BiocInstaller 1.17.6, BiocParallel 1.1.18, Biostrings 2.35.11, bitops 1.0-6, BRAIN 1.13.0, cleaver 1.5.3, data.table 1.9.4, DBI 0.3.1, digest 0.6.8, GenomeInfoDb 1.3.14, ggplot2 1.0.1, GO.db 3.1.0, hpar 1.9.1, IPPD 1.15.0, IRanges 2.1.43, isobar 1.13.2, knitr 1.9, lattice 0.20-30, MALDIquant 1.11, MALDIquantForeign 0.9, MASS 7.3-39, Matrix 1.1-5, msdata 0.5.0, MSGFgui 1.1.2, MSGFplus 1.1.3, MSnbase 1.15.13, MSnID 1.1.3, mzID 1.5.2, mzR 2.1.14, org.Hs.eg.db 3.1.0, OrgMassSpecR 0.4-4, PolynomF 0.94, ProtGenerics 0.99.3, RColorBrewer 1.1-2, Rcpp 0.11.5, RcppClassic 0.9.6, Rdisop 1.27.0, reshape2 1.4.1, RforProteomics 1.5.8, rJava 0.9-6, rols 1.9.0, rpx 1.3.0, RSQLite 1.0.0, rTANDEM 1.7.0, S4Vectors 0.5.22, xlsx 0.5.7, xlsxjars 0.6.1, XML 3.98-1.1, xtable 1.7-4, XVector 0.7.4
- Loaded via a namespace (and not attached): affy 1.45.2, affyio 1.35.0, annotate 1.45.2, base64enc 0.1-2, BiocStyle 1.5.3, biocViews 1.35.17, Category 2.33.0, chron 2.3-45, codetools 0.2-11, colorspace 1.2-6, compiler 3.2.0, distr 2.5.3, doParallel 1.0.8, downloader 0.3, evaluate 0.5.5, foreach 1.4.2, formatR 1.0, futile.logger 1.3.7, futile.options 1.0.0, genefilter 1.49.2, graph 1.45.2, grid 3.2.0, gridSVG 1.4-3, GSEABase 1.29.1, gtable 0.1.2, highr 0.4, htmltools 0.2.6, httpuv 1.3.2, impute 1.41.0, interactiveDisplay 1.5.1, interactiveDisplayBase 1.5.1, iterators 1.0.7, labeling 0.3, lambda.r 1.1.6, limma 3.23.11, mime 0.2, munsell 0.4.2, pcaMethods 1.57.2, plyr 1.8.1, preprocessCore 1.29.0, proto 0.3-10, R6 2.0.1, RBGL 1.43.0, R.cache 0.10.0, RCurl 1.95-4.5, readBrukerFlexData 1.8.2, readMzXmlData 2.8, RJSONIO 1.3-0, R.methodsS3 1.7.0, R.oo 1.19.0, RUnit 0.4.28, R.utils 2.0.0, scales 0.2.4, sfsmisc 1.0-27, shiny 0.11.1, shinyFiles 0.5.0, splines 3.2.0, SSOAP 0.8-0, startupmsg 0.9, stringr 0.6.2, survival 2.38-1, SweaveListingUtils 0.6.2, tools 3.2.0, vsn 3.35.0, XMLSchema 0.7-2, zlibbioc 1.13.2

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